

# Package ‘gwascat’

April 15, 2020

**Title** representing and modeling data in the EMBL-EBI GWAS catalog

**Version** 2.18.0

**Author** VJ Carey <stvjc@channing.harvard.edu>

**Description** Represent and model data in the EMBL-EBI GWAS catalog.

**Enhances** SNPlocs.Hsapiens.dbSNP144.GRCh37

**Depends** R (>= 3.5.0), Homo.sapiens

**Imports** methods, BiocGenerics, S4Vectors (>= 0.9.25), IRanges,  
GenomeInfoDb, GenomicRanges (>= 1.29.6), GenomicFeatures,  
Biostrings, Rsamtools, rtracklayer, AnnotationDbi, utils

**Suggests** DO.db, DT, knitr, RBGL, RUnit, snpStats, Gviz,  
VariantAnnotation, AnnotationHub, gQTLstats, graph, ggbio,  
ggplot2, DelayedArray

**VignetteBuilder** utils, knitr

**Maintainer** VJ Carey <stvjc@channing.harvard.edu>

**License** Artistic-2.0

**LazyData** no

**biocViews** Genetics

**git\_url** <https://git.bioconductor.org/packages/gwascat>

**git\_branch** RELEASE\_3\_10

**git\_last\_commit** e9cf69e

**git\_last\_commit\_date** 2019-10-29

**Date/Publication** 2020-04-14

## R topics documented:

gwascat-package	2
bindcadd_snv	3
gwastagger	4
gwaswloc-class	5
gwcex2gviz	7
gwdf_2012_02_02	7
ldtagr	9
locon6	10
makeCurrentGwascat	11

obo2graphNEL . . . . .	12
riskyAlleleCount . . . . .	13
topTraits . . . . .	14
traitsManh . . . . .	15

<b>Index</b>	<b>16</b>
--------------	-----------

---

gwascat-package	<i>representing and modeling data in the NHGRI GWAS catalog</i>
-----------------	---

---

## Description

representing and modeling data in the NHGRI GWAS catalog, using GRanges and allied infrastructure

## Details

```

Package:    gwascat
Version:    1.7.3
Suggests:
Depends:    R (>= 3.0.0), methods, IRanges, GenomicRanges
Imports:
License:    Artistic-2.0
LazyLoad:  yes

```

Index:

```
gwaswloc-class      Class "gwaswloc"
```

The GWAS catalog management has migrated to EMBL/EBI. Use `data(ebicat38)` for an image dated 3 August 2015. Use `makeCurrentGwascat()` to get a more recent image. Use `data(ebicat37)` for a GRCh37 (or hg19) liftOver result. Use `data(ebicat37UCSC)` for an image with hg19 as genome tag and UCSC seqnames.

The data objects

```
'g17SM' 'gg17N' 'gw6.rs_17' 'low17' 'rules_6.0_1kg_17' 'gwrngs'
```

are described in vignettes.

The `DataFrame` function is imported from `IRanges`.

The `SnpMatrix-class` is used to represent data related to rule-based imputation, using the `impute.snps` function.

`si.hs.38` is a `Seqinfo-class` instance for hg38.

## Author(s)

VJ Carey <stvjc@channing.harvard.edu>

Maintainer: VJ Carey <stvjc@channing.harvard.edu>

## References

<http://www.ebi.ac.uk/gwas/>

Partial support from the Computational Biology Group at Genentech, Inc.

## Examples

```
data(ebicat38)
ebicat38
```

---

bindcadd_snv	<i>bind CADD scores of Kircher et al. 2014 to a GRanges instance</i>
--------------	--

---

## Description

bind CADD scores of Kircher et al. 2014 to a GRanges instance; by default will use HTTP access at UW

## Usage

```
bindcadd_snv(gr, fn = "http://krishna.gs.washington.edu/download/CADD/v1.0/1000G.tsv.gz")
```

## Arguments

gr	query ranges to which CADD scores should be bound
fn	path to Tabix-indexed bgzipped TSV of CADD as distributed at krishna.gs.washington.edu on 1 April 2014

## Details

joins CADD fields at addresses that match query; the CADD fields for query ranges that are not matched are set to NA

## Value

GRanges instance with additional fields as obtained in the CADD resource

## Note

This software developed in part with support from Genentech, Inc.

## Author(s)

VJ Carey <stvjc@channing.harvard.edu>

## References

M Kircher, DM Witten, P Jain, BJ O’Roak, GM Cooper, J Shendure, A general framework for estimating the relative pathogenicity of human genetic variants, Nature Genetics Feb 2014, PMID 24487276

**Examples**

```

if (interactive()) {
  data(ebicat37)
  g2 = as(ebicat37, "GRanges")
  bindcadd_snv( g2[which(seqnames(g2)=="chr2")][1:20] )
}

```

---

gwastagger

*data on 1000 genomes SNPs that 'tag' GWAS catalog entries*


---

**Description**

data on 1000 genomes SNPs that 'tag' GWAS catalog entries

**Usage**

```
data(gwastagger)
```

**Format**

The format is:

```

Formal class 'GRanges' [package "GenomicRanges"] with 6 slots
..@ seqnames :Formal class 'Rle' [package "IRanges"] with 4 slots
.. ..@ values : Factor w/ 24 levels "chr1","chr2",...: 1 2 3 4 5 6 7 8 9 10 ...
.. ..@ lengths : int [1:22] 24042 23740 21522 14258 14972 34101 12330 11400 8680 15429 ...
.. ..@ elementMetadata: NULL
.. ..@ metadata : list()
..@ ranges :Formal class 'IRanges' [package "IRanges"] with 6 slots
.. ..@ start : int [1:297579] 986111 988364 992250 992402 995669 999686 1005579 1007450
1011209 1011446 ...
.. ..@ width : int [1:297579] 1 1 1 1 1 1 1 1 1 1 ...
.. ..@ NAMES : NULL
.. ..@ elementType : chr "integer"
.. ..@ elementMetadata: NULL
.. ..@ metadata : list()
..@ strand :Formal class 'Rle' [package "IRanges"] with 4 slots
.. ..@ values : Factor w/ 3 levels "+","-","*": 3
.. ..@ lengths : int 297579
.. ..@ elementMetadata: NULL
.. ..@ metadata : list()
..@ elementMetadata:Formal class 'DataFrame' [package "IRanges"] with 6 slots
.. ..@ rownames : NULL
.. ..@ nrows : int 297579
.. ..@ listData :List of 3
.. .. ..$ tagid : chr [1:297579] "rs28479311" "rs3813193" "chr1:992250" "rs60442576" ...
.. .. ..$ R2 : num [1:297579] 0.938 0.994 0.969 1 1 ...
.. .. ..$ baseid : chr [1:297579] "rs3934834" "rs3934834" "rs3934834" "rs3934834" ...
.. ..@ elementType : chr "ANY"
.. ..@ elementMetadata: NULL
.. ..@ metadata : list()
..@ seqinfo :Formal class 'Seqinfo' [package "GenomicRanges"] with 4 slots

```

```

.. ..@ seqnames : chr [1:24] "chr1" "chr2" "chr3" "chr4" ...
.. ..@ seqlengths : int [1:24] 249250621 243199373 198022430 191154276 180915260 171115067
159138663 146364022 141213431 135534747 ...
.. ..@ is_circular: logi [1:24] FALSE FALSE FALSE FALSE FALSE FALSE ...
.. ..@ genome : chr [1:24] "hg19" "hg19" "hg19" "hg19" ...
..@ metadata : list()

```

## Details

This GRanges instance includes locations for 297000 1000 genomes SNP that have been identified as exhibiting LD with NHGRI GWAS SNP as of September 2013. The tagid field tells the name of the tagging SNP, the baseid field is the SNP identifier for the GWAS catalog entry, the R2 field tells the value of R-squared relating the distributions of the tagging SNP and the GWAS entry. Only tagging SNP with R-squared 0.8 or greater are included. A self-contained R-based procedure should emerge in 2014.

## Source

NHGRI GWAS catalog; plink is used with the 1000 genomes VCF in a perl routine by Michael McGeachie, Harvard Medical School;

## Examples

```

data(gwastagger)
gwastagger[1:5]
data(ebicat37)
mean(ebicat37$SNPS %in% gwastagger$baseid)
# ideally, all GWAS SNP would be in our tagging ranges as baseid
query <- setdiff(ebicat37$SNPS, gwastagger$baseid)
# relatively recent catalog additions
sort(table(ebicat37[which(ebicat37$SNPS %in% query)]$DATE.ADDED.TO.CATALOG), decreasing=TRUE)[1:10]

```

---

gwaswloc-class

*Class "gwaswloc"*

---

## Description

A container for GRanges instances representing information in the NHGRI GWAS catalog.

## Objects from the Class

Objects can be created by calls of the form `new("gwaswloc", ...)`. Any GRanges instance can be supplied.

## Slots

**extractDate:** character set manually in .onAttach code to indicate date of retrieval of base table  
**seqnames:** Object of class "Rle" typically representing chromosome numbers of loci associated with specific traits  
**ranges:** Object of class "IRanges" genomic coordinates for locus  
**strand:** Object of class "Rle" identifier of chromosome strand

**elementMetadata:** Object of class "DataFrame" general [DataFrame-class](#) instance providing attributes for the locus-trait association

**seqinfo:** Object of class "Seqinfo"

**metadata:** Object of class "list"

## Extends

Class "[GRanges](#)", directly. Class "[GenomicRanges](#)", by class "GRanges", distance 2. Class "[Vector](#)", by class "GRanges", distance 3. Class "[GenomicRanges\\_OR\\_missing](#)", by class "GRanges", distance 3. Class "[GenomicRanges\\_OR\\_GRangesList](#)", by class "GRanges", distance 3. Class "[Annotated](#)", by class "GRanges", distance 4.

## Methods

[ **signature**(x = "gwaswloc"): a character argument to the bracket will be assumed to be a dbSNP identifier for a SNP locus, and records corresponding to this SNP are extracted; numeric indexes are supported as for [GRanges-class](#) instances.

**getRsids** **signature**(x = "gwaswloc"): extract all dbSNP identifiers as a character vector

**getTraits** **signature**(x = "gwaswloc"): extract all traits (NHGRI term 'Disease/Trait') as a character vector

**subsetByChromosome** **signature**(x = "gwaswloc"): select records by chromosome, a vector of chromosomes may be supplied

**subsetByTraits** **signature**(x = "gwaswloc"): select all records corresponding to a given vector of traits

## Note

In gwascat package 1.9.6 and earlier, the globally accessible gwaswloc instance gwrngs was created upon attachment. This is no longer the case.

## Author(s)

VJ Carey <stvjc@channing.harvard.edu>

## References

<http://www.ebi.ac.uk/gwas/>

## Examples

```
showClass("gwaswloc")
```

---

 gwce2gviz

 Prepare salient components of GWAS catalog for rendering with Gviz
 

---

### Description

Prepare salient components of GWAS catalog for rendering with Gviz

### Usage

```
gwce2gviz(basegr, contextGR = GRanges(seqnames =
  "chr17", IRanges(start = 37500000, width = 1e+06)),
  txrefpk = "TxDb.Hsapiens.UCSC.hg19.knownGene", genome
  = "hg19", genesympk = "Homo.sapiens", plot.it = TRUE,
  maxmlp = 25)
```

### Arguments

basegr	gwaswloc instance containing information about GWAS in catalog
contextGR	A GRanges instance delimiting the visualization in genomic coordinates
txrefpk	a TxDb package, typically
genesympk	string naming annotationDbi .db package
genome	character tag like 'hg19'
plot.it	logical, if FALSE, just return list
maxmlp	maximum value of $-10 \log p$ – winsorization of all larger values is performed, modifying the contents of Pvalue\_mlp in the elementMetadata for the call

### Examples

```
args(gwce2gviz)
#gwascat:::onAttach("", "gwascat")
data(ebicat37)
seqlevelsStyle(ebicat37) = "UCSC"
gwce2gviz(ebicat37)
```

---

 gwdf\_2012\_02\_02

 internal data frame for NHGRI GWAS catalog
 

---

### Description

convenience container for imported table from NHGRI GWAS catalog

### Usage

```
data("gwdf_2014_09_08")
```

**Format**

A data frame with 17832 observations on the following 34 variables.

'Date Added to Catalog' a character vector  
PUBMEDID a numeric vector  
'First Author' a character vector  
Date a character vector  
Journal a character vector  
Link a character vector  
Study a character vector  
'Disease/Trait' a character vector  
'Initial Sample Size' a character vector  
'Replication Sample Size' a character vector  
Region a character vector  
Chr\_id a character vector  
Chr\_pos a character vector  
'Reported Gene(s)' a character vector  
Mapped\_gene a character vector  
Upstream\_gene\_id a character vector  
Downstream\_gene\_id a character vector  
Snp\_gene\_ids a character vector  
Upstream\_gene\_distance a character vector  
Downstream\_gene\_distance a character vector  
'Strongest SNP-Risk Allele' a character vector  
SNPs a character vector  
Merged a character vector  
Snp\_id\_current a character vector  
Context a character vector  
Intergenic a character vector  
'Risk Allele Frequency' a character vector  
'p-Value' a character vector  
Pvalue\_mlog a character vector  
'p-Value (text)' a character vector  
'OR or beta' a character vector  
'95% CI (text)' a character vector  
'Platform [SNPs passing QC]' a character vector  
CNV a character vector

**Note**

In versions prior to 1.9.6, The `.onAttach` function specifies which data frame is transformed to GRanges. This is now managed manually.



**Source**

<http://www.ebi.ac.uk/gwas/>

**Examples**

```
## Not run:
data(gwdf_2014_09_08)
# try gwascat:::gwdf2GRanges on this data.frame

## End(Not run)
```

---

ldtagr	<i>expand a list of variants by including those in a VCF with LD exceeding some threshold</i>
--------	---

---

**Description**

expand a list of variants by including those in a VCF with LD exceeding some threshold

**Usage**

```
ldtagr(snprng, tf, samples, genome = "hg19", lbmaf = 0.05, lbR2 = 0.8, radius = 1e+05)
```

**Arguments**

snprng	a named GRanges for a single SNP. The name must correspond to the name that will be assigned by <a href="#">genotypeToSnpMatrix</a> to the corresponding column of a SnpMatrix.
tf	TabixFile instance pointing to a bgzipped tabix-indexed VCF file
samples	a vector of sample identifiers, if excluded, all samples used
genome	tag like 'hg19'
lbmaf	lower bound on variant MAF to allow consideration
lbR2	lower bound on R squared for regarding SNP to be incorporated
radius	radius of search in bp around the input range

**Details**

uses `snpStats ld()`

**Value**

a GRanges with names corresponding to 'new' variants and mcols fields 'paramRangeID' (base variant input) and 'R2'

**Note**

slow but safe approach. probably a matrix method could be substituted using the nice sparse approach already in `snpStats`

**Author(s)**

VJ Carey

**Examples**

```
require(GenomicRanges)
if (requireNamespace("gQTLstats")) {
  # install gQTLstats to test this function
  cand = GRanges("1", IRanges(113038694, width=1))
  names(cand) = "rs883593"
  require(VariantAnnotation)
  expath = dir(system.file("vcf", package="gwascat"), patt=".*exon.*gz$", full=TRUE)
  tf = TabixFile(expath)
  ldtagr( cand, tf, lbR2 = .8)
}
# should do with 1000 genomes in S3 bucket and gwascat
```

---

locon6

*location information for 10000 SNPs probed on Affy GW 6.0*

---

**Description**

location information for 10000 SNPs probed on Affy GW 6.0

**Usage**

```
data(locon6)
```

**Format**

A data frame with 10000 observations on the following 3 variables.

dbsnp\_rs\_id a character vector

chrom a character vector

physical\_pos a numeric vector

**Details**

extracted from pd.genomewidesnp.6 v 1.4.0; for demonstration purposes

**Examples**

```
data(locon6)
str(locon6)
```

---

makeCurrentGwascat	<i>read NHGRI GWAS catalog table and construct associated GRanges instance</i>
--------------------	--

---

## Description

read NHGRI table and construct associated GRanges instance

## Usage

```
makeCurrentGwascat(table.url =
  "http://www.ebi.ac.uk/gwas/api/search/downloads/alternative",
  fixNonASCII = TRUE, genome="GRCh38",
  withOnt = TRUE)
```

## Arguments

table.url	string identifying the .txt file curated at EBI/EMBL
fixNonASCII	logical, if TRUE, non-ASCII characters as identified by iconv will be replaced by asterisk
genome	character string: 'GRCh38' is default and yields current image as provided by EMBL/EBI; 'GRCh37' yields a realtime liftOver to hg19 coordinates, via AnnotationHub storage of the chain files. Any other value yields an error.
withOnt	logical indicating whether 'alternative' (ontology-present, includes repetition of loci with one:many ontological mapping) or 'full' (ontology-absent, one record per locus report) version of distributed table

## Details

records for which clear genomic position cannot be determined are dropped from the ranges instance  
 an effort is made to use reasonable data types for GRanges metadata, so some qualifying characters such as (EA) in Risk allele frequency field will simply be omitted during coercion of contents of that field to numeric.

## Value

a GRanges instance

## Author(s)

VJ Carey

## Examples

```
# if you have good internet access
if (interactive()) {
  newcatr = makeCurrentGwascat()
  newcatr
}
```

---

obo2graphNEL	<i>convert a typical OBO text file to a graphNEL instance (using Term elements)</i>
--------------	---

---

### Description

convert a typical OBO text file to a graphNEL instance (using Term elements)

### Usage

```
obo2graphNEL(obo, kill = "\\[Typedef\\]", killTrailSp=TRUE)
node2uri(nn)
uri2node(us)
```

### Arguments

obo	string naming a file in OBO format
nn	node name for converted graph, generally of form EFO:nnnnnnn
us	URI string from GWAS catalog annotation MAPPED_TRAIT_URI
kill	entity types to be excluded from processing – probably this should be in a 'keep' form, but for now this works.
killTrailSp	In the textual version of EFO ca. Aug 2015, there is a trailing blank in the tag field defining EFO:0000001, which is not present in references to this term. Set this to TRUE to eliminate this, or graphNEL construction will fail to validate.

### Details

Very rudimentary list and grep operations are used to retain sufficient information to map the DAG to a graphNEL, using formal term identifiers as node names and 'is-a' relationships as edges, and term names and other metadata are assigned to nodeData components.

### Value

a graphNEL instance

### Note

The OBO for Human Disease ontology is serialized as text with this package.

### Author(s)

VJ Carey <stvjc@channing.harvard.edu>

### References

For use with human disease ontology, [http://www.obofoundry.org/cgi-bin/detail.cgi?id=disease\\_ontology](http://www.obofoundry.org/cgi-bin/detail.cgi?id=disease_ontology)

**Examples**

```
data(efo.obo.g)
requireNamespace("graph")
hn = graph::nodes(efo.obo.g)[1:5]
hn
graph::nodeData(efo.obo.g, hn[5])
```

---

riskyAlleleCount	<i>given a matrix of subjects x SNP calls, count number of risky alleles</i>
------------------	--

---

**Description**

given a matrix of subjects x SNP calls, count number of risky alleles for various conditions, relative to NHGRI GWAS catalog

**Usage**

```
riskyAlleleCount(callmat, matIsAB = TRUE, chr,
  gwwl , snpap = "SNPlocs.Hsapiens.dbSNP144.GRCh37",
  gencode = c("A/A", "A/B", "B/B"))
```

**Arguments**

callmat	matrix with subjects as rows, SNPs as columns; entries can be generic A/A, A/B, B/B, or specific nucleotide calls
matIsAB	logical, FALSE if nucleotide codes are present, TRUE if generic call codes are present; in the latter case, <code>gwascat::ABmat2nuc</code> will be run
chr	code for chromosome, should work with the SNP annotation <code>getSNPlocs</code> function, so likely "ch[nn]"
gwwl	an instance of <a href="#">gwaswloc</a>
snpap	name of a Bioconductor <code>SNPlocs.Hsapiens.dbSNP.*</code> package
gencode	codes used for generic SNP call

**Value**

matrix with rows corresponding to subjects , columns corresponding to SNP

**Examples**

```
data(gg17N) # translated from GGdata chr 17 calls using ABmat2nuc
data(ebicat37)
library(GenomeInfoDb)
seqlevelsStyle(ebicat37) = "UCSC"
h17 = riskyAlleleCount(gg17N, matIsAB=FALSE, chr="ch17", gwwl=ebicat37)
h17[1:5,1:5]
table(as.numeric(h17))
```

---

topTraits	<i>operations on GWAS catalog</i>
-----------	-----------------------------------

---

**Description**

operations on GWAS catalog

**Usage**

```
topTraits(gwwl, n=10, tag="DISEASE/TRAIT")
locs4trait(gwwl, trait, tag="DISEASE/TRAIT")
chklocs(chrtag="20", gwwl)
```

**Arguments**

gwwl	instance of <a href="#">gwaswloc</a>
n	numeric, number of traits to report
tag	character, name of field to be used for trait enumeration
trait	character, trait to use for filtering
chrtag	character, chromosome identifier

**Value**

topTraits returns a character vector of most frequently occurring traits in the database

locs4trait returns a [gwaswloc](#) object with records defining associations to the specified trait

chklocs returns a logical that is TRUE when the asserted locations of SNP in the GWAS catalog agree with the locations given in the dbSNP package SNPlocs.Hsapiens.dbSNP144.GRCh37

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

**Examples**

```
data(ebicat38)
topTraits(ebicat38)
```

---

traitsManh	<i>use ggbio facilities to display GWAS results for selected traits in genomic coordinates</i>
------------	--

---

**Description**

use ggbio facilities to display GWAS results for selected traits in genomic coordinates

**Usage**

```
traitsManh(gwr, selr = GRanges(seqnames = "chr17", IRanges(3e+07, 5e+07)), traits = c("Asthma", "Par"))
```

**Arguments**

<code>gwr</code>	GRanges instance as managed by the gwaswloc container design, with Disease.Trait and Pvalue\_mlog among elementMetadata columns
<code>selr</code>	A GRanges instance to restrict the gwr for visualization. Not tested for noncontiguous regions.
<code>traits</code>	Character vector of traits to be exhibited; GWAS results with traits not among these will be labeled "other".
<code>truncmlp</code>	Maximum value of $-\log_{10} p$ to be displayed; in the raw data this ranges to the hundreds and can cause bad compression.
<code>...</code>	not currently used

**Details**

uses a ggbio autoplot

**Value**

autoplot value

**Note**

An xlab is added, concatenating genome tag with seqnames tag.

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

**Examples**

```
# do a p-value truncation if you want to reduce compression
## Not run: # ggbio July 2018
data(ebicat38)
library(GenomeInfoDb)
seqlevelsStyle(ebicat38) = "UCSC"
traitsManh(ebicat38)

## End(Not run)
```

# Index

- \*Topic **classes**
  - gwaswloc-class, 5
- \*Topic **datasets**
  - gwastagger, 4
  - gwdf\_2012\_02\_02, 7
  - locon6, 10
- \*Topic **graphics**
  - gwceX2gviz, 7
  - traitsManh, 15
- \*Topic **models**
  - bindcadd\_snv, 3
  - ldtagr, 9
  - makeCurrentGwascat, 11
  - obo2graphNEL, 12
  - riskyAlleleCount, 13
  - topTraits, 14
  - traitsManh, 15
- \*Topic **package**
  - gwascat-package, 2
- [, gwaswloc, ANY, ANY, ANY-method (gwaswloc-class), 5
- [, gwaswloc, ANY-method (gwaswloc-class), 5
- [, gwaswloc-method (gwaswloc-class), 5
- Annotated, 6
- bindcadd\_snv, 3
- chklocs (topTraits), 14
- DataFrame (gwascat-package), 2
- ebicat37 (gwascat-package), 2
- ebicat38 (gwascat-package), 2
- efo.obo.g (obo2graphNEL), 12
- g17SM (gwascat-package), 2
- GenomicRanges, 6
- GenomicRanges\_OR\_GRangesList, 6
- GenomicRanges\_OR\_missing, 6
- genotypeToSnpMatrix, 9
- getRsids (gwaswloc-class), 5
- getRsids, gwaswloc-method (gwaswloc-class), 5
- getTraits (gwaswloc-class), 5
- getTraits, gwaswloc-method (gwaswloc-class), 5
- gg17N (gwascat-package), 2
- GRanges, 6
- gw6.rs\_17 (gwascat-package), 2
- gwascat (gwascat-package), 2
- gwascat-package, 2
- gwastagger, 4
- gwaswloc, 13, 14
- gwaswloc-class, 5
- gwceX2gviz, 7
- gwdf\_2012\_02\_02, 7
- gwdf\_2014\_09\_08 (gwdf\_2012\_02\_02), 7
- gwrngs19 (gwascat-package), 2
- gwrngs38 (gwascat-package), 2
- impute.snps, 2
- impute.snps (gwascat-package), 2
- ldtagr, 9
- locon6, 10
- locs4trait (topTraits), 14
- low17 (gwascat-package), 2
- makeCurrentGwascat, 11
- node2uri (obo2graphNEL), 12
- obo2graphNEL, 12
- riskyAlleleCount, 13
- rules\_6.0\_1kg\_17 (gwascat-package), 2
- show, gwaswloc-method (gwaswloc-class), 5
- si.hs.38 (gwascat-package), 2
- SnpMatrix-class (gwascat-package), 2
- subsetByChromosome (gwaswloc-class), 5
- subsetByChromosome, gwaswloc-method (gwaswloc-class), 5
- subsetByTraits (gwaswloc-class), 5
- subsetByTraits, gwaswloc-method (gwaswloc-class), 5
- topTraits, 14



traitsManh, [15](#)

uri2node (obo2graphNEL), [12](#)

Vector, [6](#)