

Package ‘Uniquorn’

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Title Identification of cancer cell lines based on their weighted mutational or variational fingerprint

Version 1.4.2

Description Identifies cancer cell lines with their small variant fingerprint.

Cancer cell line misidentification and cross-

contamination represents a significant challenge for cancer researchers.

The identification is vital and in the frame of this package based on the locations or loci of somatic and germline mutations or variations.

The input format is vcf and the files have to contain a single cancer cell line sample.

The implemented method is optimized for the Next-

generation whole exome and whole genome DNA-sequencing technology. RNA-

seq data is very likely to work as well but hasn't been rigorously tested yet.

Panel-seq will require manual adjustment of thresholds.

Imports DBI, stringr, RSQLite, R.utils, WriteXLS, stats, BiocParallel

Depends R (>= 3.4)

License Artistic-2.0

LazyData TRUE

Type Package

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NeedsCompilation no

Suggests testthat, knitr, rmarkdown, BiocGenerics, RUnit

biocViews Software, StatisticalMethod, WholeGenome, ExomeSeq

VignetteBuilder knitr

R topics documented:

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add_custom_vcf_to_database

Adds a custom vcf file to the three existing cancer cell line panels

Description

Adds a custom vcf file to the three existing cancer cell line panels

Usage

```
add_custom_vcf_to_database(
  vcf_input_files,
  ref_gen = "GRCH37",
  library = "",
  test_mode = FALSE,
  n_threads = 1)
```

Arguments

vcf_input_files	Input vcf file.s This may be one or many vcf files
ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
library	The name of the library to add the CCLs to. Standard is '_CUSTOM' will automatically be added as suffix.
test_mode	Is this a test? Just for internal use
n_threads	Specifies number of threads to be used

Value

Message if the adding has succeeded

Examples

```
HT29_vcf_file = system.file("extdata/HT29.vcf.gz", package="Uniquorn");
add_custom_vcf_to_database(
vcf_input_files = HT29_vcf_file,
library = "",
ref_gen = "GRCH37",
test_mode = TRUE,
n_threads = 1)
```

create_bed_file	<i>create_bed_file</i>
-----------------	------------------------

Description

Creates BED files from the found and not found annotated mutations

Usage

```
create_bed_file(
  sim_list,
  vcf_fingerprint,
  res_table,
  output_file,
  ref_gen,
  manual_identifier
)
```

Arguments

sim_list	R table which contains the mutations from the training database for the cancer cell lines
vcf_fingerprint	contains the mutations that are present in the query cancer cell line's vcf file
res_table	Table containing the identification results
output_file	Path to output file
ref_gen	Reference genome version
manual_identifier	Manually enter a vector of CL name(s) whose bed files should be created, independently from them passing the detection threshold

Value

Returns a message which indicates if the BED file creation has succeeded

identify_vcf_file *identify_VCF_file*

Description

Identifies a cancer cell lines contained in a vcf file based on the pattern (start & length) of all contained mutations/ variations.

Usage

```
identify_vcf_file(
vcf_file,
output_file = "",
ref_gen = "GRCH37",
minimum_matching_mutations = 0,
mutational_weight_inclusion_threshold = 0.5,
only_first_candidate = FALSE,
write_xls = FALSE,
output_bed_file = FALSE,
manual_identifier_bed_file = "",
verbose = FALSE,
p_value = .05,
q_value = .05,
confidence_score = 10.0,
n_threads = 1)
```

Arguments

vcf_file	Input vcf file. Only one sample column allowed.
output_file	Path of the output file. If blank, autogenerated as name of input file plus '_uniquorn_ident.tab' suffix.
ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
minimum_matching_mutations	The minimum amount of mutations that has to match between query and training sample for a positive prediction
mutational_weight_inclusion_threshold	Include only mutations with a weight of at least x. Range: 0.0 to 1.0. 1= unique to CL. ~0 = found in many CL samples.
only_first_candidate	Only the CL identifier with highest score is predicted to be present in the sample
write_xls	Create identification results additionally as xls file for easier reading
output_bed_file	If BED files for IGV visualization should be created for the Cancer Cell lines that pass the threshold
manual_identifier_bed_file	Manually enter a vector of CL name(s) whose bed files should be created, independently from them passing the detection threshold
verbose	Print additional information

p_value	Required p-value for identification
q_value	Required q-value for identification
confidence_score	Threshold above which a positive prediction occurs default 10.0
n_threads	Number of threads to be used

Details

identify_vcf_file parses the vcf file and predicts the identity of the sample

Value

R table with a statistic of the identification result

Examples

```
HT29_vcf_file = system.file("extdata/HT29.vcf.gz", package="Uniquorn");
identification = identify_vcf_file( HT29_vcf_file )
```

```
initiate_canonical_databases
      initiate_canonical_databases
```

Description

Parses data into r list variable

Usage

```
initiate_canonical_databases(
  cosmic_file = "CosmicCLP_MutantExport.tsv",
  ccle_file = "CCLE_hybrid_capture1650_hg19_NoCommonSNPs_CDS_2012.05.07.maf",
  ref_gen = "GRCH37")
```

Arguments

cosmic_file	The path to the cosmic DNA genotype data file. Ensure that the right reference genome is used
ccle_file	The path to the ccle DNA genotype data file. Ensure that the right reference genome is used
ref_gen	Reference genome version

Value

Returns message if parsing process has succeeded

Examples

```
initiate_canonical_databases(
  cosmic_file = "CosmicCLP_MutantExport.tsv",
  ccle_file = "CCLE_hybrid_capture1650_hg19_NoCommonSNPs_CDS_2012.05.07.maf",
  ref_gen = "GRCH37")
```

```
initiate_db_and_load_data
      initiate_db_and_load_data
```

Description

Intern utility function, loads database and return the `sim_list` and `sim_list_stats` variables.

Usage

```
initiate_db_and_load_data(
  ref_gen,
  request_table,
  load_default_db )
```

Arguments

<code>ref_gen</code>	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
<code>request_table</code>	Names of the tables to be extracted from the database
<code>load_default_db</code>	Indicate whether the default db should be used as source for the data

Value

Returns the `sim_list` and `sim_list_stats` variable

```
parse_ccle_genotype_data
      parse_ccle_genotype_data
```

Description

Parses ccle genotype data

Usage

```
parse_ccle_genotype_data(ccle_file, sim_list)
```

Arguments

<code>ccle_file</code>	Path to CCLE file on hard disk
<code>sim_list</code>	Variable containing mutations and cell line

Value

The R Table `sim_list` which contains the CCLE fingerprints

`parse_cosmic_genotype_data`
parse_cosmic_genotype_data

Description

Parses cosmic genotype data

Usage

```
parse_cosmic_genotype_data(cosmic_file, sim_list)
```

Arguments

`cosmic_file` Path to cosmic clp file in hard disk
`sim_list` Variable containing mutations & cell line

Value

The R Table `sim_list` which contains the CoSMIC CLP fingerprints

`parse_vcf_file` *parse_vcf_file*

Description

Parses the vcf file and filters all information except for the start and length of variations/ mutations.

Usage

```
parse_vcf_file( vcf_file_path, n_threads)
```

Arguments

`vcf_file_path` Path to the vcf file on the operating system
`n_threads` Specifies number of threads to be used

Value

Loci-based DNA-mutational fingerprint of the cancer cell line as found in the input VCF file

```
remove_custom_vcf_from_database
```

Removes a cancer cell line training fingerprint (vcf file) from the database. The names of all training sets can be seen by using the function show_contained_cls.

Description

Removes a cancer cell line training fingerprint (vcf file) from the database. The names of all training sets can be seen by using the function show_contained_cls.

Usage

```
remove_custom_vcf_from_database(
  name_cl,
  ref_gen = "GRCH37",
  test_mode = FALSE)
```

Arguments

name_cl	name of the cancer cell line training fingerprint
ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
test_mode	Is this a test? Just for internal use

Value

Message that indicates if the removal was succesful

Examples

```
remove_custom_vcf_from_database(
  name_cl = "HT29_CELLMINER",
  ref_gen = "GRCH37",
  test_mode = TRUE )
```

```
re_calculate_cl_weights
```

Re-calculate sim_list_weights

Description

This function re-calculates the weights of mutation after a change of the training set

Usage

```
re_calculate_cl_weights(sim_list, ref_gen)
```


Arguments

sim_list	R Table which contains a mapping from mutations/ variations to their containing CLs
ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37

Value

A list containing both the sim_list at pos 1 and sim_list_stats at pos 2 data frames.

show_contained_cls	<i>show_contained_cls</i>
--------------------	---------------------------

Description

Show all cancer cell line identifier present in the database for a selected reference genome: This function shows the names, amount of mutations/ variations, overall weight of the mutations of all contained training CLs for a chosen reference genome.

Usage

```
show_contained_cls(  
  ref_gen)
```

Arguments

ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
---------	---

Value

R table which contains the identifier of all cancer cell line samples with the specific reference genome and the weight of all mutations

Examples

```
contained_cls = show_contained_cls(  
  ref_gen = "GRCH37")
```

```
show_contained_mutations  
    show_contained_mutations
```

Description

Show all mutations present in the database for a selected reference Genome: This function shows all training-set mutations for a selected reference genome, i.e. the mutations that are being used for identification of query cancer cell lines.

Usage

```
show_contained_mutations(  
  ref_gen )
```

Arguments

ref_gen Reference genome version

Value

R Table which contains all mutations associated with a particular cancer cell line for a specified reference genome

Examples

```
contained_cls = show_contained_mutations( ref_gen = "GRCH37" )
```

```
show_contained_mutations_for_cl  
    show_contained_mutations_for_cl
```

Description

Show all mutations present in the database for a selected cancer cell line and reference Genome

Usage

```
show_contained_mutations_for_cl(  
  name_cl,  
  ref_gen)
```

Arguments

name_cl Name of the cancer cell line sample stored in the database
ref_gen Reference genome version

Value

R table which contains all mutations associated with the defined cancer cell line and reference genome

Examples

```
SK_OV_3_CELLMINER_mutations = show_contained_mutations_for_cl(
  name_cl = "SK_OV_3_CELLMINER_mutations",
  ref_gen = "GRCH37")
```

```
show_which_cls_contain_mutation
      show_which_cls_contain_mutation
```

Description

Show all cancer cell lines in the database which contained the specified mutation and reference Genome. Closed interval coordinates. Format mutation: CHR_START_STOP, e.g. 1_123_123

Usage

```
show_which_cls_contain_mutation(
  mutation_name,
  ref_gen)
```

Arguments

```
mutation_name  Name of the mutation in the format CHROMOSOME_START_STOP, e.g. '11_244501_244510'
ref_gen        Reference genome version
```

Value

R table which contains all cancer cell line samples which contain the specified mutation with respect to the specified reference genome version

Examples

```
Cls_containing_mutations = show_which_cls_contain_mutation(
  mutation_name = "10_103354427_103354427",
  ref_gen = "GRCH37")
```

```
split_add      split_add
```

Description

```
split_add
```

Usage

```
split_add(vcf_matrix_row)
```

Arguments

```
vcf_matrix_row  row of the vcf file
```

Value

Transformed entry of vcf file, reduced to start and length

write_data_to_db	<i>write_data_to_db</i>
------------------	-------------------------

Description

Intern utility function, writes to database the sim_list and sim_list_stats variables

Usage

```
write_data_to_db(
  content_table,
  table_name,
  ref_gen,
  overwrite,
  test_mode )
```

Arguments

content_table	Tables to be written in db
table_name	Name of the table to be written into the DB
ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
overwrite	Overwrite the potentially existing table
test_mode	Is this a test? Just for internal use

Value

the sim_list and sim_list_stats variable

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