

# Package ‘MLP’

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**Title** MLP

**Type** Package

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**Description** Mean Log P Analysis

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**Depends** AnnotationDbi, affy, plotrix, gplots, gmodels, gdata, gtools

**Suggests** GO.db, org.Hs.eg.db, org.Mm.eg.db, org.Rn.eg.db,  
org.Cf.eg.db, KEGG.db, annotate, Rgraphviz, GOstats, limma,  
mouse4302.db, reactome.db

**Collate** 'addGeneSetDescription.R' 'getGeneSets.R' 'mlpBarplot.R'  
'MLP.R' 'plotGeneSetSignificance.R' 'plotGOgraph.R'  
'plot.MLP.R' 'plotQuantileCurves.R' 'utils.R'

**RoxygenNote** 5.0.1

**NeedsCompilation** no

## R topics documented:

addGeneSetDescription . . . . .	2
getGeneSets . . . . .	2
MLP . . . . .	3
mlpBarplot . . . . .	5
plot.MLP . . . . .	6
plotGeneSetSignificance . . . . .	6
plotGOgraph . . . . .	7
<b>Index</b>	<b>9</b>

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`addGeneSetDescription` *Utility function which adds the biological description of the gene sets as a column to the return value of the MLP function (data frame)*

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### Description

Utility function which adds the biological description of the gene sets as a column to the return value of the MLP function (data frame)

### Usage

```
addGeneSetDescription(object, geneSetSource = NULL)
```

### Arguments

`object` object of class 'MLP' as produced by the 'MLP' function

`geneSetSource` source to be used to construct the list of pathway categories; for public data sources, the user can specify a string (one of 'GOBP', 'GOMF', 'GOCC', 'KEGG' or 'REACTOME') and BioC packages will be used to construct the list of pathway categories; for non-public data sources, the user can pass the pathway data as a dataframe with (at least) the following four columns: PATHWAYID, TAXID, PATHWAYNAME and GENEID. It is assumed all columns are of type character. The 'geneSetSource' argument should be the same as the argument provided to the getGeneSets function; defaults to NULL

### Value

the data frame as returned by MLP enriched with an additional column geneSetDescription, providing a concise description of the gene set

### See Also

[MLP](#)

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`getGeneSets` *Prepare Pathway Data for the MLP Function*

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### Description

The return value of the getGeneSets function has as primary use to serve as geneSet argument for the MLP function

### Usage

```
getGeneSets(species = "Mouse", geneSetSource = NULL, entrezIdentifiers)
```

**Arguments**

species	character vector of length one indicating the species, one of 'Mouse', 'Human' or 'Rat'; defaults to 'Mouse'.
geneSetSource	source to be used to construct the list of pathway categories; for public data sources, the user can specify a string (one of 'GOBP', 'GOMF', 'GOCC', 'KEGG' or 'REACTOME') and BioC packages will be used to construct the list of pathway categories; for non-public data sources, the user can pass the pathway data as a dataframe with (at least) the following four columns: PATHWAYID, TAXID, PATHWAYNAME and GENEID. It is assumed all columns are of type character.
entrezIdentifiers	Entrez Gene identifiers used to subset the relevant gene set

**Value**

object of class `geneSetMLP` which is essentially a named list of pathway categories. Each list component contains a vector of Entrez Gene identifiers related to that particular pathway

**Examples**

```
if (require(GO.db) && require(org.Mm.eg.db)){
  pathExampleData <- system.file("exampleFiles", "expressionSetGcrma.rda", package = "MLP")
  pathExamplePValues <- system.file("exampleFiles", "examplePValues.rda", package = "MLP")
  load(pathExampleData)
  load(pathExamplePValues)
  geneSet <- getGeneSets(species = "Mouse", geneSetSource = "GOBP", entrezIdentifiers = names(examplePValues))
  head(geneSet)
}
```

MLP

---

*This function calculates p-values for each gene set based on row permutations of the gene p values or column permutations of the expression matrix; the p values can be obtained either as individual gene set p values or p values based on smoothing across gene sets of similar size.*

---

**Description**

This function calculates p-values for each gene set based on row permutations of the gene p values or column permutations of the expression matrix; the p values can be obtained either as individual gene set p values or p values based on smoothing across gene sets of similar size.

**Usage**

```
MLP(geneSet, geneStatistic, minGenes = 5, maxGenes = 100,
    rowPermutations = TRUE, nPermutations = 100, smoothPValues = TRUE,
    probabilityVector = c(0.5, 0.9, 0.95, 0.99, 0.999, 0.9999, 0.99999),
    df = 9, addGeneSetDescription = TRUE)
```

**Arguments**

<code>geneSet</code>	is the input list of gene sets (components) and gene IDs (character vectors). A gene set can, for example, be a GO category with for each category Entrez Gene identifiers; The <code>getGeneSets</code> function can be used to construct the <code>geneSet</code> argument for different pathway sources.
<code>geneStatistic</code>	is either a named numeric vector (if <code>rowPermutations</code> is TRUE) or a numeric matrix of pvalues (if <code>rowPermutations</code> is FALSE). The names of the numeric vector or row names of the matrix should represent the gene IDs.
<code>minGenes</code>	minimum number of genes in a gene set for it to be considered (lower threshold for gene set size)
<code>maxGenes</code>	maximum number of genes in a gene set for it to be considered (upper threshold for gene set size)
<code>rowPermutations</code>	logical indicating whether to use row permutations (TRUE; default) or column permutations (FALSE)
<code>nPermutations</code>	is the number of simulations. By default 100 permutations are conducted.
<code>smoothPValues</code>	logical indicating whether one wants to calculate smoothed cut-off thresholds (TRUE; default) or not (FALSE).
<code>probabilityVector</code>	vector of quantiles at which p values for each gene set are desired
<code>df</code>	degrees of freedom for the <code>smooth.spline</code> function used in <code>getSmoothedPValues</code>
<code>addGeneSetDescription</code>	logical indicating whether a column with the gene set description be added to the output data frame; defaults to TRUE.

**Value**

data frame with four (or five) columns: `totalGeneSetSize`, `testedGeneSetSize`, `geneSetStatistic` and `geneSetPValue` and (if `addDescription` is set to TRUE) `geneSetDescription`; the rows of the data frame are ordered by ascending `geneSetPValue`.

**References**

Raghavan, Nandini et al. (2007). The high-level similarity of some disparate gene expression measures, *Bioinformatics*, 23, 22, 3032-3038.

**Examples**

```
if (require(GO.db)){
  pathExampleGeneSet <- system.file("exampleFiles", "exampleGeneSet.rda", package = "MLP")
  pathExamplePValues <- system.file("exampleFiles", "examplePValues.rda", package = "MLP")
  load(pathExampleGeneSet)
  load(pathExamplePValues)
  head(examplePValues)
  head(exampleGeneSet)
  mlpResult <- MLP(geneSet = exampleGeneSet, geneStatistic = examplePValues)
  head(mlpResult)
}
```

---

`mlpBarplot`*Draw a Barplot for MLP Results*

---

**Description**

Draw a Barplot for MLP Results

**Usage**

```
mlpBarplot(object, nRow = 20, barColors = NULL, main = NULL, ylab = "",
           cex = 1)
```

**Arguments**

<code>object</code>	object of class MLP
<code>nRow</code>	number of rows of the MLP data frame to depict in the barplot; defaults to 20.
<code>barColors</code>	vector of colors to use for the bars of the barplot; defaults to NULL; if NULL, three gray shades are used reflecting the proportion of tested genes of a gene set versus the total number of genes in a geneset. If the proportion exceeds 75%, the darkest shade is used; between 50 and 75% a moderately dark shade is used; below 50% a lighter gray shade is used.
<code>main</code>	main title; if NULL (default) "Effect of the treatment on <geneSetSource> gene sets" will be used
<code>ylab</code>	string with label for the y-axis
<code>cex</code>	numeric, cex used in par

**Value**

the midpoints of all the bars are returned invisibly (using the conventions of `barplot`); an MLP-specific barplot is drawn to the current device;

**See Also**

`barplot`

**Examples**

```
pathExampleMLPResult <- system.file("exampleFiles", "exampleMLPResult.rda", package = "MLP")
load(pathExampleMLPResult)
dev.new(width = 10, height = 10)
op <- par(mar = c(30, 10, 6, 2))
mlpBarplot(exampleMLPResult)
par(op)
```

---

plot.MLP

*Plot the Results of an MLP Run*


---

**Description**

Plot the Results of an MLP Run

**Usage**

```
## S3 method for class 'MLP'
plot(x, y = NULL, type = c("barplot", "GOgraph",
  "quantileCurves"), ...)
```

**Arguments**

x	object of class 'MLP'
y	argument added to comply with generic; not used, defaults to NULL
type	character of length one; one of 'barplot', 'GOgraph' or 'quantileCurves'
...	further arguments for the plot functions for each type

**Value**

for type = "barplot", the midpoints of the barplot

**Examples**

```
pathExampleMLPResult <- system.file("exampleFiles", "exampleMLPResult.rda", package = "MLP")
load(pathExampleMLPResult)
dev.new(width = 10, height = 10)
op <- par(mar = c(30, 10, 6, 2))
plot(exampleMLPResult, type = "barplot")
par(op)
plot(exampleMLPResult, type = "quantileCurves")
if (require(GO.db) && require(Rgraphviz)){
  plot(exampleMLPResult, type = "GOgraph")
}
```

---

plotGeneSetSignificance

*Plot the Significance for the Genes of a Given Gene Set*


---

**Description**

Plot the Significance for the Genes of a Given Gene Set

**Usage**

```
plotGeneSetSignificance(geneSet, geneSetIdentifier, geneStatistic,
  annotationPackage, barColors = NULL, descriptionInMainTitle = TRUE)
```

**Arguments**

**geneSet** object of class 'geneSetMLP' as produced by function getGeneSets  
**geneSetIdentifier** identifier of the gene set for which a significance plot should be produced; character of length one  
**geneStatistic** named vector of gene statistics (e.g. p values); the names of the vector are Entrez Gene identifiers  
**annotationPackage** name of the annotation package to be used (without .db extension); character of length one  
**barColors** named vector of colors to use for the bars of the barplot; the names of the vector are Entrez Gene identifiers and the vector should be of length equal to the length of the geneStatistic vector defaults to NULL in which case 'grey50' is used  
**descriptionInMainTitle** Boolean whether or not to use the gene set description in the main title of the plot

**Value**

no return value

**Examples**

```

pathExamplePValues <- system.file("exampleFiles", "examplePValues.rda", package = "MLP")
pathExampleGeneSet <- system.file("exampleFiles", "exampleGeneSet.rda", package = "MLP")
pathExampleMLPResult <- system.file("exampleFiles", "exampleMLPResult.rda", package = "MLP")
load(pathExampleGeneSet)
load(pathExamplePValues)
load(pathExampleMLPResult)
# annotationPackage <- if (require(mouse4302mmentrezg.db)) "mouse4302mmentrezg" else "mouse4302"
annotationPackage <- "mouse4302"
geneSetID <- rownames(exampleMLPResult)[1]
dev.new(width = 10, height = 10)
op <- par(mar = c(25, 10, 6, 2))
plotGeneSetSignificance(
  geneSet = exampleGeneSet,
  geneSetIdentifier = geneSetID,
  geneStatistic = examplePValues,
  annotationPackage = annotationPackage
)
par(op)
  
```

---

plotGOgraph

*Graphical Representation of GO Based MLP Results*


---

**Description**

Graphical Representation of GO Based MLP Results

**Usage**

```
plotGOgraph(object, nRow = 5, main = NULL, nCutDescPath = 30)
```

**Arguments**

<code>object</code>	object of class MLP (as produced by the MLP function)
<code>nRow</code>	number of GO IDs for which to produce the plot
<code>main</code>	main title of the graph; if NULL (default) the main title is set to 'GO graph'
<code>nCutDescPath</code>	number of characters at which the pathway description should be cut (inserted in a new line), 30 by default

**Value**

GO graph is plotted to the current device

**Examples**

```
if (require(GO.db) && require(Rgraphviz)){  
  pathExampleMLPResult <- system.file("exampleFiles", "exampleMLPResult.rda", package = "MLP")  
  load(pathExampleMLPResult)  
  plotGOgraph(exampleMLPResult, main = "GO Graph")  
}
```



# Index

`addGeneSetDescription`, [2](#)  
`getGeneSets`, [2](#), [4](#)  
MLP, [2](#), [3](#)  
`mlpBarplot`, [5](#)  
`plot.MLP`, [6](#)  
`plotGeneSetSignificance`, [6](#)  
`plotGograph`, [7](#)