

# Package ‘DAPAR’

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**Type** Package

**Title** Tools for the Differential Analysis of Proteins Abundance with R

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**Description** This package contains a collection of functions for the  
visualisation and the statistical analysis of proteomic data.

**License** Artistic-2.0

**VignetteBuilder** knitr

**Depends** R (>= 3.3)

**Suggests** BiocGenerics, Biobase, testthat, BiocStyle, Prostar

**Imports** MSnbase, RColorBrewer,stats,preprocessCore,Cairo,png,  
lattice,reshape2,gplots,pcaMethods,ggplot2,  
limma,knitr,tmvtnorm,norm,impute, imputeLCMD, doParallel,  
parallel, foreach,grDevices, graphics, openxlsx, utils, cp4p  
(>= 0.3.5), scales, Matrix, vioplot

**biocViews** Proteomics, Normalization, Preprocessing, MassSpectrometry,  
QualityControl, DataImport

**NeedsCompilation** no

**RoxygenNote** 5.0.1

## R topics documented:

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boxPlotD	<i>Builds a boxplot from a dataframe</i>
----------	------------------------------------------

---

### Description

Boxplot for quantitative proteomics data

### Usage

```
boxPlotD(qData, dataForXAxis = NULL, labels = NULL,
         group2Color = "Condition")
```

### Arguments

qData	A dataframe that contains quantitative data.
dataForXAxis	A vector containing the types of replicates to use as X-axis. Available values are: Label, Analyt.Rep, Bio.Rep and Tech.Rep. Default is "Label".
labels	A vector of the conditions (labels) (one label per sample).
group2Color	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

### Value

A boxplot

### Author(s)

Florence Combes, Samuel Wiczorek

### See Also

[densityPlotD](#)

### Examples

```
data(UPSsep25)
qData <- Biobase::exprs(UPSsep25)
types <- c("Label", "Analyt.Rep")
dataForXAxis <- Biobase::pData(UPSsep25)[, types]
labels <- Biobase::pData(UPSsep25)[, "Label"]
boxPlotD(qData, dataForXAxis, labels)
```

---

BuildAdjacencyMatrix *Function matrix of appartenance group*

---

### Description

Method to create a binary matrix with proteins in columns and peptides in lines on a MSnSet object (peptides)

### Usage

```
BuildAdjacencyMatrix(obj.pep, protID, unique = TRUE)
```

### Arguments

obj.pep	An object (peptides) of class <a href="#">MSnbase</a> .
protID	The name of proteins ID column
unique	A boolean to indicate whether only the unique peptides must be considered (TRUE) or if the shared peptides have to be integrated (FALSE).

### Value

A binary matrix

### Author(s)

Florence Combes, Samuel Wiczorek, Alexia Dorffer

### Examples

```
data(UPS pep25)  
BuildAdjacencyMatrix(UPS pep25, "Protein.group.IDs", TRUE)
```

---

BuildColumnToProteinDataset

*creates a column for the protein dataset after agregation by using the previous peptide dataset.*

---

### Description

This function creates a column for the protein dataset after agregation by using the previous peptide dataset.

### Usage

```
BuildColumnToProteinDataset(peptideData, matAdj, columnName)
```

**Arguments**

peptideData	A data.frame of meta data of peptides. It is the fData of the MSnset object.
matAdj	The adjacency matrix used to agregate the peptides data.
columnName	The name of the column in fData(peptides_MSnset) that the user wants to keep in the new protein data.frame.

**Value**

A vector

**Author(s)**

Samuel Wieczorek

**Examples**

```
data(UPSsep25)
protID <- "Protein.group.IDs"
M <- BuildAdjacencyMatrix(UPSsep25, protID, FALSE)
data <- Biobase::fData(UPSsep25)
name <- "organism"
BuildColumnToProteinDataset(data, M, name )
```

---

compareNormalizationD *Builds a plot from a dataframe*

---

**Description**

Plot to compare the quantitative proteomics data before and after normalization

**Usage**

```
compareNormalizationD(qDataBefore, qDataAfter, labelsForLegend = NULL,
  indData2Show = NULL, group2Color = "Condition")
```

**Arguments**

qDataBefore	A dataframe that contains quantitative data before normalization.
qDataAfter	A dataframe that contains quantitative data after normalization.
labelsForLegend	A vector of the conditions (labels) (one label per sample).
indData2Show	A vector of the indices of the columns to show in the plot. The indices are those of indices of the columns int the data.frame qDataBefore.
group2Color	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

**Value**

A plot

**Author(s)**

Samuel Wiczorek

**Examples**

```
data(UPSep25)
qDataBefore <- Biobase::exprs(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
qDataAfter <- normalizeD(qDataBefore, labels, "Median Centering",
"within conditions")
compareNormalizationD(qDataBefore, qDataAfter, labels)
```

---

corrMatrixD

*Displays a correlation matrix of the quantitative data of the exprs() table.*

---

**Description**

Correlation matrix based on a [MSnSet](#) object

**Usage**

```
corrMatrixD(qData, samplesData, gradientRate = 5)
```

**Arguments**

**qData** A dataframe of quantitative data.

**samplesData** A dataframe where lines correspond to samples and columns to the meta-data for those samples.

**gradientRate** The rate parameter to control the exponential law for the gradient of colors

**Value**

A colored correlation matrix

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
data(UPSep25)
qData <- Biobase::exprs(UPSep25)
samplesData <- Biobase::pData(UPSep25)
corrMatrixD(qData, samplesData)
```

---

`CountPep`*Compute the number of peptides used to aggregate proteins*

---

**Description**

This function computes the number of peptides used to aggregate proteins.

**Usage**`CountPep(M)`**Arguments**

`M` A "valued" adjacency matrix in which lines and columns correspond respectively to peptides and proteins.

**Value**

A vector of boolean which is the adjacency matrix but with NA values if they exist in the intensity matrix.

**Author(s)**

Alexia Dorffer

**Examples**

```
data(UPSep25)
protID <- "Protein.group.IDs"
M <- BuildAdjacencyMatrix(UPSep25, protID, FALSE)
CountPep(M)
```

---

`createMSnset`*Creates an object of class `MSnSet` from text file*

---

**Description**

Builds an object of class `MSnSet` from a single tabulated-like file for quantitative and meta-data and a dataframe for the samples description. It differs from the original `MSnSet` builder which requires three separated files tabulated-like quantitative proteomic data into a `MSnSet` object, including meta-data.

**Usage**

```
createMSnset(file, metadata = NULL, indExpData, indFData, indiceID = NULL,
             logData = FALSE, replaceZeros = FALSE, pep_prot_data = NULL)
```

**Arguments**

file	The name of a tab-separated file that contains the data.
metadata	A dataframe describing the samples (in lines).
indExpData	A vector of string where each element is the name of a column in designTable that have to be integrated in the fData() table of the MSnSet object.
indFData	The name of column in file that will be the name of rows for the exprs() and fData() tables
indiceID	The indice of the column containing the ID of entities (peptides or proteins)
logData	A boolean value to indicate if the data have to be log-transformed (Default is FALSE)
replaceZeros	A boolean value to indicate if the 0 and NaN values of intensity have to be replaced by NA (Default is FALSE)
pep_prot_data	A string that indicates whether the dataset is about peptides or proteins.

**Value**

An instance of class `MSnSet`.

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```

exprsFile <- system.file("extdata", "UPSep25.txt", package="DAPAR")
metadataFile <- system.file("extdata", "samples.txt", package="DAPAR")
metadata = read.table(metadataFile, header=TRUE, sep="\t", as.is=TRUE)
indExpData <- c(56:61)
indFData <- c(1:55,62:71)
indiceID <- 64
createMSnset(exprsFile, metadata,indExpData, indFData, indiceID,
pep_prot_data = "peptide")

```

---

deleteLinesFromIndices

*Delete the lines in the matrix of intensities and the metadata table given their indice.*

---

**Description**

Delete the lines of exprs() table identified by their indice.

**Usage**

```
deleteLinesFromIndices(obj, deleteThat = NULL, processText = NULL)
```

**Arguments**

obj	An object of class <code>MSnSet</code> containing quantitative data.
deleteThat	A vector of integers which are the indices of lines to delete.
processText	A string to be included in the <code>MSnSet</code> object for log.



**Value**

An instance of class `MSnSet` that have been filtered.

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
data(UPSep25)
mvFilter(UPSep25, c(1:10))
```

---

densityPlotD	<i>Builds a densityplot from a dataframe</i>
--------------	----------------------------------------------

---

**Description**

Densityplot of quantitative proteomics data over samples.

**Usage**

```
densityPlotD(qData, labelsForLegend = NULL, indData2Show = NULL,
             group2Color = "Condition")
```

**Arguments**

<code>qData</code>	A dataframe that contains quantitative data.
<code>labelsForLegend</code>	A vector of the conditions (labels) (one label per sample).
<code>indData2Show</code>	A vector of indices to show in densityplot. If <code>NULL</code> , then all labels are displayed.
<code>group2Color</code>	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

**Value**

A density plot

**Author(s)**

Florence Combes, Samuel Wiczorek

**See Also**

[boxPlotD](#), [varianceDistD](#)

**Examples**

```
data(UPSep25)
qData <- Biobase::exprs(UPSep25)
labels <- lab2Show <- Biobase::pData(UPSep25)[,"Label"]
densityPlotD(qData, labels)
```

---

diffAna	<i>This function performs a differential analysis on an MSnSet object (adapted from <a href="#">limma</a>)</i>
---------	----------------------------------------------------------------------------------------------------------------

---

**Description**

Performs a differential analysis on an [MSnSet](#) object, based on [limma](#) functions.

**Usage**

```
diffAna(qData, design)
```

**Arguments**

qData	A dataframe that contains quantitative data.
design	The design matrix as described in the <a href="#">limma</a> package documentation

**Value**

A dataframe with the p-value and log(Fold Change) associated to each element (peptide/protein)

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
data(UPSsep25)
qData <- Biobase::exprs(UPSsep25)
design <- cbind(cond1=1, cond2 = rep(0,nrow(Biobase::pData(UPSsep25))))
rownames(design) <- rownames(Biobase::pData(UPSsep25))
labels <- Biobase::pData(UPSsep25)[,"Label"]
indices <- getIndicesConditions(labels, "25fmol", "10fmol")
design[indices$iCond2,2] <- 1
diffAna(qData, design)
```

---

diffAnaComputeFDR	<i>Computes the FDR corresponding to the p-values of the differential analysis using</i>
-------------------	------------------------------------------------------------------------------------------

---

**Description**

This function is a wrapper to the function `adjust.p` from the `cp4p` package. It returns the FDR corresponding to the p-values of the differential analysis. The FDR is computed with the function `p.adjust{stats}`.

**Usage**

```
diffAnaComputeFDR(data, threshold_PVal = 0, threshold_LogFC = 0,
  pi0Method = 1)
```

**Arguments**

data	The result of the differential analysis processed by <a href="#">diffAna</a>
threshold_PVal	The threshold on p-value to distinguish between differential and non-differential data
threshold_LogFC	The threshold on log(Fold Change) to distinguish between differential and non-differential data
pi0Method	The parameter pi0.method of the method adjust.p in the package cp4p

**Value**

The computed FDR value (floating number)

**Author(s)**

Samuel Wieczorek

**Examples**

```
data(UPSsep25)
obj <- wrapper.mvImputation(UPSsep25, "QRILC")
condition1 <- '25fmol'
condition2 <- '10fmol'
qData <- Biobase::exprs(obj)
samplesData <- Biobase::pData(obj)
labels <- Biobase::pData(obj)[,"Label"]
limma <- diffAnaLimma(qData,samplesData, labels, condition1, condition2)
diffAnaComputeFDR(limma)
```

---

diffAnaGetSignificant *Returns a MSnSet object with only proteins significant after differential analysis.*

---

**Description**

Returns a MSnSet object with only proteins significant after differential analysis.

**Usage**

```
diffAnaGetSignificant(obj)
```

**Arguments**

obj An object of class [MSnSet](#).

**Value**

A MSnSet

**Author(s)**

Alexia Dorffer

**Examples**

```

data(UPSep25)
condition1 <- "25fmol"
condition2 <- "10fmol"
resLimma <- wrapper.diffAnaLimma(UPSep25, condition1, condition2)
obj <- diffAnaSave(UPSep25, resLimma, "limma", condition1, condition2)
signif <- diffAnaGetSignificant(obj)

```

---

diffAnaLimma	<i>Performs differential analysis on an MSnSet object, calling the limma package functions</i>
--------------	------------------------------------------------------------------------------------------------

---

**Description**

Method to perform differential analysis on an [MSnSet](#) object (calls the limma package function).

**Usage**

```
diffAnaLimma(qData, samplesData, labels, condition1, condition2)
```

**Arguments**

qData	A dataframe that contains quantitative data.
samplesData	A dataframe where lines correspond to samples and columns to the meta-data for those samples.
labels	A vector of the conditions (labels) (one label per sample).
condition1	A vector that contains the names of the conditions considered as condition 1
condition2	A vector that contains the names of the conditions considered as condition 2

**Value**

A dataframe as returned by the limma package

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```

data(UPSep25)
condition1 <- '25fmol'
condition2 <- '10fmol'
qData <- Biobase::exprs(UPSep25)
samplesData <- Biobase::pData(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
diffAnaLimma(qData, samplesData, labels, condition1, condition2)

```

---

diffAnaSave	Returns a <a href="#">MSnSet</a> object with the results of the differential analysis performed with <a href="#">limma</a> package.
-------------	-------------------------------------------------------------------------------------------------------------------------------------

---

### Description

This method returns a [MSnSet](#) object with the results of differential analysis.

### Usage

```
diffAnaSave(obj, data, method = "limma", condition1, condition2,  
            threshold_pVal = 1e-60, threshold_logFC = 0, fdr = 0,  
            calibrationMethod = "pounds")
```

### Arguments

obj	An object of class <a href="#">MSnSet</a> .
data	The result of the differential analysis processed by <a href="#">diffAna</a>
method	The method used for differential analysis. Available choices are : "limma", "Welch"
condition1	A vector containing the names (some values of the slot "Label" of <code>pData()</code> of the first condition.
condition2	A vector containing the names (some values of the slot "Label" of <code>pData()</code> of the second condition.
threshold_pVal	A float that indicates the threshold on p-value chosen to discriminate differential proteins.
threshold_logFC	A float that indicates the threshold on log(Fold Change) to discriminatedifferential proteins.
fdr	The FDR based on the values of <code>threshold_pVal</code> and <code>threshold_logFC</code>
calibrationMethod	The calibration method used to compute the calibration plot

### Value

A [MSnSet](#)

### Author(s)

Alexia Dorffer, Samuel Wiczorek

### Examples

```
data(UPSsep25)  
condition1 <- '25fmol'  
condition2 <- '10fmol'  
limma <- wrapper.diffAnaLimma(UPSsep25, condition1, condition2)  
obj <- diffAnaSave(UPSsep25, limma, "limma", condition1, condition2)
```

---

diffAnaVolcanoplot      *Volcanoplot of the differential analysis*

---

### Description

Plots a volcano plot after the differential analysis. Typically, the log of Fold Change is represented on the X-axis and the log<sub>10</sub> of the p-value is drawn on the Y-axis. When the `threshold_pVal` and the `threshold_logFC` are set, two lines are drawn respectively on the y-axis and the X-axis to visually distinguish between differential and non differential data.

### Usage

```
diffAnaVolcanoplot(logFC = NULL, pVal = NULL, threshold_pVal = 1e-60,  
  threshold_logFC = 0, conditions = NULL)
```

### Arguments

<code>logFC</code>	A vector of the log(fold change) values of the differential analysis.
<code>pVal</code>	A vector of the p-value values returned by the differential analysis.
<code>threshold_pVal</code>	A floating number which represents the p-value that separates differential and non-differential data.
<code>threshold_logFC</code>	A floating number which represents the log of the Fold Change that separates differential and non-differential data.
<code>conditions</code>	A list of the names of condition 1 and 2 used for the differential analysis.

### Value

A volcano plot

### Author(s)

Florence Combes, Samuel Wiczorek

### Examples

```
data(UPSep25)  
condition1 <- '25fmol'  
condition2 <- '10fmol'  
data <- wrapper.diffAnaLimma(UPSep25, condition1, condition2)  
diffAnaVolcanoplot(data$logFC, data$P.Value)
```

---

diffAnaWelch	<i>Performs a differential analysis on a <code>MSnSet</code> object using the Welch t-test</i>
--------------	------------------------------------------------------------------------------------------------

---

**Description**

Computes differential analysis on an `MSnSet` object, using the Welch t-test (`t.test{stats}`).

**Usage**

```
diffAnaWelch(qData, labels, condition1, condition2)
```

**Arguments**

qData	A dataframe that contains quantitative data.
labels	A vector of the conditions (labels) (one label per sample).
condition1	A vector containing the names of the conditions qData as condition 1
condition2	A vector containing the names of the conditions considered as condition 2

**Value**

A dataframe with two slots : P.Value (for the p-value) and logFC (the log of the Fold Change).

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
data(UPSep25)
condition1 <- '25fmol'
condition2 <- '10fmol'
qData <- Biobase::exprs(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
diffAnaWelch(qData, labels, condition1, condition2)
```

---

getIndicesConditions	<i>Gets the conditions indices.</i>
----------------------	-------------------------------------

---

**Description**

Returns a list for the two conditions where each slot is a vector of indices for the samples.

**Usage**

```
getIndicesConditions(labels, cond1, cond2)
```

**Arguments**

labels	A vector of strings containing the column "Label" of the pData().
cond1	A vector of Labels (a slot in the pData() table) for the condition 1.
cond2	A vector of Labels (a slot in the pData() table) for the condition 2.

**Value**

A list with two slots iCond1 and iCond2 containing respectively the indices of samples in the pData() table of the dataset.

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
data(UPSsep25)
labels <- Biobase::pData(UPSsep25)[,"Label"]
getIndicesConditions(labels, "25fmol", "10fmol")
```

---

getIndicesOfLinesToRemove

*Get the indices of the lines to delete, based on a prefix string*

---

**Description**

This function returns the indice of the lines to delete, based on a prefix string

**Usage**

```
getIndicesOfLinesToRemove(obj, idLine2Delete = NULL, prefix = NULL)
```

**Arguments**

obj	An object of class <a href="#">MSnSet</a> .
idLine2Delete	The name of the column that correspond to the data to filter
prefix	A character string that is the prefix to find in the data

**Value**

A vector of integers.

**Author(s)**

Samuel Wiczorek

**Examples**

```
data(UPSsep25)
getIndicesOfLinesToRemove(UPSsep25, "Potential.contaminant", prefix="+")
```



---

getNumberOf	<i>Number of lines with prefix</i>
-------------	------------------------------------

---

**Description**

Returns the number of lines, in a given column, where content matches the prefix.

**Usage**

```
getNumberOf(obj, name = NULL, prefix = NULL)
```

**Arguments**

obj	An object of class <a href="#">MSnSet</a> .
name	The name of a column.
prefix	A string

**Value**

An integer

**Author(s)**

Samuel Wieczorek

**Examples**

```
data(UPSpep25)
getNumberOf(UPSpep25, "Potential.contaminant", "+")
```

---

getNumberOfEmptyLines	<i>Returns the number of empty lines in the data</i>
-----------------------	------------------------------------------------------

---

**Description**

Returns the number of empty lines in a matrix.

**Usage**

```
getNumberOfEmptyLines(qData)
```

**Arguments**

qData	A matrix corresponding to the quantitative data.
-------	--------------------------------------------------

**Value**

An integer

**Author(s)**

Samuel Wieczorek

**Examples**

```
data(UPSep25)
qData <- Biobase::exprs(UPSep25)
getNumberOfEmptyLines(qData)
```

---

getPaletteForLabels     *Palette for plots in DAPAR*

---

**Description**

Selects colors for the plots in DAPAR based on the different conditions in the dataset. The palette is derived from the brewer palette "Dark2" (see [RColorBrewer](#)).

**Usage**

```
getPaletteForLabels(labels)
```

**Arguments**

labels                    A vector of labels (strings).

**Value**

A palette designed for the data manipulated in DAPAR

**Author(s)**

Florence Combes, Samuel Wieczorek

**Examples**

```
data(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
getPaletteForLabels(labels)
```

---

`getPaletteForReplicates`*Palette for plot the replicates in DAPAR*

---

**Description**

Selects colors for the plots in DAPAR based on the replicates in the dataset. The palette is derived from the brewer palette "Dark2" (see [RColorBrewer](#)).

**Usage**

```
getPaletteForReplicates(nColors)
```

**Arguments**

`nColors`            The desired number of colors

**Value**

A palette designed for the data manipulated in DAPAR

**Author(s)**

Samuel Wieczorek

**Examples**

```
data(UPSsep25)
n <- nrow(Biobase::pData(UPSsep25))
getPaletteForLabels(5)
```

---

`getPourcentageOfMV`*Percentage of missing values*

---

**Description**

Returns the percentage of missing values in the quantitative data (`exprs()` table of the dataset).

**Usage**

```
getPourcentageOfMV(obj)
```

**Arguments**

`obj`                An object of class [MSnSet](#).

**Value**

A floating number

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
data(UPSpep25)
getPourcentageOfMV(UPSpep25)
```

---

getProcessingInfo	<i>Returns the contains of the slot processing of an object of class MSnSet</i>
-------------------	---------------------------------------------------------------------------------

---

**Description**

Returns the contains of the slot processing of an object of class MSnSet.

**Usage**

```
getProcessingInfo(obj)
```

**Arguments**

obj                    An object (peptides) of class [MSnbase](#).

**Value**

The slot processing of obj@processingData

**Author(s)**

Samuel Wiczorek

**Examples**

```
data(UPSpep25)
getProcessingInfo(UPSpep25)
```

---

getProteinsStats	<i>computes the number of proteins that are only defined by specific peptides, shared peptides or a mixture of two.</i>
------------------	-------------------------------------------------------------------------------------------------------------------------

---

**Description**

This function computes the number of proteins that are only defined by specific peptides, shared peptides or a mixture of two.

**Usage**

```
getProteinsStats(matUnique, matShared)
```

**Arguments**

matUnique        The adjacency matrix with only specific peptides.  
matShared        The adjacency matrix with both specific and shared peptides.

**Value**

A list

**Author(s)**

Samuel Wiczorek

**Examples**

```
data(UPSep25)  
protID <- "Protein.group.IDs"  
MShared <- BuildAdjacencyMatrix(UPSep25, protID, FALSE)  
MUnique <- BuildAdjacencyMatrix(UPSep25, protID, TRUE)  
getProteinsStats(MUnique, MShared)
```

---

GraphPepProt	<i>Function to create a histogram that shows the repartition of peptides w.r.t. the proteins</i>
--------------	--------------------------------------------------------------------------------------------------

---

**Description**

Method to create a plot with proteins and peptides on a MSnSet object (peptides)

**Usage**

```
GraphPepProt(mat)
```

**Arguments**

mat                An adjacency matrix.

**Value**

A histogram

**Author(s)**

Alexia Dorffer, Samuel Wiczorek

**Examples**

```
data(UPSep25)  
mat <- BuildAdjacencyMatrix(UPSep25, "Protein.group.IDs")  
GraphPepProt(mat)
```

---

heatmap.DAPAR	<i>This function is inspired from the function <a href="#">heatmap.2</a> that displays quantitative data in the <code>exprs()</code> table of an object of class <code>MSet</code>. For more information, please refer to the help of the <code>heatmap.2</code> function.</i>
---------------	--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------

---

## Description

Heatmap inspired by the `heatmap.2` function.

## Usage

```
heatmap.DAPAR(x, col = heat.colors(100), srtCol = NULL, labCol = NULL,  
  labRow = NULL, key = TRUE, key.title = NULL, main = NULL,  
  ylab = NULL)
```

## Arguments

<code>x</code>	A dataframe that contains quantitative data.
<code>col</code>	colors used for the image. Defaults to heat colors ( <code>heat.colors</code> ).
<code>srtCol</code>	angle of column labels, in degrees from horizontal
<code>labCol</code>	character vectors with column labels to use.
<code>labRow</code>	character vectors with row labels to use.
<code>key</code>	logical indicating whether a color-key should be shown.
<code>key.title</code>	main title of the color key. If set to <code>NA</code> no title will be plotted.
<code>main</code>	main title; default to none.
<code>ylab</code>	y-axis title; default to none.

## Value

A heatmap

## Author(s)

Samuel Wieczorek

## Examples

```
data(testWithoutNA)  
qData <- Biobase::exprs(testWithoutNA)  
heatmapD(qData)
```

---

heatmapD	<i>This function is a wrapper to <a href="#">heatmap.2</a> that displays quantitative data in the <code>exprs()</code> table of an object of class <code>MSnSet</code></i>
----------	----------------------------------------------------------------------------------------------------------------------------------------------------------------------------

---

**Description**

Heatmap of the quantitative proteomic data of a `MSnSet` object

**Usage**

```
heatmapD(qData, distance = "euclidean", cluster = "average",  
         dendro = FALSE)
```

**Arguments**

qData	A dataframe that contains quantitative data.
distance	The distance used by the clustering algorithm to compute the dendrogram. See <code>help(heatmap.2)</code>
cluster	the clustering algorithm used to build the dendrogram. See <code>help(heatmap.2)</code>
dendro	A boolean to indicate if the dendrogram has to be displayed

**Value**

A heatmap

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
data(testWithoutNA)  
qData <- Biobase::exprs(testWithoutNA)  
heatmapD(qData)
```

---

limmaCompleteTest	<i>Computes a hierarchical differential analysis</i>
-------------------	------------------------------------------------------

---

**Description**

This function is a `limmaCompleteTest`

**Usage**

```
limmaCompleteTest(qData, Conditions, RepBio, RepTech, Contrast = 1)
```

**Arguments**

qData	A matrix of quantitative data, without any missing values.
Conditions	A vector of factor which indicates the name of the biological condition for each replicate.
RepBio	A vector of factor which indicates the number of the bio rep for each replicate.
RepTech	A vector of factor which indicates the number of the tech rep for each replicate.
Contrast	Indicates if the test consists of the comparison of each biological condition versus each of the other ones (Contrast=1; for example H0:"C1=C2" vs H1:"C1!=C2", etc.) or each condition versus all others (Contrast=2; e.g. H0:"C1=(C2+C3)/2" vs H1:"C1!=(C2+C3)/2", etc. if there are three conditions).

**Value**

fdsgfdg

**Author(s)**

Quentin Giai-Gianetto

**Examples**

```
data(UPSsep25)
obj <- wrapper.mvImputation(UPSsep25, "QRILC")
condition1 <- '25fmol'
condition2 <- '10fmol'
qData <- Biobase::exprs(obj)
RepBio <- RepTech <- factor(1:6)
conds <- factor(c(rep(condition1, 3), (rep(condition2, 3))))
limma <- limmaCompleteTest(qData,conds,RepBio, RepTech)
```

---

MeanPeptides	<i>Compute the intensity of proteins as the mean of the intensities of their peptides.</i>
--------------	--------------------------------------------------------------------------------------------

---

**Description**

This function computes the intensity of proteins as the mean of the intensities of their peptides.

**Usage**

```
MeanPeptides(matAdj, expr)
```

**Arguments**

matAdj	An adjacency matrix in which lines and columns correspond respectively to peptides and proteins.
expr	A matrix of intensities of peptides

**Value**

A matrix of intensities of proteins



**Author(s)**

Alexia Dorffer

**Examples**

```
data(UPSep25)
protID <- "Protein.group.IDs"
matAdj <- BuildAdjacencyMatrix(UPSep25, protID, FALSE)
MeanPeptides(matAdj, Biobase::exprs(UPSep25))
```

---

mvFilter

*Filter lines in the matrix of intensities w.r.t. some criteria*

---

**Description**

Filters the lines of `exprs()` table with conditions on the number of missing values. The user chooses the minimum amount of intensities that is acceptable and the filter delete lines that do not respect this condition. The condition may be on the whole line or condition by condition.

**Usage**

```
mvFilter(obj, type, th, processText = NULL)
```

**Arguments**

<code>obj</code>	An object of class <code>MSnSet</code> containing quantitative data.
<code>type</code>	Method used to choose the lines to delete. Values are : "none", "wholeMatrix", "allCond", "atLeastOneCond"
<code>th</code>	An integer value of the threshold
<code>processText</code>	A string to be included in the <code>MSnSet</code> object for log.

**Details**

The different methods are : "wholeMatrix": given a threshold `th`, only the lines that contain at least `th` values are kept. "allCond": given a threshold `th`, only the lines which contain at least `th` values for each of the conditions are kept. "atLeastOneCond": given a threshold `th`, only the lines that contain at least `th` values, and for at least one condition, are kept.

**Value**

An instance of class `MSnSet` that have been filtered.

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
data(UPSep25)
mvFilter(UPSep25, "wholeMatrix", 2)
```

mvFilterFromIndices     *Filter lines in the matrix of intensities w.r.t. some criteria*

---

### Description

Filters the lines of `exprs()` table with conditions on the number of missing values. The user chooses the minimum amount of intensities that is acceptable and the filter delete lines that do not respect this condition. The condition may be on the whole line or condition by condition.

### Usage

```
mvFilterFromIndices(obj, keepThat = NULL, processText = NULL)
```

### Arguments

<code>obj</code>	An object of class <code>MSnSet</code> containing quantitative data.
<code>keepThat</code>	A vector of integers which are the indices of lines to keep.
<code>processText</code>	A string to be included in the <code>MSnSet</code> object for log.

### Details

The different methods are : "wholeMatrix": given a threshold `th`, only the lines that contain at least `th` values are kept. "allCond": given a threshold `th`, only the lines which contain at least `th` values for each of the conditions are kept. "atLeastOneCond": given a threshold `th`, only the lines that contain at least `th` values, and for at least one condition, are kept.

### Value

An instance of class `MSnSet` that have been filtered.

### Author(s)

Florence Combes, Samuel Wiczorek

### Examples

```
data(UPSep25)
mvFilter(UPSep25, c(1:10))
```

---

mvFilterGetIndices     *Filter lines in the matrix of intensities w.r.t. some criteria*

---

### Description

Returns the indices of the lines of `exprs()` table to delete w.r.t. the conditions on the number of missing values. The user chooses the minimum amount of intensities that is acceptable and the filter delete lines that do not respect this condition. The condition may be on the whole line or condition by condition.

**Usage**

```
mvFilterGetIndices(obj, type, th)
```

**Arguments**

obj	An object of class <code>MSnSet</code> containing quantitative data.
type	Method used to choose the lines to delete. Values are : "none", "wholeMatrix", "allCond", "atLeastOneCond"
th	An integer value of the threshold

**Details**

The different methods are : "wholeMatrix": given a threshold `th`, only the lines that contain at least `th` values are kept. "allCond": given a threshold `th`, only the lines which contain at least `th` values for each of the conditions are kept. "atLeastOneCond": given a threshold `th`, only the lines that contain at least `th` values, and for at least one condition, are kept.

**Value**

An vector of indices that correspond to the lines to keep.

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
data(UPSep25)
mvFilterGetIndices(UPSep25, "wholeMatrix", 2)
```

---

mvHisto

*Histogram of missing values*


---

**Description**

This method plots a histogram of missing values.

**Usage**

```
mvHisto(qData, samplesData, labels, indLegend = "auto", showValues = FALSE)
```

**Arguments**

qData	A dataframe that contains quantitative data.
samplesData	A dataframe where lines correspond to samples and columns to the meta-data for those samples.
labels	A vector of the conditions (labels) (one label per sample).
indLegend	The indices of the column name's in <code>pData()</code> tab
showValues	A logical that indicates wether numeric values should be drawn above the bars.

**Value**

A histogram

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
data(UPSep25)
qData <- Biobase::exprs(UPSep25)
samplesData <- Biobase::pData(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
mvHisto(qData, samplesData, labels, indLegend="auto", showValues=TRUE)
```

---

mvImage

*Heatmap of missing values*

---

**Description**

Plots a heatmap of the quantitative data. Each column represent one of the conditions in the object of class [MSnSet](#) and the color is proportional to the mean of intensity for each line of the dataset. The lines have been sorted in order to visualize easily the different number of missing values. A white square is plotted for missing values.

**Usage**

```
mvImage(qData, labels)
```

**Arguments**

qData	A dataframe that contains quantitative data.
labels	A vector of the conditions (labels) (one label per sample).

**Value**

A heatmap

**Author(s)**

Samuel Wiczorek, Thomas Burger

**Examples**

```
data(UPSep25)
qData <- Biobase::exprs(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
mvImage(qData, labels)
```

---

mvImputation	<i>Missing values imputation from a matrix</i>
--------------	------------------------------------------------

---

**Description**

This method is a wrapper to the `imputeLCMD` package adapted to a matrix.

**Usage**

```
mvImputation(qData, method)
```

**Arguments**

qData	A dataframe that contains quantitative data.
method	The imputation method to be used. Choices are QRILC, KNN, BPCA and MLE.

**Value**

The matrix imputed

**Author(s)**

Samuel Wieczorek

**Examples**

```
data(UPSsep25)
qData <- Biobase::exprs(UPSsep25)
mvImputation(qData, "QRILC")
```

---

mvPerLinesHisto	<i>Bar plot of missing values per lines</i>
-----------------	---------------------------------------------

---

**Description**

This method plots a bar plot which represents the distribution of the number of missing values (NA) per lines (ie proteins).

**Usage**

```
mvPerLinesHisto(qData, samplesData, indLegend = "auto", showValues = FALSE)
```

**Arguments**

qData	A dataframe that contains the data to plot.
samplesData	A dataframe which contains informations about the replicates.
indLegend	The indice of the column name's in <code>pData()</code> tab
showValues	A logical that indicates wether numeric values should be drawn above the bars.

**Value**

A bar plot

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
data(UPSsep25)
qData <- Biobase::exprs(UPSsep25)
samplesData <- Biobase::pData(UPSsep25)
mvPerLinesHisto(qData, samplesData)
```

---

mvPerLinesHistoPerCondition

*Bar plot of missing values per lines and per condition*

---

**Description**

This method plots a bar plot which represents the distribution of the number of missing values (NA) per lines (ie proteins) and per conditions.

**Usage**

```
mvPerLinesHistoPerCondition(qData, samplesData, indLegend = "auto",
  showValues = FALSE)
```

**Arguments**

qData	A dataframe that contains quantitative data.
samplesData	A dataframe where lines correspond to samples and columns to the meta-data for those samples.
indLegend	The indice of the column name's in pData() tab
showValues	A logical that indicates wether numeric values should be drawn above the bars.

**Value**

A bar plot

**Author(s)**

Samuel Wiczorek

**Examples**

```
data(UPSsep25)
qData <- Biobase::exprs(UPSsep25)
samplesData <- Biobase::pData(UPSsep25)
mvPerLinesHistoPerCondition(qData, samplesData)
```

mvTypePlot

*Distribution of missing values with respect to intensity values***Description**

This method plots a scatter plot which represents the distribution of missing values. The colors correspond to the different conditions (slot Label in in the dataset of class `MsnSet`). The x-axis represent the mean of intensity for one condition and one entity in the dataset (i. e. a protein) whereas the y-axis count the number of missing values for this entity and the considered condition. The data have been jittered for an easier vizualisation.

**Usage**

```
mvTypePlot(qData, labels, threshold = 0)
```

**Arguments**

qData	A dataframe that contains quantitative data.
labels	A vector of the conditions (labels) (one label per sample).
threshold	An integer for the intensity that delimits MNAR and MCAR missing values.

**Value**

A scatter plot

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
data(UPSep25)
qData <- Biobase::exprs(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
mvTypePlot(qData, labels, threshold=0)
```

normalized

*Normalisation***Description**

Provides several methods to normalize data from a matrix. They are organized in four main families : Strong Rescaling, Median Centering, Mean Centering, Mean CenteringScaling. For the first family, two sub-categories are available : the sum by columns and the quantiles method. For the three other families, two categories are available : "Overall" which means that the value for each protein (ie line in the expression data tab) is computed over all the samples ; "within conditions" which means that the value for each protein (ie line in the matrix) is computed condition by condition.

**Usage**

```
normalized(qData, labels, family, method)
```

**Arguments**

qData	A dataframe that contains quantitative data.
labels	A vector of strings containing the column "Label" of the pData().
family	One of the following : Global Rescaling, Median Centering, Mean Centering, Mean Centering Scaling.
method	"Overall" or "within conditions".

**Value**

A matrix normalized

**Author(s)**

Florence Combes, Samuel Wiczorek

**Examples**

```
data(UPSsep25)
qData <- Biobase::exprs(UPSsep25)
labels <- Biobase::pData(UPSsep25)[,"Label"]
normalizedD(qData, labels, "Median Centering", "within conditions")
```

---

pepAgregate	<i>Function agregate peptides to proteins</i>
-------------	-----------------------------------------------

---

**Description**

Method to agregate with a method peptides to proteins on a MSnSet object (peptides)

**Usage**

```
pepAgregate(obj.pep, protID, method = "sum overall", matAdj = NULL,
  n = NULL)
```

**Arguments**

obj.pep	An object (peptides) of class <a href="#">MSnbase</a> .
protID	The name of proteins ID column
method	The method used to aggregate the peptides into proteins. Values are "sum", "mean" or "sum on top n" : do the sum / mean of intensity on all peptides belonging to proteins. Default is "sum"
matAdj	An adjacency matrix
n	The number of peptides considered for the aggregation.

**Value**

An object of class [MSnbase](#) with proteins



**Author(s)**

Alexia Dorffer, Samuel Wiczorek

**Examples**

```
data(UPSep25)
protID <- "Protein.group.IDs"
mat <- BuildAdjacencyMatrix(UPSep25, protID, TRUE)
pepAggregate(UPSep25, protID, "sum overall", mat)
```

---

proportionConRev      *Barplot of proportion of contaminants and reverse*

---

**Description**

Plots a barplot of proportion of contaminants and reverse

**Usage**

```
proportionConRev(obj, idContaminants = NULL, prefixContaminants = NULL,
  idReverse = NULL, prefixReverse = NULL)
```

**Arguments**

obj                    An object of class [MSnSet](#).  
idContaminants      The name of a column of Contaminants  
prefixContaminants      The prefix to identify contaminants  
idReverse            The name of a column of Reverse  
prefixReverse        The prefix to identify Reverse

**Value**

A barplot

**Author(s)**

Samuel Wiczorek

**Examples**

```
data(UPSep25)
pref <- "+"
proportionConRev(UPSep25, "Potential.contaminant", pref, "Reverse", pref)
```

---

removeLines	<i>Removes lines in the dataset based on a prefix string.</i>
-------------	---------------------------------------------------------------

---

**Description**

This function removes lines in the dataset based on a prefix string.

**Usage**

```
removeLines(obj, idLine2Delete = NULL, prefix = NULL)
```

**Arguments**

obj	An object of class <a href="#">MSnSet</a> .
idLine2Delete	The name of the column that correspond to the data to filter
prefix	A character string that is the prefix to find in the data

**Value**

An object of class [MSnSet](#).

**Author(s)**

Samuel Wieczorek

**Examples**

```
data(UPSsep25)
removeLines(UPSsep25, "Potential.contaminant")
removeLines(UPSsep25, "Reverse")
```

---

SumPeptides	<i>Compute the intensity of proteins with the sum of the intensities of their peptides.</i>
-------------	---------------------------------------------------------------------------------------------

---

**Description**

This function computes the intensity of proteins based on the sum of the intensities of their peptides.

**Usage**

```
SumPeptides(matAdj, expr)
```

**Arguments**

matAdj	An adjacency matrix in which lines and columns correspond respectively to peptides and proteins.
expr	A matrix of intensities of peptides

**Value**

A matrix of intensities of proteins

**Author(s)**

Alexia Dorffer

**Examples**

```
data(UPSep25)
protID <- "Protein.group.IDs"
M <- BuildAdjacencyMatrix(UPSep25, protID, FALSE)
SumPeptides(M, Biobase::exprs(UPSep25))
```

---

test	<i>Test dataset</i>
------	---------------------

---

**Description**

Partial (small) dataset for unit tests containing missing values.

**Format**

An object of class [MSnSet](#)

---

testWithoutNA	<i>Test dataset</i>
---------------	---------------------

---

**Description**

Partial (small) dataset for unit tests without any missing values.

**Format**

An object of class [MSnSet](#)

---

TopnPeptides	<i>Compute the intensity of proteins as the sum of the intensities of their n best peptides.</i>
--------------	--------------------------------------------------------------------------------------------------

---

### Description

This function computes the intensity of proteins as the sum of the intensities of their n best peptides.

### Usage

```
TopnPeptides(matAdj, expr, n)
```

### Arguments

matAdj	An adjacency matrix in which lines and columns correspond respectively to peptides and proteins.
expr	A matrix of intensities of peptides
n	The maximum number of peptides used to aggregate a protein.

### Value

A matrix of intensities of proteins

### Author(s)

Alexia Dorffer

### Examples

```
data(UPSpep25)
protID <- "Protein.group.IDs"
matAdj <- BuildAdjacencyMatrix(UPSpep25, protID, FALSE)
TopnPeptides(matAdj, Biobase::exprs(UPSpep25), 3)
```

---

UPSpep25	<i>UPSpep25 dataset</i>
----------	-------------------------

---

### Description

This dataset is the final outcome of a quantitative mass spectrometry-based proteomic analysis of two samples containing different concentrations of 48 human proteins (UPS1 standard from Sigma-Aldrich) within a constant yeast background (see Gaii Gianetto et al. (2016) for details). It contains the abundance values of the different human and yeast peptides identified and quantified in these two conditions. The two conditions represent the measured abundances of peptides when respectively 25fmol and 10fmol of UPS1 human proteins were mixed with the yeast extract before mass spectrometry analyses. Three technical replicates were acquired for each condition.

To identify and quantify peptides, spectra were searched using MaxQuant (version 1.5.1.2) against the Uniprot database, the UPS database and the frequently observed contaminants database. Maximum false discovery rates were set to 0.01 by employing a reverse database strategy.

The dataset is either available as a CSV file (see inst/extdata/UPSpep25.txt), or as a [MSnSet](#) structure (UPSpep25). In the latter case, the quantitative data are those of the raw intensities.

**Usage**

```
data(UPSep25)
```

**Format**

An object of class `MSnSet` related to peptide quantification. It contains 6 samples divided into two conditions (25fmol and 10fmol) and 13918 peptides.

The data frame `exprs(UPSep25)` contains six columns that are the quantitation of peptides for the six replicates.

The data frame `fData(UPSep25)` contains the meta data about the peptides.

The data frame `pData(UPSep25)` contains the experimental design and gives few informations about the samples.

**Value**

An object of class `MSnSet`.

**References**

Cox J., Hein M.Y., Lubner C.A., Paron I., Nagaraj N., Mann M. Accurate proteome-wide label-free quantification by delayed normalization and maximal peptide ratio extraction, termed MaxLFQ. *Mol Cell Proteomics*. 2014 Sep, 13(9):2513-26.

Giai Gianetto, Q., Combes, F., Ramus, C., Bruley, C., Coute, Y., Burger, T. (2016). Calibration plot for proteomics: A graphical tool to visually check the assumptions underlying FDR control in quantitative experiments. *Proteomics*, 16(1), 29-32.

---

varianceDistD	<i>Distribution of variance of proteins</i>
---------------	---------------------------------------------

---

**Description**

Builds a densityplot of the variance of entities in the `exprs()` table of a object. The variance is calculated for each condition (Label) present in the dataset (see the slot 'Label' in the `pData()` table)

**Usage**

```
varianceDistD(qData, labels = NULL)
```

**Arguments**

<code>qData</code>	A dataframe that contains quantitative data.
<code>labels</code>	A vector of the conditions (labels) (one label per sample).

**Value**

A density plot

**Author(s)**

Florence Combes, Samuel Wiczorek

**See Also**

[densityPlotD](#).

**Examples**

```
data(UPSsep25)
labels <- Biobase::pData(UPSsep25)[,"Label"]
varianceDistD(UPSsep25)
```

---

violinPlotD

*Builds a violinplot from a dataframe*

---

**Description**

ViolinPlot for quantitative proteomics data

**Usage**

```
violinPlotD(qData, dataForXAxis = NULL, labels = NULL,
            group2Color = "Condition")
```

**Arguments**

qData	A dataframe that contains quantitative data.
dataForXAxis	A vector containing the types of replicates to use as X-axis. Available values are: Label, Analyt.Rep, Bio.Rep and Tech.Rep. Default is "Label".
labels	A vector of the conditions (labels) (one label per sample).
group2Color	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

**Value**

A violinplot

**Author(s)**

Florence Combes, Samuel Wiczorek

**See Also**

[densityPlotD](#)

**Examples**

```
data(UPSsep25)
library(vioplot)
qData <- Biobase::exprs(UPSsep25)
types <- c("Label", "Analyt.Rep")
dataForXAxis <- Biobase::pData(UPSsep25)[,types]
labels <- Biobase::pData(UPSsep25)[,"Label"]
violinPlotD(qData, dataForXAxis, labels)
```

---

wrapper.boxPlotD      *Wrapper to the boxplotD function on an object MSnSet*

---

### Description

This function is a wrapper for using the boxPlotD function with objects of class [MSnSet](#)

### Usage

```
wrapper.boxPlotD(obj, dataForXAxis = "Label", group2Color = "Condition")
```

### Arguments

obj	An object of class <a href="#">MSnSet</a> .
dataForXAxis	A vector of strings containing the names of columns in <code>pData()</code> to print labels on X-axis (Default is "Label").
group2Color	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

### Value

A boxplot

### Author(s)

Florence Combes, Samuel Wiczorek

### See Also

[wrapper.densityPlotD](#)

### Examples

```
data(UPSspep25)
types <- c("Label", "Analyt.Rep")
wrapper.boxPlotD(UPSspep25, types)
```

---

wrapper.compareNormalizationD  
*Builds a plot from a dataframe*

---

### Description

Wrapper to the function that plot to compare the quantitative proteomics data before and after normalization

### Usage

```
wrapper.compareNormalizationD(objBefore, objAfter, labelsForLegend = NULL,
  indData2Show = NULL, group2Color = "Condition")
```

**Arguments**

objBefore	A dataframe that contains quantitative data before normalization.
objAfter	A dataframe that contains quantitative data after normalization.
labelsForLegend	A vector of the conditions (labels) (one label per sample).
indData2Show	A vector of the indices of the columns to show in the plot. The indices are those of indices of the columns in the data.frame qDataBefore.
group2Color	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

**Value**

A plot

**Author(s)**

Samuel Wieczorek

**Examples**

```
data(UPSsep25)
labels <- Biobase::pData(UPSsep25)[,"Label"]
objAfter <- wrapper.normalized(UPSsep25, "Median Centering",
"within conditions")
wrapper.compareNormalizationD(UPSsep25, objAfter, labels)
```

---

wrapper.corrMatrixD	<i>Displays a correlation matrix of the quantitative data of the exprs() table</i>
---------------------	------------------------------------------------------------------------------------

---

**Description**

Builds a correlation matrix based on a [MSnSet](#) object.

**Usage**

```
wrapper.corrMatrixD(obj, rate = 5)
```

**Arguments**

obj	An object of class <a href="#">MSnSet</a> .
rate	A float that defines the gradient of colors.

**Value**

A colored correlation matrix

**Author(s)**

Alexia Dorffer



## Examples

```
data(UPSep25)
wrapper.corrMatrixD(UPSep25)
```

---

wrapper.densityPlotD *Builds a densityplot from an object of class [MSnSet](#)*

---

## Description

This function is a wrapper for using the densityPlotD function with objects of class [MSnSet](#)

## Usage

```
wrapper.densityPlotD(obj, labelsForLegend = NULL, indData2Show = NULL,
  group2Color = "Condition")
```

## Arguments

obj	An object of class <a href="#">MSnSet</a> .
labelsForLegend	A vector of labels to show in densityplot.
indData2Show	A vector of the indices of the columns to show in the plot. The indices are those of indices of the columns in the data frame qDataBefore in the density plot.
group2Color	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

## Value

A density plot

## Author(s)

Alexia Dorffer

## See Also

[wrapper.boxPlotD](#), [wrapper.varianceDistD](#)

## Examples

```
data(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
wrapper.densityPlotD(UPSep25, labels)
```

---

wrapper.diffAnaLimma *Performs differential analysis on an MSnSet object, calling the limma package functions*

---

### Description

Method to perform differential analysis on a [MSnSet](#) object (calls the limma package function).

### Usage

```
wrapper.diffAnaLimma(obj, condition1, condition2)
```

### Arguments

obj                    An object of class [MSnSet](#).  
condition1            A vector that contains the names of the conditions considered as condition 1.  
condition2            A vector that contains the names of the conditions considered as condition 2.

### Value

A dataframe as returned by the limma package

### Author(s)

Alexia Dorffer

### Examples

```
data(UPSep25)  
condition1 <- '25fmol'  
condition2 <- '10fmol'  
wrapper.diffAnaLimma(UPSep25, condition1, condition2)
```

---

wrapper.diffAnaWelch *Performs a differential analysis on a MSnSet object using the Welch t-test*

---

### Description

Computes differential analysis on a [MSnSet](#) object, using the Welch t-test (`t.test{stats}`).

### Usage

```
wrapper.diffAnaWelch(obj, condition1, condition2)
```

### Arguments

obj                    An object of class [MSnSet](#).  
condition1            A vector containing the names of the conditions considered as condition 1.  
condition2            A vector containing the names of the conditions considered as condition 2.

**Value**

A dataframe with two slots : P.Value (for the p-value) and logFC (the log of the Fold Change).

**Author(s)**

Alexia Dorffer

**Examples**

```
data(UPSep25)
condition1 <- '25fmol'
condition2 <- '10fmol'
wrapper.diffAnaWelch(UPSep25, condition1, condition2)
```

---

wrapper.heatmapD

*This function is a wrapper to [heatmap.2](#) that displays quantitative data in the `exprs()` table of an object of class `MSnSet`*

---

**Description**

Builds a heatmap of the quantitative proteomic data of a `MSnSet` object.

**Usage**

```
wrapper.heatmapD(obj, distance = "euclidean", cluster = "average",
  dendro = FALSE)
```

**Arguments**

obj	An object of class <code>MSnSet</code> .
distance	The distance used by the clustering algorithm to compute the dendrogram. See <code>help(heatmap.2)</code> .
cluster	the clustering algorithm used to build the dendrogram. See <code>help(heatmap.2)</code>
dendro	A boolean to indicate if the dendrogram has to be displayed

**Value**

A heatmap

**Author(s)**

Alexia Dorffer

**Examples**

```
data(testWithoutNA)
wrapper.heatmapD(testWithoutNA)
```

---

wrapper.mvHisto      *Histogram of missing values from a MSnSet object*

---

### Description

This method plots from a [MSnSet](#) object a histogram of missing values.

### Usage

```
wrapper.mvHisto(obj, indLegend = "auto", showValues = FALSE)
```

### Arguments

`obj`                    An object of class [MSnSet](#).  
`indLegend`            The indices of the column name's in `pData()` tab.  
`showValues`           A logical that indicates wether numeric values should be drawn above the bars.

### Value

A histogram

### Author(s)

Alexia Dorffer

### Examples

```
data(UPSsep25)
wrapper.mvHisto(UPSsep25, showValues=TRUE)
```

---

wrapper.mvImage      *Heatmap of missing values from a MSnSet object*

---

### Description

Plots a heatmap of the quantitative data. Each column represent one of the conditions in the object of class [MSnSet](#) and the color is proportional to the mean of intensity for each line of the dataset. The lines have been sorted in order to visualize easily the different number of missing values. A white square is plotted for missing values.

### Usage

```
wrapper.mvImage(obj)
```

### Arguments

`obj`                    An object of class [MSnSet](#).

### Value

A heatmap

**Author(s)**

Alexia Dorffer

**Examples**

```
data(UPSsep25)
wrapper.mvImage(UPSsep25)
```

---

wrapper.mvImputation *Missing values imputation from a [MSnSet](#) object*

---

**Description**

This method is a wrapper to the `imputeLCMD` package adapted to objects of class [MSnSet](#).

**Usage**

```
wrapper.mvImputation(obj, method)
```

**Arguments**

obj	An object of class <a href="#">MSnSet</a> .
method	The imputation method to be used. Choices are QRILC, KNN, BPCA and MLE.

**Value**

The object `obj` which has been imputed

**Author(s)**

Alexia Dorffer

**Examples**

```
data(UPSsep25)
wrapper.mvImputation(UPSsep25, "QRILC")
```

---

wrapper.mvPerLinesHisto  
*Histogram of missing values per lines from an object [MSnSet](#)*

---

**Description**

This method is a wrapper to plots from a [MSnSet](#) object a histogram which represents the distribution of the number of missing values (NA) per lines (ie proteins).

**Usage**

```
wrapper.mvPerLinesHisto(obj, indLegend = "auto", showValues = FALSE)
```

**Arguments**

obj	An object of class <a href="#">MSnSet</a> .
indLegend	The indice of the column name's in pData() tab .
showValues	A logical that indicates wether numeric values should be drawn above the bars.

**Value**

A histogram

**Author(s)**

Alexia Dorffer

**Examples**

```
data(UPSsep25)
wrapper.mvPerLinesHisto(UPSsep25)
```

---

wrapper.mvPerLinesHistoPerCondition

*Bar plot of missing values per lines and per conditions from an object*  
[MSnSet](#)

---

**Description**

This method is a wrapper to plots from a [MSnSet](#) object a bar plot which represents the distribution of the number of missing values (NA) per lines (ie proteins) and per conditions.

**Usage**

```
wrapper.mvPerLinesHistoPerCondition(obj, indLegend = "auto",
  showValues = FALSE)
```

**Arguments**

obj	An object of class <a href="#">MSnSet</a> .
indLegend	The indice of the column name's in pData() tab .
showValues	A logical that indicates wether numeric values should be drawn above the bars.

**Value**

A bar plot

**Author(s)**

Samuel Wieczorek

**Examples**

```
data(UPSsep25)
wrapper.mvPerLinesHistoPerCondition(UPSsep25)
```

---

wrapper.mvTypePlot      *Distribution of missing values with respect to intensity values from a MSnSet object*

---

### Description

This method plots a scatter plot which represents the distribution of missing values. The colors correspond to the different conditions (slot Label in in the dataset of class [MSnSet](#)). The x-axis represent the mean of intensity for one condition and one entity in the dataset (i. e. a protein) whereas the y-axis count the number of missing values for this entity and the considered condition. The data have been jittered for an easier vizualisation.

### Usage

```
wrapper.mvTypePlot(obj, threshold = 0)
```

### Arguments

obj                      An object of class [MSnSet](#).  
threshold                An integer for the intensity that delimits MNAR and MCAR missing values.

### Value

A scatter plot

### Author(s)

Florence Combes, Samuel Wiczorek

### Examples

```
data(UPSep25)  
wrapper.mvTypePlot(UPSep25)
```

---

wrapper.normalized      *Normalisation*

---

### Description

Provides several methods to normalize quantitative data from a [MSnSet](#) object. They are organized in four main families : Strong Rescaling, Median Centering, Mean Centering, Mean CenteringScaling. For the first family, two sub-categories are available : the sum by columns and the quantiles method. For the three other families, two categories are available : "Overall" which means that the value for each protein (ie line in the expression data tab) is computed over all the samples ; "within conditions" which means that the value for each protein (ie line in the exprs() data tab) is computed condition by condition.

### Usage

```
wrapper.normalized(obj, family, method)
```

**Arguments**

obj	An object of class <a href="#">MSnSet</a> .
family	One of the following : Global Rescaling, Median Centering, Mean Centering, Mean Centering Scaling.
method	"Overall" or "within conditions".

**Value**

An instance of class [MSnSet](#) where the quantitative data in the `exprs()` tab has been normalized.

**Author(s)**

Alexia Dorffer

**Examples**

```
data(UPSep25)
wrapper.normalized(UPSep25, "Median Centering", "within conditions")
```

---

wrapper.varianceDistD *Distribution of variance of proteins*

---

**Description**

Builds a densityplot of the variance of entities in the `exprs()` table of an object [MSnSet](#). The variance is calculated for each condition (Label) present in the dataset (see the slot 'Label' in the `pData()` table).

**Usage**

```
wrapper.varianceDistD(obj)
```

**Arguments**

obj	An object of class <a href="#">MSnSet</a> .
-----	---------------------------------------------

**Value**

A density plot

**Author(s)**

Alexia Dorffer

**See Also**

[wrapper.densityPlotD](#)

**Examples**

```
data(UPSep25)
wrapper.varianceDistD(UPSep25)
```



---

wrapper.violinPlotD    *Wrapper to the violinPlotD function on an object MSnSet*

---

### Description

This function is a wrapper for using the violinPlotD function with objects of class [MSnSet](#)

### Usage

```
wrapper.violinPlotD(obj, dataForXAxis = "Label", group2Color = "Condition")
```

### Arguments

obj	An object of class <a href="#">MSnSet</a> .
dataForXAxis	A vector of strings containing the names of columns in <code>pData()</code> to print labels on X-axis (Default is "Label").
group2Color	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

### Value

A violin plot

### Author(s)

Samuel Wieczorek

### See Also

[wrapper.densityPlotD](#), [wrapper.boxPlotD](#)

### Examples

```
data(UPSsep25)
library(vioplot)
types <- c("Label", "Analyt.Rep")
wrapper.violinPlotD(UPSsep25, types)
```

---

wrapperCalibrationPlot

*Performs a calibration plot on an [MSnSet](#) object, calling the cp4p package functions.*

---

### Description

This function is a wrapper to the calibration.plot method of the cp4p package for use with [MSnSet](#) objects.

**Usage**

```
wrapperCalibrationPlot(vPVal, pi0Method = "pounds")
```

**Arguments**

vPVal            A dataframe that contains quantitative data.  
pi0Method        A vector of the conditions (labels) (one label per sample).

**Value**

A plot

**Author(s)**

Samuel Wieczorek

**Examples**

```
data(UPSsep25)
condition1 <- '25fmol'
condition2 <- '10fmol'
qData <- Biobase::exprs(UPSsep25)
labels <- Biobase::pData(UPSsep25)[,"Label"]
diffAnaWelch(qData, labels, condition1, condition2)
```

---

writeMSnsetToExcel        *This function exports a [MSnSet](#) object to a Excel file.*

---

**Description**

This function exports a [MSnSet](#) data object to a Excel file. Each of the three data.frames in the [MSnSet](#) object (ie experimental data, phenoData and metaData are respectively integrated into separate sheets in the Excel file).

**Usage**

```
writeMSnsetToExcel(obj, filename)
```

**Arguments**

obj            An object of class [MSnSet](#).  
filename        A character string for the name of the Excel file.

**Value**

A Excel file (.xlsx)

**Author(s)**

Samuel Wieczorek

**Examples**

```
data(UPSpep25)  
writeMSnsetToExcel(UPSpep25, "foo")
```

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