

# Package ‘MethCP’

October 17, 2020

**Type** Package

**Title** Differential methylation analysis for bisulfite sequencing data

**Version** 1.2.0

**Description** MethCP is a differentially methylated region (DMR) detecting method for whole-genome bisulfite sequencing (WGBS) data, which is applicable for a wide range of experimental designs beyond the two-group comparisons, such as time-course data. MethCP identifies DMRs based on change point detection, which naturally segments the genome and provides region-level differential analysis.

**License** Artistic-2.0

**Encoding** UTF-8

**LazyData** TRUE

**RoxygenNote** 7.1.0

**biocViews** DifferentialMethylation, Sequencing, WholeGenome, TimeCourse

**Imports** methods, utils, stats, S4Vectors, bsseq, DSS, methylKit, DNACopy, GenomicRanges, IRanges, GenomeInfoDb, BiocParallel

**Suggests** testthat, knitr, rmarkdown

**VignetteBuilder** knitr

**Depends** R (>= 3.6.0)

**BugReports** <https://github.com/boyinggong/methcp/issues>

**git\_url** <https://git.bioconductor.org/packages/MethCP>

**git\_branch** RELEASE\_3\_11

**git\_last\_commit** 41cf2b6

**git\_last\_commit\_date** 2020-04-27

**Date/Publication** 2020-10-16

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## R topics documented:

calcLociStat . . . . .	2
calcLociStatTimeCourse . . . . .	3
createBsseqObject . . . . .	5
getSigRegion . . . . .	6
MethCP-class . . . . .	7
MethCPFromStat . . . . .	9
segmentMethCP . . . . .	10
show,MethCP-method . . . . .	12
<b>Index</b>	<b>13</b>

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calcLociStat	<i>Calculate the per-cytosine statistics for simple two-group comparisons.</i>
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### Description

calcLociStat calculates per-cytosine based statistics between two population groups.

### Usage

```
calcLociStat(
  bs.object, group1, group2, test = c("DSS", "methylKit"),
  BPPARAM = bpparam())
```

### Arguments

bs.object	a BSseq object from the bsseq package.
group1	a character vector containing the sample names of the treatment group.
group2	a character vector containing the sample names of the control group.
test	a character string containing the names of the test to be performed per cytosine.
BPPARAM	An optional BiocParallelParam instance determining the parallel back-end to be used during evaluation, or a list of BiocParallelParam instances, to be applied in sequence for nested calls to BiocParallel functions. Default bpparam().

### Details

For each cytosine, calcLociStat calculates a statistics using either package DSS or methylKit to test the differences between two groups, and returns a MethCP object. For customized per-cytosine statistics, please use the function methcpFromStat. The input bs.object is a BSseq object from the bsseq package which contains the raw data including coverages, methylated counts and position information for every cytosine in the dataset.

### Value

a MethCP object that is not segmented.

**Examples**

```

library(bsseq)
library(GenomicRanges)
library(IRanges)

set.seed(0286374)

# Simulate a small dataset with 11 cytosine and 6 samples,
# 3 in the treatment group and 3 in the control group. The
# methylation ratios are generated using Binomial distribution
# with probability 0.3.
nC <- 2000
sim_cov <- rbinom(6*nC, 5, 0.5) + 5
sim_M <- vapply(
  sim_cov, function(x) rbinom(1, x, 0.3),
  FUN.VALUE = numeric(1))
sim_cov <- matrix(sim_cov, ncol = 6)
sim_M <- matrix(sim_M, ncol = 6)
# methylation ratios in the DMRs in the treatment group are
# generated using Binomial(0.7)
DMRs <- c(600:622, 1089:1103, 1698:1750)
sim_M[DMRs, 1:3] <- vapply(
  sim_cov[DMRs, 1:3], function(x) rbinom(1, x, 0.7),
  FUN.VALUE = numeric(1))
# sample names
sample_names <- c(paste0("treatment", 1:3), paste0("control", 1:3))
colnames(sim_cov) <- sample_names
colnames(sim_M) <- sample_names

# create a bs.object
bs_object <- BSseq(gr = GRanges(
  seqnames = "Chr01", IRanges(
    start = (1:nC)*10, width = 1)),
  Cov = sim_cov, M = sim_M, sampleNames = sample_names)
# methcp_obj1 <- calcLociStat(
#   bs_object,
#   group1 = paste0("treatment", 1:3),
#   group2 = paste0("control", 1:3),
#   test = "DSS")
methcp_obj2 <- calcLociStat(
  bs_object,
  group1 = paste0("treatment", 1:3),
  group2 = paste0("control", 1:3),
  test = "methylKit")

```

---

calcLociStatTimeCourse

*Calculate the per-cytosine statistics for time-course data.*

---

**Description**

For each cytosine, calcLociStatTimeCourse fits a linear model on the arcsin-transformed methylation ratios, and test the differences of the slope between the treatment and the control group.

**Usage**

```
calcLociStatTimeCourse(
  bs.object, meta, force.slope = FALSE,
  BPPARAM = bpparam())
```

**Arguments**

bs.object	a BSseq object from the bsseq package.
meta	a data.frame. See details.
force.slope	if TRUE, we force the slope in the linear model to be the same between two conditions. Otherwise, the slopes are fitted separately but not tested.
BPPARAM	An optional BiocParallelParam instance determining the parallel back-end to be used during evaluation, or a list of BiocParallelParam instances, to be applied in sequence for nested calls to BiocParallel functions. Default bpparam().

**Details**

bs.object is a BSseq object from the bsseq package, which contains the raw data including covers, methylated counts and position information for every cytosine in the dataset. meta must contain columns named Condition, Time and SampleName in the dataframe. They are used to fit the linear model.

**Value**

A MethCP object that is not segmented.

**Examples**

```
library(bsseq)
# Simulate a small dataset with 2000 cyotsine and 10 samples,
# 5 in the treatment group and 5 in the control group. The
# methylation ratio are generated using Binomial distribution
# with probability 0.3, 0.4, 0.5, 0.6 and 0.7 for 5 time points.
nC <- 2000
nsamples <- 5
sim_cov <- rnbinom(10*nC, 5, 0.5) + 5
sim_cov <- matrix(sim_cov, ncol = 10)
time_point <- rep(1:nsamples, 2)
ratios <- time_point/10 + 0.2
sim_M <- sapply(1:(2*nsamples), function(i){
  sapply(sim_cov[, i], function(j) rbinom(1, j, ratios[i]))
})
sim_M <- matrix(sim_M, ncol = 2*nsamples)
# methylation ratios in the DMRs in the treatment group are
# generated using Binomial(0.3)
DMRs <- c(600:622, 1089:1103, 1698:1750)
sim_M[DMRs, 1:5] <- sapply(
  sim_cov[DMRs, 1:5], function(x) rbinom(1, x, 0.3))
# sample names
sample_names <- c(paste0("treatment", 1:nsamples),
  paste0("control", 1:nsamples))
colnames(sim_cov) <- sample_names
colnames(sim_M) <- sample_names
```

```

# create a bs.object
bs_object_ts <- BSseq(gr = GRanges(
  seqnames = "Chr01", IRanges(
    start = (1:nC)*10, width = 1)),
  Cov = sim_cov, M = sim_M, sampleNames = sample_names)
DMRs_pos_ts <- DMRs*10
meta <- data.frame(
  Condition = rep(
    c("treatment", "control"),
    each = nsamples),
  SampleName = sample_names,
  Time = time_point)
obj_ts <- calcLociStatTimeCourse(bs_object_ts, meta)
obj_ts

```

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createBsseqObject      *Helper function to read text files and create a bsseq object.*

---

## Description

Create a bsseq object when the data for each sample is stored in a separate text file.

## Usage

```

createBsseqObject(
  files, sample_names,
  chr_col, pos_col, m_col, cov_col, header = TRUE)

```

## Arguments

files	a character vector of file names with full path to the file.
sample_names	a character vector of sample names. It should have the same length as files vector.
chr_col	name or index of the chromosome column in data files.
pos_col	name or index of the position column in data files.
m_col	name or index of the methylated counts column.
cov_col	name or index of the coverage counts column.
header	a logical value indicating whether the file contains the names of the variables as its first line. default TRUE.

## Value

a MethCP object that is not segmented.

**Examples**

```

library(bsseq)
# The dataset is consist of 6 samples. 3 samples are H2A.Z mutant
# plants, and 3 samples are controls.
sample_names <- c(
  paste0("control", seq_len(3)),
  paste0("treatment", seq_len(3))
)

# Get the vector of file path and names
raw_files <- system.file(
  "extdata", paste0(sample_names, ".txt"), package = "MethCP")

# load the data
bs_object <- createBsseqObject(
  files = raw_files, sample_names = sample_names,
  chr_col = 'Chr', pos_col = 'Pos', m_col = "M", cov_col = 'Cov')

```

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<code>getSigRegion</code>	<i>Obtain the significant DMRs.</i>
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**Description**

`getSigRegion` returns the significant DMRs giving the segmented MethCP object.

**Usage**

```

getSigRegion(
  object, sig.level = 0.01, mean.coverage = 1,
  mean.diff = 0.1, nC.valid = 10)

```

**Arguments**

<code>object</code>	a MethCP object that is segmented using function <code>segmentMethCP</code> .
<code>sig.level</code>	significance level to call a region DMR.
<code>mean.coverage</code>	The minimum average coverage required for the reported DMRs.
<code>mean.diff</code>	The minimum differences between groups required for the reported DMRs.
<code>nC.valid</code>	number of valid cytosines required for the reported DMRs.

**Value**

a data.frame containing the DMRs.

**Examples**

```

library(bsseq)
# Simulate a small dataset with 2000 cyotsine and 6 samples,
# 3 in the treatment group and 3 in the control group. The
# methylation ratio are generated using Binomial distribution
# with probability 0.3.
nC <- 2000

```

```

sim_cov <- rnbinom(6*nC, 5, 0.5) + 5
sim_M <- vapply(
  sim_cov, function(x) rbinom(1, x, 0.3), FUN.VALUE = numeric(1))
sim_cov <- matrix(sim_cov, ncol = 6)
sim_M <- matrix(sim_M, ncol = 6)
# methylation ratios in the DMRs in the treatment group are
# generated using Binomial(0.7)
DMRs <- c(600:622, 1089:1103, 1698:1750)
sim_M[DMRs, 1:3] <- vapply(
  sim_cov[DMRs, 1:3], function(x) rbinom(1, x, 0.7),
  FUN.VALUE = numeric(1))
# sample names
sample_names <- c(paste0("treatment", 1:3), paste0("control", 1:3))
colnames(sim_cov) <- sample_names
colnames(sim_M) <- sample_names

# create a bs.object
bs_object <- BSseq(gr = GRanges(
  seqnames = "Chr01",
  IRanges(start = (1:nC)*10, width = 1)),
  Cov = sim_cov, M = sim_M, sampleNames = sample_names)
DMRs_pos <- DMRs*10
methcp_obj1 <- calcLociStat(
  bs_object,
  group1 = paste0("treatment", 1:3),
  group2 = paste0("control", 1:3),
  test = "methylKit")
methcp_obj1 <- segmentMethCP(
  methcp_obj1, bs_object,
  region.test = "fisher")
methcp_res1 <- getSigRegion(methcp_obj1)

```

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MethCP-class

*Class* MethCP

---

### Description

A class for performing DMR (differentially methylated region) detection analysis using method MethCP on whole genome bisulfite sequencing data.

The constructor function for MethCP objects.

### Usage

```

MethCP(
  test = NA_character_,
  group1 = NA,
  group2 = NA,
  chr,
  pos,
  pvals,
  effect.size
)

```

```
MethCP(
  test = NA_character_, group1 = NA, group2 = NA, chr, pos,
  pvals, effect.size)
```

### Arguments

<code>test</code>	a character string of the name of the per-cytosine statistics.
<code>group1</code>	a character vector containing the sample names of the treatment group.
<code>group2</code>	a character vector containing the sample names of the control group.
<code>chr</code>	a character vector containing the cytosine chromosome information.
<code>pos</code>	a numeric vector containing the cytosine positions.
<code>pvals</code>	a numeric vector containing the p-values for each cytosine.
<code>effect.size</code>	a numeric vector containing the effect sizes for each cytosine.

### Details

If not specified by function `calcLociStatTimeCourse`, `calcLociStat`, the parameter `test` can be set to any user-specified string indicating the name of the test performed.

In the cases where the goal is not to compare between treatment and control groups, parameter `group1` and `group2` can be set to `NA`.

If generated by `calcLociStat`, parameter `stat` will be a `GRangesList` object where each element in the list contains statistics for each of the chromosome in the dataset.

### Value

A `MethCP` objects.

### Slots

<code>test</code>	a character string of the name of the per-cytosine statistics.
<code>group1</code>	a character vector containing the sample names of the treatment group.
<code>group2</code>	a character vector containing the sample names of the control group.
<code>stat</code>	a <code>GRangesList</code> object containing the results of per-cytosine tests.
<code>segmentation</code>	a <code>GRanges</code> object containing the segments and their information such as region-based statistics, coverages, etc.

### Examples

```
obj <- MethCP(
  test = "myTest",
  group1 = paste0("Treatment", 1:3),
  group2 = paste0("Control", 1:3),
  chr = rep("Chr01", 5),
  pvals = c(0, 0.1, 0.9, NA, 0.02),
  pos = c(2, 5, 9, 10, 18),
  effect.size = c(1, -1, NA, 9, Inf))
# MethCP will omit the NAs and infinite values.
obj
```



MethCPFromStat

*Create a MethCP object given the per-cytosine based tests.***Description**

Given the per-cytosine based p-values and effect sizes, MethCPFromStat create a MethCP object for segmentation.

**Usage**

```
MethCPFromStat(
  data,
  test.name,
  pvals.field = "pvals",
  effect.size.field = "effect.size",
  seqnames.field = c("seqnames", "seqname", "chromosome", "chrom", "chr",
    "chromosome_name", "seqid"),
  pos.field = "pos"
)
```

**Arguments**

<code>data</code>	a data.frame or GPos or GRanges object.
<code>test.name</code>	a character string containing the name of the test to be performed per cytosine.
<code>pvals.field</code>	A character vector of recognized names for the column (if 'data' is a data.frame) or meta data column (is 'data' is GPos or GRanges object) in 'data' that contains the p-value.
<code>effect.size.field</code>	A character vector of recognized names for the column (if 'data' is a data.frame) or meta data column (is 'data' is GPos or GRanges object) in 'data' that contains the effect size.
<code>seqnames.field</code>	A character vector of recognized names for the column in 'data' that contains the chromosome name (sequence name) associated with each position. Only the first name in seqnames.field that is found in colnames(data) is used. If no one is found, then an error is raised. This column is only used when 'data' is a data.frame. Otherwise, chromosome name is obtained from GPos or GRanges object.
<code>pos.field</code>	A character vector of recognized names for the column in df that contains the position integer associated with each position. Only the first name in pos.field that is found in colnames(data) is used. If no one is found, then an error is raised. This column is only used when 'data' is a data.frame. Otherwise, position is obtained from GPos or GRanges object.

**Value**

a MethCP object that is not segmented.

**Examples**

```
# ===== construct using data frame
data <- data.frame(
  chr = rep("Chr01", 5),
  pos = c(2, 5, 9, 10, 18),
  effect.size = c(1,-1, NA, 9, Inf),
  pvals = c(0, 0.1, 0.9, NA, 0.02))
obj <- MethCPFromStat(
  data, test.name="myTest",
  pvals.field = "pvals",
  effect.size.field="effect.size",
  seqnames.field="chr",
  pos.field="pos"
)
# ===== construct using GRanges
library(GenomicRanges)
data <- GRanges(
  "Chr01", IRanges(c(2, 5, 9, 10, 18), c(2, 5, 9, 10, 18)),
  pvals=c(0, 0.1, 0.9, NA, 0.02), effect.size = c(1,-1, NA, 9, Inf))
obj <- MethCPFromStat(
  data, test.name="myTest",
  pvals.field = "pvals",
  effect.size.field="effect.size"
)
```

segmentMethCP

*Perform segmentation on a MethCP object.***Description**

Perform CBS algorithm that segments the genome into similar levels of significance.

**Usage**

```
segmentMethCP(
  methcp.object, bs.object,
  region.test = c(
    "fisher", "stouffer", "weighted-variance", "weighted-coverage"),
  min.width = 2, sig.level = 0.01,
  presegment_dist = 600, BPPARAM = bpparam(), ...)
```

**Arguments**

methcp.object	a MethCP object.
bs.object	a BSseq object from the bsseq package.
region.test	The meta-analysis method used to create region-based test statistics.
min.width	the minimum width for the segments, which is used as termination rule for the segmentation algorithm.
sig.level	the significance level of the segments, which is used as termination rule for the segmentation algorithm.

presegment_dist	the maximum distance between cytosines for the presegmentation.
BPPARAM	An optional BiocParallelParam instance determining the parallel back-end to be used during evaluation, or a list of BiocParallelParam instances, to be applied in sequence for nested calls to BiocParallel functions. Default bpparam().
...	argument to be passed to segment function in DNAcopy package

## Details

The MethCP object `methcp.object` can be generated from functions `calcLociStat`, `calcLociStatTimeCourse`, or `methcpFromStat`.

If `region.test = "fisher"`, Fisher's combined probability test is used.

If `region.test = stouffer` Stouffer's test is applied.

If `region.test = "weighted-variance"` we use the variance of the test to combine per-cytosine based statistics into a region-based statistic.

If `region.test = "weighted-coverage"` we use the coverage of the test to combine per-cytosine based statistics into a region-based statistic.

## Value

a MethCP object that is not segmented.

## Examples

```
library(bsseq)
# Simulate a small dataset with 2000 cyotsine and 6 samples,
# 3 in the treatment group and 3 in the control group. The
# methylation ratio are generated using Binomial distribution
# with probability 0.3.
nC <- 2000
sim_cov <- rnbinom(6*nC, 5, 0.5) + 5
sim_M <- vapply(
  sim_cov, function(x) rbinom(1, x, 0.3), FUN.VALUE = numeric(1))
sim_cov <- matrix(sim_cov, ncol = 6)
sim_M <- matrix(sim_M, ncol = 6)
# methylation ratios in the DMRs in the treatment group are
# generated using Binomial(0.7)
DMRs <- c(600:622, 1089:1103, 1698:1750)
sim_M[DMRs, 1:3] <- vapply(
  sim_cov[DMRs, 1:3], function(x) rbinom(1, x, 0.7),
  FUN.VALUE = numeric(1))
# sample names
sample_names <- c(paste0("treatment", 1:3), paste0("control", 1:3))
colnames(sim_cov) <- sample_names
colnames(sim_M) <- sample_names

# create a bs.object
bs_object <- BSseq(gr = GRanges(
  seqnames = "Chr01", IRanges(start = (1:nC)*10, width = 1)),
  Cov = sim_cov, M = sim_M,
  sampleNames = sample_names)
DMRs_pos <- DMRs*10
methcp_obj1 <- calcLociStat(
  bs_object,
```

```
group1 = paste0("treatment", 1:3),
group2 = paste0("control", 1:3),
test = "methylKit")
methcp_obj1 <- segmentMethCP(
  methcp_obj1, bs_object,
  region.test = "fisher")
```

---

show,MethCP-method      *The show method*

---

### **Description**

Print MethCP object information.

### **Usage**

```
## S4 method for signature 'MethCP'
show(object)
```

### **Arguments**

object                  a MethCP object.

### **Value**

No value will be returned.

# Index

[calcLociStat](#), [2](#)  
[calcLociStatTimeCourse](#), [3](#)  
[createBsseqObject](#), [5](#)  
  
[getSigRegion](#), [6](#)  
  
[MethCP \(MethCP-class\)](#), [7](#)  
[MethCP-class](#), [7](#)  
[MethCPFromStat](#), [9](#)  
  
[segmentMethCP](#), [10](#)  
[show, MethCP-method](#), [12](#)