

Package ‘GLAD’

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Title Gain and Loss Analysis of DNA

Depends R (>= 2.10)

SystemRequirements gsl. Note: users should have GSL installed. Windows users: 'consult the README file available in the inst directory of the source distribution for necessary configuration instructions'.

Imports aws

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Description Analysis of array CGH data : detection of breakpoints in genomic profiles and assignment of a status (gain, normal or loss) to each chromosomal regions identified.

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URL <http://bioinfo.curie.fr>

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array	<i>Bladder cancer CGH data</i>
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Description

Bladder cancer data from 3 arrays CGH (Comparative Genomic Hybridization). Arrays dimension are 4 blocs per column, 4 blocs per row, 21 columns per bloc and 22 rows by blocs.

Usage

```
data(arrayCGH)
```

Format

A data frame composed of the following elements :

Log2Rat Log 2 ratio.

Position BAC position on the genome.

CHROMOSOME Chromosome.

Col Column location on the array.

Row Row location on the array.

Source

Institut Curie, <glad@curie.fr>.

Examples

```
data(arrayCGH)
data <- array1 #array1 to array3
```

arrayCGH	<i>Object of Class arrayCGH</i>
----------	---------------------------------

Description

Description of the object arrayCGH.

Value

The object arrayCGH is a list with at least a data.frame named arrayValues and a vector named arrayDesign. The data.frame arrayValues must contain the following fields :

Col	Vector of columns coordinates.
Row	Vector of rows coordinates.
...	Other elements can be added.

The vector arrayDesign is composed of 4 values : c(arrayCol, arrayRow, SpotCol, SpotRow). The array CGH is represented by arrayRow*arrayCol blocs and each bloc is composed of SpotRow*SpotCol spots.

N.B. : Col takes the values in 1:arrayRow*SpotRow and Row takes the values in 1:arrayCol*SpotCol

Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

Author(s)

Philippe HupÃ©, <glad@curie.fr>.

See Also

[glad](#).

Examples

```
data(arrayCGH)

# object of class arrayCGH

array <- list(arrayValues=array2, arrayDesign=c(4,4,21,22))
class(array) <- "arrayCGH"
```

arrayPersp

*Perspective image of microarray spots statistic***Description**

The function `arrayPersp` creates perspective images of shades of gray or colors that correspond to the values of a statistic for each spot on the array. The statistic can be the intensity log-ratio, a spot quality measure (e.g. spot size or shape), or a test statistic. This function can be used to explore whether there are any spatial effects in the data, for example, print-tip or cover-slip effects.

Usage

```
## Default S3 method:
arrayPersp(Statistic, Col, Row,
           ArrCol, ArrRow, SpotCol, SpotRow,
           mediancenter=FALSE,
           col=myPalette("green", "red", "yellow"),
           zlim=zlim, bar=TRUE, ...)
```

```
## S3 method for class 'arrayCGH'
arrayPersp(arrayCGH, variable,
           mediancenter=FALSE,
           col=myPalette("green", "red", "yellow"),
           zlim=zlim, bar=TRUE, ...)
```

Arguments

<code>arrayCGH</code>	Object of class <code>arrayCGH</code> .
<code>variable</code>	Variable to be plotted
<code>Statistic</code>	Statistic to be plotted.
<code>Col</code>	Vector of columns coordinates.
<code>Row</code>	Vector of rows coordinates.
<code>ArrCol</code>	Number of columns for the blocs.
<code>ArrRow</code>	Number of rows for the blocs.
<code>SpotCol</code>	Number of column for each bloc.
<code>SpotRow</code>	Number of rows for each bloc.
<code>mediancenter</code>	If <code>mediancenter=TRUE</code> , values of <code>Statistic</code> are median-centered.
<code>col</code>	List of colors such as that generated by Palettes. In addition to these color palettes functions, a new function <code>myPalette</code> was defined to generate color palettes from user supplied low, middle, and high color values.

`zlim` Numerical vector of length 2 giving the extreme values of `z` to associate with colors low and high of `myPalette`. By default `zlim` is the range of `z`. Any values of `z` outside the interval `zlim` will be truncated to the relevant limit.

`bar` If `bar=TRUE`, a calibration color bar is shown to the right of the image.

... Graphical parameters can be given as arguments to function `persp`.

N.B. : `Col` takes the values in `1:arrayRow*SpotRow` and `Row` takes the values in `1:arrayCol*SpotCol`

Value

An image is created on the current graphics device.

Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

Author(s)

Philippe HupÃ©, <glad@curie.fr>.

See Also

`persp`, `arrayPlot`, `myPalette`.

Examples

```
## Not run:
data(arrayCGH)

# object of class arrayCGH

array <- list(arrayValues=array2, arrayDesign=c(4,4,21,22))
class(array) <- "arrayCGH"

arrayPersp(array,"Log2Rat", main="Perspective image of array CGH",
           box=FALSE, theta=110, phi=40)

## End(Not run)
```

arrayPlot

*Spatial image of microarray spots statistic***Description**

The function `arrayPlot` creates spatial images of shades of gray or colors that correspond to the values of a statistic for each spot on the array. The statistic can be the intensity log-ratio, a spot quality measure (e.g. spot size or shape), or a test statistic. This function can be used to explore whether there are any spatial effects in the data, for example, print-tip or cover-slip effects.

Usage

```
## Default S3 method:
arrayPlot(Statistic, Col, Row,
          ArrCol, ArrRow, SpotCol, SpotRow,
          mediancenter=FALSE,
          col=myPalette("green", "red", "yellow"),
          contour=FALSE, nlevels=5,
          zlim=NULL, bar=c("none", "horizontal", "vertical"),
          layout=TRUE, ...)

## S3 method for class 'arrayCGH'
arrayPlot(arrayCGH, variable,
          mediancenter=FALSE,
          col=myPalette("green", "red", "yellow"),
          contour=FALSE, nlevels=5,
          zlim=NULL, bar=c("none", "horizontal", "vertical"),
          layout=TRUE, ...)
```

Arguments

<code>arrayCGH</code>	Object of class <code>arrayCGH</code> .
<code>variable</code>	Variable to be plotted
<code>Statistic</code>	Statistic to be plotted.
<code>Col</code>	Vector of columns coordinates.
<code>Row</code>	Vector of rows coordinates.
<code>ArrCol</code>	Number of columns for the blocs.
<code>ArrRow</code>	Number of rows for the blocs.
<code>SpotCol</code>	Number of column for each bloc.
<code>SpotRow</code>	Number of rows for each bloc.
<code>mediancenter</code>	If <code>mediancenter=TRUE</code> , values of <code>Statistic</code> are median-centered.
<code>col</code>	List of colors such as that generated by <code>Palettes</code> . In addition to these color palettes functions, a new function <code>myPalette</code> was defined to generate color palettes from user supplied low, middle, and high color values.

contour	If contour=TRUE, contour are plotted, otherwise they are not shown.
nlevels	Numbers of levels added by contour if contour=TRUE.
zlim	Numerical vector of length 2 giving the extreme values of z to associate with colors low and high of myPalette . By default zlim is the range of z. Any values of z outside the interval zlim will be truncated to the relevant limit.
bar	If bar=='horizontal' (resp. 'vertical'), an horizontal (resp. vertical) calibration color bar is shown to the right of the image.
layout	If layout==TRUE plot layout is automatically set when a color bar is asked for
...	Graphical parameters can be given as arguments to function image.

N.B. : Col takes the values in 1:arrayRow*SpotRow and Row takes the values in 1:arrayCol*SpotCol

Details

This function is very similar to the `maImage` written by Sandrine Dudoit (available in `marrayPlots` package) with added options `zlim`, `mediacenter` and `layout`.

Value

An image is created on the current graphics device.

Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

Author(s)

Philippe HupÃ©, <glad@curie.fr>.

See Also

`image`, `contour`, [arrayPersp](#), [myPalette](#).

Examples

```
data(arrayCGH)

pdf(file="arrayCGH.pdf",height=21/cm(1),width=29.7/cm(1))
arrayPlot(array2$Log2Rat, array2$Col, array2$Row, 4,4,21,22, main="Spatial Image of array CGH")
dev.off()

# object of class arrayCGH

array <- list(arrayValues=array2, arrayDesign=c(4,4,21,22))
class(array) <- "arrayCGH"

arrayPlot(array,"Log2Rat", main="Spatial Image of array CGH")
```

as.data.frame.profileCGH

profileCGH consercion

Description

Convert a profileCGH object into a data.frame.

Usage

```
## S3 method for class 'profileCGH'  
as.data.frame(x, row.names = NULL, optional = FALSE, ...)
```

Arguments

x	The object to converted into data.frame.
row.names	NULL or a character vector giving the row names for the data frame. Missing values are not allowed.
optional	logical. If 'TRUE', setting row names and converting column names (to syntactic names) is optional.
...	...

Details

The attributes profileValues and profileValuesNA are binded into a data.frame.

Value

A data.frame object

Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

Author(s)

Philippe HupÃ©, <glad@curie.fr>

See Also

[as.profileCGH](#)

Examples

```

data(snijders)

### Creation of "profileCGH" object
profileCGH <- as.profileCGH(gm13330)

#####
###
### glad function as described in HupÃ© et al. (2004)
###
#####

res <- glad(profileCGH, mediancenter=FALSE,
            smoothfunc="lawsglad", bandwidth=10, round=2,
            model="Gaussian", lkern="Exponential", qlambda=0.999,
            base=FALSE,
            lambdabreak=8, lambdacluster=8, lambdaclusterGen=40,
            type="tricubic", param=c(d=6),
            alpha=0.001, msize=5,
            method="centroid", nmax=8,
            verbose=FALSE)

res <- as.data.frame(res)

```

as.profileCGH	<i>Create an object of class profileCGH</i>
---------------	---

Description

Create an object of class profileCGH.

Usage

```

as.profileCGH(object,...)
## S3 method for class 'data.frame'
as.profileCGH(object, infaction=c("value","empty"),
value=20, keepSmoothing=FALSE, ...)

```

Arguments

object	A data.frame to be convert into profileCGH.
infaction	If "value" then the LogRatio with infinite values (-Inf, Inf) are replace by + or - value according to the sign. If "empty" then NAs are put instead.

value	replace Inf by value if infaction is "value".
keepSmoothing	if TRUE the smoothing value in object is kept
...	...

Details

The data.frame to be convert must at least contain the following fields: LogRatio, PosOrder, and Chromosome. If the field Chromosome is of mode character, it is automatically converted into a numeric vector (see [ChrNumeric](#)); a field ChromosomeChar contains the character labels. The data.frame to be converted into a profileCGH objet is split into two data.frame: profileValuesNA contains the rows for which there is at least a missing value for either LogRatio, PosOrder or Chromosome; profileValues contains the remaining rows.

Value

A list with the following attributes

profileValues	A data.frame
profileValuesNA	A data.frame

Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

Author(s)

Philippe HupÃ©, <glad@curie.fr>

See Also

[as.data.frame.profileCGH](#)

Examples

```
data(snijders)

### Creation of "profileCGH" object
profileCGH <- as.profileCGH(gm13330)

attributes(profileCGH)
```

ChrNumeric	<i>Convert chromosome into numeric values</i>
------------	---

Description

Convert chromosome into numeric values.

Usage

```
ChrNumeric(Chromosome)
```

Arguments

Chromosome A vector with chromosome labels.

Details

For sexual chromosome, labels must contains X or Y which are coded by 23 and 24 respectively.

Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

Author(s)

Philippe HupÃ©, <glad@curie.fr>

Examples

```
Chromosome <- c("1","X","Y","chr X", "ChrX", "chrX", "Chr Y")
ChrNumeric(Chromosome)
```

ColorBar	<i>Calibration bar for color images</i>
----------	---

Description

This function produces a color image (color bar) which can be used for the legend to another color image obtained from the functions [image](#) or [arrayPlot](#).

Usage

```
ColorBar(x, horizontal=TRUE, col=heat.colors(50), scale=1:length(x), k=10, ...)
```

Arguments

x	If "numeric", a vector containing the "z" values in the color image, i.e., the values which are represented in the color image. Otherwise, a "character" vector representing colors.
horizontal	If TRUE, the values of x are represented as vertical color strips in the image, else, the values are represented as horizontal color strips.
col	Vector of colors such as that generated by rainbow, heat.colors, topo.colors, terrain.colors, or similar functions. In addition to these color palette functions, a new function <code>myPalette</code> was defined to generate color palettes from user supplied low, middle, and high color values.
scale	A "numeric" vector specifying the "z" values in the color image. This is used when the argument x is a "character" vector representing color information.
k	Object of class "numeric", for the number of labels displayed on the bar.
...	Optional graphical parameters, see <code>par</code> .

Author(s)

Sandrine Dudoit, Yee Hwa (Jean) Yang.

See Also

[image](#), [arrayPlot](#) [myPalette](#).

Examples

```
par(mfrow=c(3,1))
Rcol <- myPalette(low="white", high="red", k=10)
Gcol <- myPalette(low="white", high="green", k=50)
RGcol <- myPalette(low="green", high="red", k=100)
ColorBar(Rcol)
ColorBar(Gcol, scale=c(-5,5))
ColorBar(1:50, col=RGcol)

par(mfrow=c(1,3))
x<-seq(-1, 1, by=0.01)
ColorBar(x, col=Gcol, horizontal=FALSE, k=11)
ColorBar(x, col=Gcol, horizontal=FALSE, k=21)
ColorBar(x, col=Gcol, horizontal=FALSE, k=51)
```

cytoband

Cytogenetic banding

Description

Cytogenetic banding

Usage

```
data(cytoband)
```

Examples

```
data(cytoband)
cytoband
```

 daglad

Analysis of array CGH data

Description

This function allows the detection of breakpoints in genomic profiles obtained by array CGH technology and affects a status (gain, normal or lost) to each clone.

Usage

```
## S3 method for class 'profileCGH'
daglad(profileCGH, mediancenter=FALSE,
normalrefcenter=FALSE, genomestep=FALSE,
OnlySmoothing = FALSE, OnlyOptimCall = FALSE,
smoothfunc="lawsglad", lkern="Exponential",
model="Gaussian", qlambda=0.999, bandwidth=10,
sigma=NULL, base=FALSE, round=2,
lambdabreak=8, lambdaclusterGen=40, param=c(d=6),
alpha=0.001, msize=2, method="centroid", nmin=1, nmax=8, region.size=2,
amplicon=1, deletion=-5, deltaN=0.10, forceGL=c(-0.15,0.15),
nbsigma=3, MinBkpWeight=0.35, DelBkpInAmp=TRUE, DelBkpInDel=TRUE,
CheckBkpPos=TRUE, assignGNLOut=TRUE,
breaksFdrQ = 0.0001, haarStartLevel = 1,
haarEndLevel = 5, weights.name = NULL,
verbose=FALSE, ...)
```

Arguments

profileCGH	Object of class profileCGH
mediancenter	If TRUE, LogRatio are center on their median.
genomestep	If TRUE, a smoothing step over the whole genome is performed and a "clustering throughout the genome" allows to identify a cluster corresponding to the Normal DNA level. The threshold used in the daglad function (deltaN, forceGL, amplicon, deletion) and then compared to the median of this cluster.
normalrefcenter	If TRUE, the LogRatio are centered through the median of the cluster identified during the genomestep.

OnlySmoothing	If TRUE, only segmentation is performed without optimization of breakpoints and calling.
OnlyOptimCall	If TRUE, the user can provide data which have been already segmented. In this case, <code>profileCGH\$profileValues</code> must contain a field with the name "Smoothing". The <code>daglad</code> function skip the smoothing step but both the optimization of breakpoints and calling are performed.
smoothfunc	Type of algorithm used to smooth <code>LogRatio</code> by a piecewise constant function. Choose either <code>lawsglad</code> , <code>haarseg</code> , <code>aws</code> or <code>laws</code> (<code>aws</code> package).
lkern	<code>lkern</code> determines the location kernel to be used (see <code>laws</code> in <code>aws</code> package for details).
model	<code>model</code> determines the distribution type of <code>LogRatio</code> (see <code>laws</code> in <code>aws</code> package for details).
qlambda	<code>qlambda</code> determines the scale parameter <code>qlambda</code> for the stochastic penalty (see <code>laws</code> in <code>aws</code> package for details).
base	If TRUE, the position of clone is the physical position onto the chromosome, otherwise the rank position is used.
sigma	Value to be passed to either argument <code>sigma2</code> of <code>aws</code> (see <code>aws</code> package) function or shape of <code>laws</code> (see <code>aws</code> package). If NULL, <code>sigma</code> is calculated from the data.
bandwidth	Set the maximal bandwidth <code>hmax</code> in the <code>aws</code> or <code>laws</code> functions in <code>aws</code> package. For example, if <code>bandwidth=10</code> then the <code>hmax</code> value is set to $10 * X_N$ where X_N is the position of the last clone.
round	The smoothing results of either <code>aws</code> or <code>laws</code> functions (in <code>aws</code> package) are rounded or not depending on the <code>round</code> argument. The <code>round</code> value is passed to the argument <code>digits</code> of the <code>round</code> function.
lambdabreak	Penalty term (λ') used during the "Optimization of the number of breakpoints" step.
lambdaclusterGen	Penalty term (λ^*) used during the "clustering throughout the genome" step.
param	Parameter of kernel used in the penalty term.
alpha	Risk <code>alpha</code> used for the "Outlier detection" step.
msize	The outliers MAD are calculated on regions with a cardinality greater or equal to <code>msize</code> .
method	The agglomeration method to be used during the "clustering throughout the genome" steps.
nmin	Minimum number of clusters (N^*max) allowed during the "clustering throughout the genome" clustering step.
nmax	Maximum number of clusters (N^*max) allowed during the "clustering throughout the genome" clustering step.
region.size	The breakpoints which define regions with a number of probes lower or equal to this value are discarded.
amplicon	Level (and outliers) with a smoothing value (log-ratio value) greater than this threshold are consider as amplicon. Note that first, the data are centered on the normal reference value computed during the "clustering throughout the genome" step.

deletion	Level (and outliers) with a smoothing value (log-ratio value) lower than this threshold are consider as deletion. Note that first, the data are centered on the normal reference value computed during the "clustering throughout the genome" step.
deltaN	Region with smoothing values in between the interval $[-\text{deltaN}, +\text{deltaN}]$ are supposed to be normal.
forceGL	Level with smoothing value greater (lower) than $\text{rangeGL}[1]$ ($\text{rangeGL}[2]$) are considered as gain (lost). Note that first, the data are centered on the normal reference value computed during the "clustering throughout the genome" step.
nbsigma	For each breakpoints, a weight is calculated which is a function of absolute value of the Gap between the smoothing values of the two consecutive regions. $\text{Weight} = 1 - \text{kernelpen}(\text{abs}(\text{Gap}), \text{param} = c(d = \text{nbsigma} * \text{Sigma}))$ where Sigma is the standard deviation of the LogRatio.
MinBkpWeight	Breakpoints which $\text{GNLchange} == 0$ and Weight less than MinBkpWeight are discarded.
DelBkpInAmp	If TRUE, the breakpoints identified inside amplicon regions are deleted. For amplicon, the log-ratio values are highly variable which lead to identification of false positive breakpoints.
DelBkpInDel	If TRUE, the breakpoints identified inside deletion regions are deleted. For deletion, the log-ratio values are highly variable which lead to identification of false positive breakpoints.
CheckBkpPos	If TRUE, the accuracy position of each breakpoints is checked.
assignGNLOut	If FALSE the status (gain/normal/loss) is not assigned for outliers.
breaksFdrQ	breaksFdrQ for HaarSeg algorithm.
haarStartLevel	haarStartLevel for HaarSeg algorithm.
haarEndLevel	haarEndLevel for HaarSeg algorithm.
weights.name	The name of the fields which contains the weights used for the haarseg algorithm. By default, the value is set to NULL meaning that all the observations have the same weights. If provided, the field must contain positive values.
verbose	If TRUE some information are printed.
...	...

Details

The function `daglad` implements a slightly modified version of the methodology described in the article : Analysis of array CGH data: from signal ratio to gain and loss of DNA regions (HupÃ© et al., Bioinformatics, 2004). For smoothing, it is possible to use either the AWS algorithm (Polzehl and Spokoiny, 2002) or the HaarSeg algorithm (Ben-Yaacov and Eldar, Bioinformatics, 2008). The `daglad` function allows to choose some threshold to help the algorithm to identify the status of the genomic regions. The thresholdls are given in the following parameters:

- `deltaN`
- `forceGL`
- `deletion`
- `amplicon`

Value

	An object of class "profileCGH" with the following attributes:
profileValues	is a data.frame with the following information: <ul style="list-style-type: none"> • SmoothingThe smoothing values correspond to the median of each Level • BreakpointsThe last position of a region with identical amount of DNA is flagged by 1 otherwise it is 0. Note that during the "Optimization of the number of breakpoints" step, removed breakpoints are flagged by -1. • LevelEach position with equal smoothing value are labelled the same way with an integer value starting from one. The label is incremented by one when a new level occurs or when moving to the next chromosome. • OutliersAwsEach AWS outliers are flagged -1 (if it is in the $\alpha/2$ lower tail of the distribution) or 1 (if it is in the $\alpha/2$ upper tail of the distribution) otherwise it is 0. • OutliersMadEach MAD outliers are flagged -1 (if it is in the $\alpha/2$ lower tail of the distribution) or 1 (if it is in the $\alpha/2$ upper tail of the distribution) otherwise it is 0. • OutliersTotOutliersAws + OutliersMad. • NormalRefClusters which have been used to set the normal reference during the "clustering throughout the genome" step are code by 0. Note that if genomestep=FALSE, all the value are set to 0. • ZoneGNLStatus of each clone: Gain is coded by 1, Loss by -1, Amplicon by 2, deletion by -10 and Normal by 0.
BkpInfo	is a data.frame sum up the information for each breakpoint: <ul style="list-style-type: none"> • ChromosomeChromosome name. • SmoothingSmoothing value for the breakpoint. • Gapabsolute value of the gap between the smoothing values of the two consecutive regions. • SigmaThe estimation of the standard-deviation of the chromosome. • Weight$1 - \text{kernelpen}(\text{Gap}, \text{type}, \text{param} = c(d = n \cdot \text{sigma} \cdot \text{Sigma}))$ • ZoneGNLStatus of the level where is the breakpoint. • GNLchangeTakes the value 1 if the ZoneGNL of the two consecutive regions are different. • LogRatioTest over Reference log-ratio.
NormalRef	If genomestep=TRUE and normalrefcenter=FALSE, then NormalRef is the median of the cluster which has been used to set the normal reference during the "clustering throughout the genome" step. Otherwise NormalRef is 0.

Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

Author(s)

Philippe HupÃ©, <glad@curie.fr>.

References

- HupÃ© et al. (Bioinformatics, 2004): Analysis of array CGH data: from signal ratio to gain and loss of DNA regions.
- Polzehl and Spokoiny (WIAS-Preprint 787, 2002): Local likelihood modelling by adaptive weights smoothing.
- Ben-Yaacov and Eldar (Bioinformatics, 2008): A fast and flexible method for the segmentation of aCGH data.

See Also

[glad](#).

Examples

```
data(snijders)
gm13330$Clone <- gm13330$BAC
profileCGH <- as.profileCGH(gm13330)

#####
###
### daglad function
###
#####

res <- daglad(profileCGH, mediancenter=FALSE, normalrefcenter=FALSE, genomestep=FALSE,
  smoothfunc="lawsglad", lkern="Exponential", model="Gaussian",
  qlambda=0.999, bandwidth=10, base=FALSE, round=1.5,
  lambdabreak=8, lambdaclusterGen=40, param=c(d=6), alpha=0.001, msize=2,
  method="centroid", nmin=1, nmax=8,
  amplicon=1, deletion=-5, deltaN=0.10, forceGL=c(-0.15,0.15), nbsigma=3,
  MinBkpWeight=0.35, CheckBkpPos=TRUE)

### data for cytoband
data(cytoband)

### Genomic profile on the whole genome
plotProfile(res, unit=3, Bkp=TRUE, labels=FALSE, Smoothing="Smoothing",
  main="Breakpoints detection: DAGLAD analysis", cytoband = cytoband)

###Genomic profile for chromosome 1
plotProfile(res, unit=3, Bkp=TRUE, labels=TRUE, Chromosome=1,
  Smoothing="Smoothing", main="Chromosome 1: DAGLAD analysis", cytoband = cytoband)

### The standard-deviation of LogRatio are:
res$SigmaC
```

```
### The list of breakpoints is:
res$BkpInfo
```

glad

Analysis of array CGH data

Description

This function allows the detection of breakpoints in genomic profiles obtained by array CGH technology and affects a status (gain, normal or lost) to each clone.

Usage

```
## S3 method for class 'profileCGH'
glad(profileCGH, mediancenter=FALSE,
      smoothfunc="lawsglad", bandwidth=10, round=1.5,
      model="Gaussian", lkern="Exponential", qlambda=0.999,
      base=FALSE, sigma,
      lambdabreak=8, lambdacluster=8, lambdaclusterGen=40,
      type="tricubic", param=c(d=6),
      alpha=0.001, msize=5,
      method="centroid", nmax=8, assignGNLOut=TRUE,
      breaksFdrQ = 0.0001, haarStartLevel = 1, haarEndLevel = 5,
      verbose=FALSE, ...)
```

Arguments

profileCGH	Object of class profileCGH
mediancenter	If TRUE, LogRatio are centered on their median.
smoothfunc	Type of algorithm used to smooth LogRatio by a piecewise constant function. Choose either lawsglad, haarseg, aws or laws in aws package.
bandwidth	Set the maximal bandwidth hmax in the aws or laws functions in aws package. For example, if bandwidth=10 then the hmax value is set to $10 * X_N$ where X_N is the position of the last clone.
round	The smoothing results are rounded or not depending on the round argument. The round value is passed to the argument digits of the round function.
model	Determines the distribution type of the LogRatio. Keep always the model as "Gaussian" (see laws in aws package).
lkern	Determines the location kernel to be used (see aws or laws in aws package).
qlambda	Determines the scale parameter for the stochastic penalty (see aws or laws in aws package)

base	If TRUE, the position of clone is the physical position on the chromosome, otherwise the rank position is used.
sigma	Value to be passed to either argument sigma2 of aws function or shape of laws (see aws package). If NULL, sigma is calculated from the data.
lambdabreak	Penalty term (λ') used during the Optimization of the number of breakpoints step.
lambdacluster	Penalty term (λ^*) used during the MSHR clustering by chromosome step.
lambdaclusterGen	Penalty term (λ^*) used during the HCSR clustering throughout the genome step.
type	Type of kernel function used in the penalty term during the Optimization of the number of breakpoints step, the MSHR clustering by chromosome step and the HCSR clustering throughout the genome step.
param	Parameter of kernel used in the penalty term.
alpha	Risk alpha used for the Outlier detection step.
msize	The outliers MAD are calculated on regions with a cardinality greater or equal to msize.
method	The agglomeration method to be used during the MSHR clustering by chromosome and the HCSR clustering throughout the genome clustering steps.
nmax	Maximum number of clusters (N^*max) allowed during the the MSHR clustering by chromosome and the HCSR clustering throughout the genome clustering steps.
assignGNLOut	If FALSE the status (gain/normal/loss) is not assigned for outliers.
breaksFdrQ	breaksFdrQ for HaarSeg algorithm.
haarStartLevel	haarStartLevel for HaarSeg algorithm.
haarEndLevel	for HaarSeg algorithm.
verbose	If TRUE some information are printed
...	...

Details

The function glad implements the methodology which is described in the article: Analysis of array CGH data: from signal ratio to gain and loss of DNA regions (HupÅ© et al., Bioinformatics, 2004).

The principles of the GLAD algorithm: First, the detection of breakpoints is based on the estimation of a piecewise constant function with the Adaptive Weights Smoothing (AWS) procedure (Polzehl and Spokoiny, 2002). Alternatively, it is possible to use the HaarSeg algorithm (Ben-Yaacov and Eldar, Bioinformatics, 2008). Then, a procedure based on penalized maximum likelihood optimizes the number of breakpoints and removes the undesirable breakpoints. Finally, based on the regions previously identified, a two-step unsupervised classification (**MSHR clustering by chromosome** and the **HCSR clustering throughout the genome**) with model selection criteria allows a status to be assigned for each region (gain, loss or normal).

Main parameters to be tuned:

qlambda	if you want the smoothing to fit some very local effect, choose a smaller qlambda.
bandwidth	choose a bandwidth not too small otherwise you will have a lot of little discontinuities.
lambdabreak	The higher the parameter is, the higher the number of undesirable breakpoints is.
lambdacluster	The higher the parameter is, the higher is the number of the regions within a chromosome which belong to the same cluster.
lambdaclusterGen	More the parameter is high more the regions over the whole genome are supposed to belong to the same cluster.

Value

profileValues:	<p>An object of class "profileCGH" with the following attributes:</p> <p>a data.frame with the following added information:</p> <ul style="list-style-type: none"> • SmoothingThe smoothing values correspond to the median of each MSHR (i.e. Region). • BreakpointsThe last position of a region with identical amount of DNA is flagged by 1 otherwise it is 0. Note that during the "Optimization of the number of breakpoints" step, removed breakpoints are flagged by -1. • RegionEach position between two breakpoints are labelled the same way with an integer value starting from one. The label is incremented by one when a new breakpoint is found or when moving to the next chromosome. The variable region is what we call MSHR. • LevelEach position with equal smoothing value is labelled the same way with an integer value starting from one. The label is incremented by one when a new level is found or when moving to the next chromosome. • OutliersAwsEach AWS outliers are flagged -1 or 1 otherwise it is 0. • OutliersMadEach MAD outliers are flagged -1 (if it is in the $\alpha/2$ lower tail of the distribution) or 1 (if it is in the $\alpha/2$ upper tail of the distribution) otherwise it is 0. • OutliersTotOutliersAws + OutliersMad. • ZoneChrClusters identified after MSHR (i.e. Region) clustering by chromosome. • ZoneGenClusters identified after HCSR clustering throughout the genome. • ZoneGNLStatus of each clone : Gain is coded by 1, Loss by -1 and Normal by 0.
BkpInfo:	<p>the data.frame attribute BkpInfo which gives the list of breakpoints:</p> <ul style="list-style-type: none"> • PosOrderThe rank position of each clone on the genome. • PosBaseThe base position of each clone on the genome. • ChromosomeChromosome name.
SigmaC:	<p>the data.frame attribute SigmaC gives the estimation of the LogRatio standard-deviation for each chromosome:</p> <ul style="list-style-type: none"> • ChromosomeChromosome name. • ValueThe estimation is based on the Inter Quartile Range.

Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

Author(s)

Philippe HupÃ©, <glad@curie.fr>.

References

- HupÃ© et al. (Bioinformatics, 2004) Analysis of array CGH data: from signal ratio to gain and loss of DNA regions.
- Polzehl and Spokoiny (WIAS-Preprint 787, 2002) Local likelihood modelling by adaptive weights smoothing.
- Ben-Yaacov and Eldar (Bioinformatics, 2008) A fast and flexible method for the segmentation of aCGH data.

See Also

[profileCGH](#), [as.profileCGH](#), [plotProfile](#).

Examples

```
data(snijders)

### Creation of "profileCGH" object
gm13330$Clone <- gm13330$BAC
profileCGH <- as.profileCGH(gm13330)

#####
###
### glad function as described in HupÃ© et al. (2004)
###
#####

res <- glad(profileCGH, mediancenter=FALSE,
            smoothfunc="lawsglad", bandwidth=10, round=1.5,
            model="Gaussian", lkern="Exponential", qlambda=0.999,
            base=FALSE,
            lambdabreak=8, lambdacluster=8, lambdaclusterGen=40,
            type="tricubic", param=c(d=6),
            alpha=0.001, msize=5,
            method="centroid", nmax=8,
            verbose=FALSE)

### cytoband data to plot chromosomes
data(cytoband)

### Genomic profile on the whole genome
plotProfile(res, unit=3, Bkp=TRUE, labels=FALSE, Smoothing="Smoothing",
            main="Breakpoints detection: GLAD analysis", cytoband = cytoband)

### Genomic profile for chromosome 1
```

```
plotProfile(res, unit=3, Bkp=TRUE, labels=TRUE, Chromosome=1,
Smoothing="Smoothing", main="Chromosome 1: GLAD analysis", cytoband = cytoband)
```

```
### The standard-deviation of LogRatio are:
res$SigmaC
```

```
### The list of breakpoints is:
res$BkpInfo
```

GLAD-internal

GLAD-internal

Description

Internal functions

Usage

" "

Value

" "

Author(s)

Philippe HupÃ©, glad@curie.fr.

See Also

" "

hclustglad

Hierarchical Clustering

Description

Hierarchical cluster analysis on a set of dissimilarities and methods for analyzing it.

Usage

```
hclustglad(d, method = "complete", members=NULL)
```

Arguments

d	a dissimilarity structure as produced by <code>dist</code> .
method	the agglomeration method to be used. This should be (an unambiguous abbreviation of) one of "ward", "single", "complete", "average", "mcquitty", "median" or "centroid".
members	NULL or a vector with length size of d.

Details

This function performs a hierarchical cluster analysis using a set of dissimilarities for the n objects being clustered. Initially, each object is assigned to its own cluster and then the algorithm proceeds iteratively, at each stage joining the two most similar clusters, continuing until there is just a single cluster. At each stage distances between clusters are recomputed by the Lance–Williams dissimilarity update formula according to the particular clustering method being used.

A number of different clustering methods are provided. *Ward's* minimum variance method aims at finding compact, spherical clusters. The *complete linkage* method finds similar clusters. The *single linkage* method (which is closely related to the minimal spanning tree) adopts a ‘friends of friends’ clustering strategy. The other methods can be regarded as aiming for clusters with characteristics somewhere between the single and complete link methods.

If `members` != NULL, then `d` is taken to be a dissimilarity matrix between clusters instead of dissimilarities between singletons and `members` gives the number of observations per cluster. This way the hierarchical cluster algorithm can be “started in the middle of the dendrogram”, e.g., in order to reconstruct the part of the tree above a cut (see examples). Dissimilarities between clusters can be efficiently computed (i.e., without `hclustglad` itself) only for a limited number of distance/linkage combinations, the simplest one being squared Euclidean distance and centroid linkage. In this case the dissimilarities between the clusters are the squared Euclidean distances between cluster means.

In hierarchical cluster displays, a decision is needed at each merge to specify which subtree should go on the left and which on the right. Since, for n observations there are $n - 1$ merges, there are $2^{(n-1)}$ possible orderings for the leaves in a cluster tree, or dendrogram. The algorithm used in `hclustglad` is to order the subtree so that the tighter cluster is on the left (the last, i.e. most recent, merge of the left subtree is at a lower value than the last merge of the right subtree). Single observations are the tightest clusters possible, and merges involving two observations place them in order by their observation sequence number.

Value

An object of class **hclust** which describes the tree produced by the clustering process. The object is a list with components:

merge	an $n - 1$ by 2 matrix. Row i of merge describes the merging of clusters at step i of the clustering. If an element j in the row is negative, then observation $-j$ was merged at this stage. If j is positive then the merge was with the cluster formed at the (earlier) stage j of the algorithm. Thus negative entries in merge indicate agglomerations of singletons, and positive entries indicate agglomerations of non-singletons.
height	a set of $n - 1$ non-decreasing real values. The clustering <i>height</i> : that is, the value of the criterion associated with the clustering method for the particular agglomeration.

order	a vector giving the permutation of the original observations suitable for plotting, in the sense that a cluster plot using this ordering and matrix merge will not have crossings of the branches.
labels	labels for each of the objects being clustered.
call	the call which produced the result.
method	the cluster method that has been used.
dist.method	the distance that has been used to create d (only returned if the distance object has a "method" attribute).

Author(s)

The hclustglad function is based an Algorithm contributed to STATLIB by F. Murtagh.

References

- Everitt, B. (1974). *Cluster Analysis*. London: Heinemann Educ. Books.
- Hartigan, J. A. (1975). *Clustering Algorithms*. New York: Wiley.
- Sneath, P. H. A. and R. R. Sokal (1973). *Numerical Taxonomy*. San Francisco: Freeman.
- Anderberg, M. R. (1973). *Cluster Analysis for Applications*. Academic Press: New York.
- Gordon, A. D. (1999). *Classification*. Second Edition. London: Chapman and Hall / CRC
- Murtagh, F. (1985). "Multidimensional Clustering Algorithms", in *COMPSTAT Lectures 4*. Wuerzburg: Physica-Verlag (for algorithmic details of algorithms used).

Examples

```
data(USArrests)
hc <- hclustglad(dist(USArrests), "ave")
plot(hc)
plot(hc, hang = -1)

## Do the same with centroid clustering and squared Euclidean distance,
## cut the tree into ten clusters and reconstruct the upper part of the
## tree from the cluster centers.
hc <- hclustglad(dist(USArrests)^2, "cen")
memb <- cutree(hc, k = 10)
cent <- NULL
for(k in 1:10){
  cent <- rbind(cent, colMeans(USArrests[memb == k, , drop = FALSE]))
}
hc1 <- hclustglad(dist(cent)^2, method = "cen", members = table(memb))
opar <- par(mfrow = c(1, 2))
plot(hc, labels = FALSE, hang = -1, main = "Original Tree")
plot(hc1, labels = FALSE, hang = -1, main = "Re-start from 10 clusters")
par(opar)
```

kernelpen	<i>Kernelpen function</i>
-----------	---------------------------

Description

Kernel function used in the penalty term.

Usage

```
kernelpen(x, type="tricubic", param)
```

Arguments

x	Real Value.
type	Type of kernelpen to be used
param	a named vector.

Details

The only kernel available is the "tricubic" kernel which takes the values $(1 - (x/d)^3)^3$. The value of d is given by param=c(d=6) for example.

Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

Author(s)

Philippe HupÃ©, <glad@curie.fr>

myPalette	<i>Microarray color palette</i>
-----------	---------------------------------

Description

This function returns a vector of color names corresponding to a range of colors specified in the arguments.

Usage

```
myPalette(low = "white", high = c("green", "red"), mid=NULL, k =50)
```

Arguments

low	Color for the lower end of the color palette, specified using any of the three kinds of R colors, i.e., either a color name (an element of <code>colors</code>), a hexadecimal string of the form "#rrggbb", or an integer <code>i</code> meaning <code>palette()[i]</code> .
high	Color for the upper end of the color palette, specified using any of the three kinds of R colors, i.e., either a color name (an element of <code>colors</code>), a hexadecimal string of the form "#rrggbb", or an integer <code>i</code> meaning <code>palette()[i]</code> .
mid	Color for the middle portion of the color palette, specified using any of the three kinds of R colors, i.e., either a color name (an element of <code>colors</code>), a hexadecimal string of the form "#rrggbb", or an integer <code>i</code> meaning <code>palette()[i]</code> .
k	Number of colors in the palette.

Value

A "character" vector of color names. This can be used to create a user-defined color palette for subsequent graphics by `palette`, in a `col=` specification in graphics functions, or in `par`.

Author(s)

Sandrine Dudoit, Yee Hwa (Jean) Yang.

See Also

`palette`, `rgb`, `colors`, `col2rgb`, `image`, `ColorBar`, `arrayPlot`.

Examples

```
par(mfrow=c(1,4))
pal <- myPalette(low="red", high="green")
ColorBar(seq(-2,2, 0.2), col=pal, horizontal=FALSE, k=21)
pal <- myPalette(low="red", high="green", mid="yellow")
ColorBar(seq(-2,2, 0.2), col=pal, horizontal=FALSE, k=21)
pal <- myPalette()
ColorBar(seq(-2,2, 0.2), col=pal, horizontal=FALSE, k=21)
pal <- myPalette(low="purple", high="purple", mid="white")
ColorBar(seq(-2,2, 0.2), col=pal, horizontal=FALSE, k=21)
```

Description

Plot genomic profile with breakpoints, outliers, smoothing line and cytogenetic banding.

Usage

```
## S3 method for class 'profileCGH'
plotProfile(profileCGH, variable="LogRatio", Chromosome=NULL,
            Smoothing=NULL, GNL="ZoneGNL", Bkp=FALSE,
            labels=TRUE, plotband=TRUE, unit=0,
            colDAGLAD=c("black","blue","red","green","yellow"),
            pchSymbol=c(20,13),
            colCytoBand=c("white","darkblue"),
            colCentro="red", text=NULL,
            cytoband = NULL, main="", ylim=NULL, ...)
```

Arguments

profileCGH	Object of class profileCGH
variable	The variable to be plot.
Chromosome	A numeric vector with chromosome number to be plotted. Use 23 and 24 for chromosome X and Y respectively. If NULL, all the genome is plotted.
Smoothing	The variable used to plot the smoothing line. If NULL, nothing is plotted.
GNL	The variable used to plot the Gain, Normal and Loss color code.
Bkp	If TRUE, the breakpoints are represented by a vertical red dashed line.
labels	If TRUE, the labels of the cytogenetic banding are written.
plotband	If TRUE, the cytogenetic banding are plotted.
unit	Give the unit of the PosBase. For example if unit=3, PosBase are in Kb, if unit=6, PosBase are in Mb, ...
colDAGLAD	Color code to plot Deletion, Amplification, Gain, Lost and Normal status.
pchSymbol	A vector of two elements to specify the symbol tu be used for plotting point. pchSymbol[2] is the symbol for outliers.
colCytoBand	Color code for cytogenetic banding.
colCentro	Color code for centromere.
text	A list with the parameters to be passed to the function text .
cytoband	cytodand data. For human, cytoband data are avaibale using data(cytoband).
main	title of the plot.
ylim	range of the y-axis
...	...

Details

" "

Value

A plot

Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

Author(s)

Philippe HupÃ©, <glad@curie.fr>.

See Also

" "

Examples

```
### Cytogenetic banding information
data(cytoband)

###
data(snijders)

### Creation of "profileCGH" object
profileCGH <- as.profileCGH(gm13330)

#####
###
### glad function as described in HupÃ© et al. (2004)
###
#####

res <- glad(profileCGH, mediancenter=FALSE,
            smoothfunc="lawsglad", bandwidth=10, round=2,
            model="Gaussian", lkern="Exponential", qlambda=0.999,
            base=FALSE,
            lambdabreak=8, lambdacluster=8, lambdaclusterGen=40,
            type="tricubic", param=c(d=6),
            alpha=0.001, msize=5,
            method="centroid", nmax=8,
            verbose=FALSE)

### cytoband data to plot chromosome
data(cytoband)

### Genomic profile on the whole genome
plotProfile(res, unit=3, Bkp=TRUE, labels=FALSE,
            Smoothing="Smoothing", plotband=FALSE, cytoband = cytoband)
```

```

### Genomic profile on the whole genome and cytogenetic banding
plotProfile(res, unit=3, Bkp=TRUE, labels=FALSE, Smoothing="Smoothing",
            cytoband = cytoband)

### Genomic profile for chromosome 1
text <- list(x=c(90000,200000),y=c(0.15,0.3),labels=c("NORMAL","GAIN"), cex=2)
plotProfile(res, unit=3, Bkp=TRUE, labels=TRUE, Chromosome=1,
            Smoothing="Smoothing", plotband=FALSE, text=text, cytoband = cytoband)

### Genomic profile for chromosome 1 and cytogenetic banding with labels
text <- list(x=c(90000,200000),y=c(0.15,0.3),labels=c("NORMAL","GAIN"), cex=2)
plotProfile(res, unit=3, Bkp=TRUE, labels=TRUE, Chromosome=1,
            Smoothing="Smoothing", text=text, main="Chromosome 1", cytoband = cytoband)

```

profileCGH

Objects of Class profileCGH and profileChr

Description

Description of the objects profileCGH and profileChr. The last object corresponds to data of only one chromosome.

Details

LogRatio, Chromosome and PosOrder are compulsory.

Value

Objects profileCGH and profileChr are composed of a list with the first element profileValues which is a data.frame with the following columns names:

LogRatio	Test over Reference log-ratio.
PosOrder	The rank position of each clone on the genome.
PosBase	The base position of each clone on the genome.
Chromosome	Chromosome name.
Clone	The name of the corresponding clone.
...	Other elements can be added.

Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

Author(s)

Philippe HupÃ©, <glad@curie.fr>.

See Also

[glad](#), [as.profileCGH](#).

Examples

```
data(snijders)
gm13330$Clone <- gm13330$BAC
profileCGH <- as.profileCGH(gm13330)
class(profileCGH) <- "profileCGH"

profileChr <- as.profileCGH(gm13330[which(gm13330$Chromosome==1),])
class(profileChr) <- "profileChr"
```

snijders

Public CGH data of Snijders

Description

The data consist of 15 human cell strains with known karyotype (12 fibroblast cell strains, 2 chorionic villus cell strains, 1 lymphoblast cell strain) from the NIGMS Human Genetics Cell Repository (<http://locus.umdj.edu/nigms>). Each cell strain has been hybridized onto a CGH-array of 2276 BAC's spotted in triplicate.

Usage

```
data(snijders)
```

Source

http://www.nature.com/ng/journal/v29/n3/supinfo/ng754_S1.html

References

A M Snijders, N Nowak, R Segreaves, S Blackwood, N Brown, J Conroy, G Hamilton, A K Hindle, B Huey, K Kimura, S Law, K Myambo, J Palmer, B Ylstra, J P Yue, J W Gray, A N Jain, D Pinkel & D G Albertson , Assembly of microarrays for genome-wide measurement of DNA copy number, *Nature Genetics* 29, pp 263 - 264 (2001) Brief Communications.

Examples

```
data(snijders)
array <- gm13330
```

veltman

Public CGH data of Veltman

Description

The data consist of 2 bladder cancer tumors obtained by Veltman et al (2003).

Usage

```
data(veltman)
```

Source

<http://cancerres.aacrjournals.org/cgi/content/full/63/11/2872>

References

Joris A. Veltman, Jane Fridlyand, Sunanda Pejavar, Adam B. Olshen, James E. Korkola, Sandy DeVries, Peter Carroll, Wen-Lin Kuo, Daniel Pinkel, Donna Albertson, Carlos Cordon-Cardo, Ajay N. Jain and Frederic M. Waldman. Array-based Comparative Genomic Hybridization for Genome-Wide Screening of DNA Copy Number in Bladder Tumors. *Cancer Research* 63, 2872-2880, 2003.

Examples

```
data(veltman)  
P9
```

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