

Package ‘Harman’

October 12, 2021

Type Package

Title The removal of batch effects from datasets using a PCA and constrained optimisation based technique

Version 1.20.0

Date 2016-03-31

Description Harman is a PCA and constrained optimisation based technique that maximises the removal of batch effects from datasets, with the constraint that the probability of overcorrection (i.e. removing genuine biological signal along with batch noise) is kept to a fraction which is set by the end-user.

NeedsCompilation yes

Suggests HarmanData, BiocGenerics, BiocStyle, knitr, rmarkdown, RUnit, RColorBrewer, bladderbatch, limma, minfi, lumi, msmsEDA, affydata, minfiData, sva

Depends R (>= 3.6)

Imports Rcpp (>= 0.11.2), graphics, stats, methods

LinkingTo Rcpp

License GPL-3 + file LICENCE

LazyData true

biocViews BatchEffect, Microarray, MultipleComparison, PrincipalComponent, Normalization, Preprocessing, DNAMethylation, Transcription, Software, StatisticalMethod

VignetteBuilder knitr

URL <http://www.bioinformatics.csiro.au/harman/>

BugReports <https://github.com/JasonR055/Harman/issues>

RoxygenNote 7.0.2

Encoding UTF-8

Author Josh Bowden [aut], Jason Ross [aut, cre], Yalchin Oytam [aut]

Maintainer Jason Ross <jason.ross@csiro.au>

git_url <https://git.bioconductor.org/packages/Harman>

git_branch RELEASE_3_13

git_last_commit 44013b7

git_last_commit_date 2021-05-19

Date/Publication 2021-10-12

R topics documented:

arrowPlot	2
callHarman	3
detachHarman	4
harman	5
harmanresults	6
harmanScores	7
pcaPlot	8
plot.harmanresults	9
prcompPlot	10
print.summary.harmanresults	11
reconstructData	11
shiftBetas	12
summary.harmanresults	13
Index	14

arrowPlot	<i>PCA before and after arrow plot for harman results</i>
-----------	---

Description

Generates an arrow plot for an instance of `harmanresults`. The tail of the arrow is the starting point (original) in principle coordinates, while the arrow head is the new point (corrected) in principle coordinates. It can be observed that on principle components that have undergone correction (`codeharmanresults$stats$correction < 1.0`), the samples within a batch will be coordinately moved towards 0 on that principle component.

Usage

```
arrowPlot(harmanresults, pc_x = 1, pc_y = 2, colBy = "batch",
  palette = "rainbow", col, length = 0.1, legend = TRUE, ...)
```

Arguments

harmanresults	an instance of harmanresults.
pc_x	integer, principle component for the plot x dimension.
pc_y	integer, principle component for the plot y dimension.
colBy	string, colour the points by the experimental or batch variable; legal values are expt and batch. The palette function specified in palette is used. This parameter is overridden by col.
palette	string, the function to call to create a vector of contiguous colours with the levels of factor in colBy steps.
col,	colour vector for the points. This parameter overrides palette.
length	length of the arrow heads, default is 0.1.
legend	logical, whether to display a legend on the plot
...	further arguments passed to or from other methods.

Details

Generates a Principle Component plot for an instance of harmanresults. If a vector of colours is supplied via the col argument, then a legend will not be drawn.

Value

None

See Also

[harmanresults](#) [plot.harmanresults](#)

Examples

```
library(HarmanData)
data(OLF)
expt <- olf.info$Treatment
batch <- olf.info$Batch
olf.harman <- harman(olf.data, expt, batch)
arrowPlot(olf.harman, pc_x=2, pc_y=3, length=0.2)
```

callHarman

Wrapper function to call the shared C/C++ library code

Description

This wrapper should probably not be addressed directly except for debugging. Instead use [harman](#). Input of PCA scores and the experiment structure (treatments and batches) and returns a batch corrected version of the PCA scores matrix

Usage

```
.callHarman(pc_data_scores, group, limit, numrepeats, randseed, forceRand,
            printInfo)
```

Arguments

pc_data_scores	2D NumericMatrix of PCA scores data (from the prcomp\$x slot), rows = samples, cols = PC scores
group	The structure of the experiment, consisting of batch numbers and treatment numbers forming 2 rows or columns (HarmanMain works out which). Each entry for a sample describes what batch it came from and what treatment it was given. Has to be integer formatted data.
limit	A double precision value indicating the limit of confidence in which to stop removing a batch effect
numrepeats	The number of repeats in which to run the simulated batch mean distribution estimator. Probably should be greater than 100,000.
randseed	Random seed to pass to the random number generator (0 for use default from system time)
forceRand	Force algorithm
printInfo	Print update information to screen

Value

SEXP R list: scores.corrected = harman_res_list["corrected_scores"]
 correction = harman_res_list["correction"]
 confidence = harman_res_list["confidence"]

Note

A data matrix with samples in columns must be transposed before PCA analysis and these scores in turn are tweaked a little before handing over to .callHarman. See the example below.

 detachHarman

Detach the Harman package and its shared C/C++ library code

Description

A helper function that can be called if [harman](#) had to be aborted.

Usage

```
detachHarman()
```

Value

None

harman

*Harman batch correction method***Description**

Harman is a PCA and constrained optimisation based technique that maximises the removal of batch effects from datasets, with the constraint that the probability of overcorrection (i.e. removing genuine biological signal along with batch noise) is kept to a fraction which is set by the end-user (Oytam et al, 2016; <http://dx.doi.org/10.1186/s12859-016-1212-5>).

Harman expects unbounded data, so for example, with HumanMethylation450 arrays do not use the Beta statistic (with values constrained between 0 and 1), instead use the logit transformed M-values.

Usage

```
harman(datamatrix, expt, batch, limit = 0.95, numrepeats = 100000L,
       randseed, forceRand = FALSE, printInfo = FALSE)
```

Arguments

<code>datamatrix</code>	matrix or data.frame, the data values to correct with samples in columns and data values in rows. Internally, a data.frame will be coerced to a matrix. Matrices need to be of type integer or double.
<code>expt</code>	vector or factor with the experimental variable of interest (variance to be kept).
<code>batch</code>	vector or factor with the batch variable (variance to be removed).
<code>limit</code>	numeric, confidence limit. Indicates the limit of confidence in which to stop removing a batch effect. Must be between 0 and 1.
<code>numrepeats</code>	integer, the number of repeats in which to run the simulated batch mean distribution estimator using the random selection algorithm. (N.B. 32 bit Windows versions may have an upper limit of 300000 before catastrophic failure)
<code>randseed</code>	integer, the seed for random number generation.
<code>forceRand</code>	logical, to enforce Harman to use a random selection algorithm to compute corrections. Force the simulated mean code to use random selection of scores to create the simulated batch mean (rather than full explicit calculation from all permutations).
<code>printInfo</code>	logical, whether to print information during computation or not.

Details

The `datamatrix` needs to be of type integer or numeric, or alternatively a data.frame that can be coerced into one using `as.matrix`. The matrix is to be constructed with data values (typically microarray probes or sequencing counts) in rows and samples in columns, much like the ‘assayData’ slot in the canonical Bioconductor eSet object, or any object which inherits from it. The data should have normalisation and any other global adjustment for noise reduction (such as background correction) applied prior to using Harman. For converge, the number of simulations, `numrepeats` parameter should probably should be at least 100,000. The underlying principle of Harman rests upon PCA, which is a parametric technique. This implies Harman should be optimal when the data is normally distributed. However, PCA is known to be rather robust to very non-normal data.

Value

A [harmanresults](#) S3 object.

References

Oytam et al (2016) BMC Bioinformatics 17:1. DOI: 10.1186/s12859-016-1212-5

See Also

[harman](#), [reconstructData](#), [pcaPlot](#), [arrowPlot](#)

Examples

```
library(HarmanData)
data(OLF)
expt <- olf.info$Treatment
batch <- olf.info$Batch
olf.harman <- harman(olf.data, expt, batch)
plot(olf.harman)
olf.data.corrected <- reconstructData(olf.harman)

## Reading from a csv file
datafile <- system.file("extdata", "NPM_data_first_1000_rows.csv.gz",
  package="Harman")
infofile <- system.file("extdata", "NPM_info.csv.gz", package="Harman")
datamatrix <- read.table(datafile, header=TRUE, sep=",", row.names="probeID")
batches <- read.table(infofile, header=TRUE, sep=",", row.names="Sample")
res <- harman(datamatrix, expt=batches$Treatment, batch=batches$Batch)
arrowPlot(res, 1, 3)
```

harmanresults

Harman results object

Description

The S3 object returned after running [harman](#).

Details

[harmanresults](#) is the S3 object used to store the results from [harman](#). This object may be presented to summary and data exploration functions such as [plot.harmanresults](#) and [summary.harmanresults](#) as well as the [reconstructData](#) function which creates a corrected matrix of data with the batch effect removed.

Slots

factors A data.frame of the expt and batch vectors.
parameters The harman runtime parameters. See [harman](#) for details.
stats Confidence intervals and the degree of correction for each principal component.
center The centering vector returned by [prcomp](#) with center=TRUE.
rotation The matrix of eigenvectors (by column) returned from [prcomp](#).
original The original PC scores returned by [prcomp](#).
corrected The harman corrected PC scores.

See Also

[harman](#), [reconstructData](#), [pcaPlot](#), [arrowPlot](#)

Examples

```

## HarmanResults
library(HarmanData)
data(OLF)
expt <- olf.info$Treatment
batch <- olf.info$Batch
olf.harman <- harman(as.matrix(olf.data), expt, batch)
plot(olf.harman)
summary(olf.harman)
pcaPlot(olf.harman, pc_x=2, pc_y=3)
pcaPlot(olf.harman, pc_x=2, pc_y=3, colBy='expt', pch=1)
olf.data.corrected <- reconstructData(olf.harman)

```

harmanScores

A Principal components prcomp function tweaked for Harman

Description

A tweaking of `stats::prcomp` such that for the svd, the transpose of `u` is used instead of `v` when the number of assays is less than the number of samples.

Usage

```
harmanScores(x)
```

Arguments

`x` matrix, data matrix of values to perform PCA on.

Value

scores, a `prcomp`-like object with rotation, scores and the center values. The scores are corrected, but all three are needed later to reconstruct the data.

pcaPlot

PCA plot for harman results

Description

Generates a Principle Component plot for an instance of [harmanresults](#).

Usage

```
pcaPlot(harmanresults, pc_x = 1, pc_y = 2, this = "corrected",  
        colBy = "batch", pchBy = "expt", palette = "rainbow", legend = TRUE,  
        col, pch, ...)
```

Arguments

harmanresults	An instance of harmanresults.
pc_x	integer, principle component for the plot x dimension.
pc_y	integer, principle component for the plot y dimension.
this	string, legal values are original or corrected.
colBy	string, colour the points by the experimental or batch variable; legal values are expt and batch. The palette function specified in palette is used. This parameter is overridden by col.
pchBy	string, point-type by the experimental or batch variable; legal values are expt and batch. This parameter is overridden by pch.
palette	string, the function to call to create a vector of contiguous colours with the levels of factor in colBy steps.
legend	logical, whether to display a legend on the plot.
col,	colour vector for the points. This parameter overrides colBy and palette.
pch,	integer vector giving the point type. This parameter overrides pchBy.
...	further arguments passed to or from other methods.

Details

If a vector of colours is supplied via the col argument, then a legend will not be drawn.

Value

None

See Also

[harmanresults plot.harmanresults](#)

Examples

```
library(HarmanData)
data(OLF)
expt <- olf.info$Treatment
batch <- olf.info$Batch
olf.harman <- harman(as.matrix(olf.data), expt, batch)
pcaPlot(olf.harman)
pcaPlot(olf.harman, colBy='expt')
pcaPlot(olf.harman, pc_x=2, pc_y=3, this='original', pch=17)
```

plot.harmanresults *Plot method for harman*

Description

Plot method for instances of [harmanresults](#).

Usage

```
## S3 method for class 'harmanresults'
plot(x, ...)
```

Arguments

x	An instance of harmanresults.
...	further plotting parameters.

Value

None

See Also

[harmanresults](#) [pcaPlot](#)

Examples

```
library(HarmanData)
data(OLF)
expt <- olf.info$Treatment
batch <- olf.info$Batch
olf.harman <- harman(olf.data, expt, batch)
plot(olf.harman)
```

prcompPlot

*PCA plot***Description**

Generates a Principle Component plot for data.frames, matrices, or a pre-made [prcomp](#) object.

Usage

```
prcompPlot(object, pc_x = 1, pc_y = 2, scale = FALSE, colFactor = NULL,
           pchFactor = NULL, palette = "rainbow", legend = TRUE, ...)
```

Arguments

object	data.frame, matrix or prcomp object.
pc_x	integer, principle component for the plot x dimension.
pc_y	integer, principle component for the plot y dimension.
scale	logical, whether to scale to unit variance before PCA.
colFactor	factor or vector, colour the points by this factor, default is NULL.
pchFactor	factor or vector, point-type by this factor, default is NULL.
palette	string, the function to call to create a vector of contiguous colours with <code>levels(colFactor)</code> steps.
legend	logical, whether to display a legend on the plot.
...	further arguments passed to or from other methods.

Details

A data.frame object will be coerced internally to a matrix. Matrices must be of type double or integer. The prcompPlot function will then perform a principle component analysis on the data prior to plotting. The function is call is `prcomp(t(object), retx=TRUE, center=TRUE, scale.=scale)`. Instead of specifying a data.frame or matrix, a pre-made prcomp object can be given to prcompPlot. In this case, care should be taken in setting the appropriate value of scale.. If a vector is given to colFactor or pchFactor, they will be coerced internally to factors.

For the default NULL values of colFactor and pchFactor, all colours will be black and circles the point type, respectively.

Value

None

See Also

[prcomp rainbow](#)

Examples

```
library(HarmanData)
data(IMR90)
expt <- imr90.info$Treatment
batch <- imr90.info$Batch
prcompPlot(imr90.data, colFactor=expt)
pca <- prcomp(t(imr90.data), scale.=TRUE)
prcompPlot(pca, 1, 3, colFactor=batch, pchFactor=expt, palette='topo.colors',
main='IMR90 PCA plot of Dim 1 and 3')
```

```
print.summary.harmanresults
```

Printing Harmanresults summaries.

Description

Print method for summary.harmanresults.

Usage

```
## S3 method for class 'summary.harmanresults'
print(x, ...)
```

Arguments

x an object of class summary.harmanresults, usually, a result of a call to summary.harmanresults.
... further parameters.

Value

Prints summary information from an object of class summary.harmanresults.

```
reconstructData
```

Reconstruct corrected data from Harman results

Description

Method which reverts the PCA factorisation for instances of [harmanresults](#). This allows the original or corrected data to be returned back from the PCA domain into the original data domain.

Usage

```
reconstructData(object, this = "corrected")
```

Arguments

object An instance of harmanresults.
this string, legal values are original or corrected.

Value

matrix of data

See Also

[harman harmanresults](#)

Examples

```
library(HarmanData)
data(OLF)
expt <- olf.info$Treatment
batch <- olf.info$Batch
olf.harman <- harman(olf.data, expt, batch)
olf.data.corrected <- reconstructData(olf.harman)
```

shiftBetas

Shift beta values from 0 and 1 to avoid infinite M values

Description

A convenience function for methylation data.

Usage

```
shiftBetas(betas, shiftBy = 1e-04)
```

Arguments

betas matrix, beta values.
shiftBy numeric, the amount to shift values of 0 and 1 by.

Value

None

Examples

```
betas <- seq(0, 1, by=0.05)
range(betas)
newBetas <- shiftBetas(betas, shiftBy=1e-4)
newBetas
range(newBetas)
```

summary.harmanresults *Summarizing harman results.*

Description

Summary method for class [harmanresults](#).

Usage

```
## S3 method for class 'harmanresults'  
summary(object, ...)
```

Arguments

object	An object of class harmanresults.
...	further parameters.

Value

Returns an object of class summary.harmanresults.

See Also

[harmanresults](#)

Examples

```
library(HarmanData)  
data(OLF)  
expt <- olf.info$Treatment  
batch <- olf.info$Batch  
olf.harman <- harman(olf.data, expt, batch)  
summary(olf.harman)
```

Index

`.callHarman` (`callHarman`), 3

`arrow`, 3
`arrowPlot`, 2, 6, 7
`as.matrix`, 5

`callHarman`, 3

`detachHarman`, 4

`harman`, 3, 4, 5, 6, 7, 12
`harmanresults`, 2, 3, 6, 6, 8, 9, 11–13
`harmanScores`, 7

`pcaPlot`, 6, 7, 8, 9
`plot.harmanresults`, 3, 6, 8, 9
`prcomp`, 7, 10
`prcompPlot`, 10
`print.summary.harmanresults`, 11

`rainbow`, 10
`reconstructData`, 6, 7, 11

`shiftBetas`, 12
`summary.harmanresults`, 6, 13