

# MergeMaid

October 5, 2010

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`mergeCoeff`

*Class mergeCoeff, a class for storing regression coefficients.*

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## Description

This is the class representation for output from regression coefficient calculations

## Slots

**coeff** Object of class matrix, A matrix storing default coefficients.

**coeff.std** Object of class matrix, A matrix storing standardized coefficients.

**zscore** Object of class matrix, A matrix storing zscores.

## Methods

Class-specific methods:

**coeff (mergeCoeff)** Accessor function for the coeff slot.

**coeff<- (mergeCoeff)** Replacement function for the coeff slot.

**stdcoeff (mergeCoeff)** Accessor function for the coeff.std slot.

**stdcoeff<- (mergeCoeff)** Replacement function for the coeff.std slot.

**zscore (mergeCoeff)** Accessor function for the zscore slot.

**zscore<- (mergeCoeff)** Replacement function for the zscore slot.

Standard generic methods:

**plot (list)** This method is not formally defined for mergeCoeff objects but for a matrix. This function would typically be called with the following syntax, `plot(coeff(mergeCoeff))`. The result is pairwise scatterplots of the columns of the selected matrix. If there are two studies, this is a single scatterplot.

## See Also

[mergeExprs](#), [modelOutcome](#), [mergeExpressionSet-class](#)

**Examples**

```

if(require(Biobase) & require(MASS) & require(survival)){

data(mergeData)
merged <- mergeExprs(sample1, sample2, sample3)

log.coeff <- modelOutcome(merged, outcome=c(1,1,1), method="logistic")
plot(coeff(log.coeff))
plot(stdcoeff(log.coeff), pch=4, labels=c("study A", "study B", "study C"), col=3)

linear.coeff <- modelOutcome(merged[1:2], outcome=c(3,3), method="linear")
plot(zscore(linear.coeff))
plot(zscore(linear.coeff), xlab="study A", ylab="study B", col=2)
}

```

mergeCor

---

*Class mergeCor, a class for storing data relevant to integrative correlation coefficients.*

---

**Description**

This is the class representation for integrative correlation coefficients.

**Details**

If 'n' is the number of studies then for  $i < j \leq n$ , the pairwise correlation of correlations for studies  $i$  and  $j$  is stored in column  $((i-1)*(n-1)-(i-2)*(i-1))/2 + j - i$  of the pairwise.cors slot.

**Slots**

**pairwise.cors** Object of class matrix. Each column contains correlation of correlation score for genes in a pair of study.

**maxcors** Object of class vector. Each slot represents maximal canonical correlation (pairwise canonical correlations) for each pair of studies.

**notes** Object of class vector. Each slot contains notes for each study.

**Methods**

Class-specific methods:

**cors(mergeCor)** Accessor function for the cors slot.

**pairs.cors(mergeCor)** Accessor function for the pairwise.cors slot

**integrate.cors(mergeCor)** Accessor function, returns integrative correlation (average pairwise correlation of correlations) for each gene. If adjust is TRUE, returns the integrate correlation divided by the corresponding canonical correlation, and the default value is FALSE.

**maxcors(mergeCor)** Accessor function, returns maximal canonical correlation (pairwise canonical correlations) for each pair of studies.

Standard generic methods:

**notes(mergeCor)** An accessor function for the notes slot.

**hist(mergeCor,...)** Draw histograms of integrative correlation, here we use the approximate method to calculate the integrative correlation.

**See Also**

[mergeCor-class](#), [intCor](#), [modelOutcome](#), [mergeExpressionSet-class](#)

**Examples**

```

if(require(Biobase) & require(MASS)){
  data(mergeData)
  merged <- mergeExprs(sample1, sample2, sample3)
  intcor3 <- intCor(merged, method="pearson")
  plot(merged)
  intcor2 <- intCor(merged[1:2], exact=FALSE)
  plot(merged, pch=4, col=5)

  pairwise.cors(intcor3)
  integrative.cors(intcor3)
  integrative.cors(intcor2) ["Hs.12101"]
  maxcors(intcor2)
}

```

---

`mergeExpressionSet` *Class mergeExpressionSet, a class for merged microarray data, and methods for processing them*

---

**Description**

This is class representation for merged Microarray Data.

**Details**

The `mergeExpressionSet` class is conceived as an extension of the `ExpressionSet` class provided in `Biobase` for the storage of expression array data. A `mergeExpressionSet` object is primarily a list of `ExpressionSet` objects, along with an incidence matrix indicating which genes appear in which studies. A `mergeExpressionSet` object with a single study reverts to the `ExpressionSet` class. A number of accessor functions are defined for this class, as well as a few convenient analysis and plotting functions.

**Slots**

We assume there are  $K$  studies, representing a total of  $M$  unique genes.

**data** A list of `ExpressionSet` objects.

**geneStudy** Binary incidence matrix with  $M$  rows and  $K$  columns. Each column represents a study, and each row represents a gene. If study "s" contains gene "g", then `geneStudy[g,s]=1`, otherwise `geneStudy[g,s]=0`.

**notes** Object of class "character" This slot is available for storage of descriptive information.

## Methods

Derived from [ExpressionSet](#):

**exprs(mergeExpressionSet)** An accessor function for the data slot.

**exprs<- (mergeExpressionSet)** A replace function for the data slot

**notes (mergeExpressionSet)** An accessor function for the notes slot.

**notes<- (mergeExpressionSet)** A replace function for the notes slot.

**geneNames (mergeExpressionSet)** Accessor function for union of gene ids in all studies.

**geneNames<- (mergeExpressionSet)** A replace function for gene ids.

Class-specific methods:

**geneStudy (mergeExpressionSet)** Accessor function for the geneStudy slot.

**phenoData (mergeExpressionSet)** Accessor function for phenodata in ExpressionSet's. Returns a list, one phenodata matrix per study.

**phenoData<- (mergeExpressionSet)** A replace function for phenodata in ExpressionSet's. Returns a list, one phenodata matrix per study.

**intersection (mergeExpressionSet)** Represent data for genes common to all studies as a single ExpressionSet object.

**modelOutcome (mergeExpressionSet)** Calculate regression coefficients for each study/gene.

**intCor (mergeExpressionSet)** Calculate the integrative correlation coefficients for mergeExpressionSet data.

**intcorDens (mergeExpressionSet)** Plot the distribution of the integrative correlation coefficients and the null distribution obtained by permutation here we use the approximate method to calculate the integrative correlation.

Standard generic methods:

**length (mergeExpressionSet)** Function returning the number of studies in the mergeExpressionSet.

**names (mergeExpressionSet)** Function returning study names.

**names<- (mergeExpressionSet)** A replace function for study names.

**[ (mergeExpressionSet)** A subset operator. Returns a mergeExpressionSet containing a subset of the studies. A mergeExpressionSet with only one study is returned as a single ExpressionSet.

**summary (mergeExpressionSet)** Obtain the basic information for 'mergeExpressionSet'.

**plot (mergeExpressionSet)** Draw scatterplots to compare integrative correlations for genes, here we use the approximate method to calculate the integrative correlation.

## See Also

[mergeExprs](#), [intCor](#), [modelOutcome](#), [intcorDens](#), [ExpressionSet](#)

## Examples

```
if(require(Biobase) & require(MASS)){
  data(mergeData)
  merged <-mergeExprs(sample1, sample2, sample3)

  merged[1:2]
  i<-c(1, 3)
```

```

merged[i]

exprs (merged)

names (merged) <- c ("study1", "study2", "study3")

length (merged)

summary (merged)

plot (merged)

plot (merged[1:2])

intcorDens (merged)

inter  <- intersection (merged)
}

```

---

intCor

*Correlation of Correlations*


---

### Description

Given a mergeExpressionSet, this function calculates the study specific correlation matrices, and, for each gene, the correlation of correlations.

### Usage

```
intCor(x, method= c("pearson", "spearman"), exact, ...)
```

### Arguments

x	Object of class mergeExpressionSet.
method	Method used to calculate correlation coefficient. If exact is TRUE, the available methods to use is "spearman" and "pearson"; If exact is FALSE, the available methods to use is "pearson".
exact	If exact is TRUE, we use the standard method the calculate the integrative correlation; If exact is FALSE, we use the approximate method the calculate.
...	Not implemented at this time

### Details

Integrative correlation coefficients are calculated as follows. The first step is to identify the n genes common to all studies. Within each study, we calculate the correlation coefficient between gene g, and every other common gene. This gives a vector of length n-1. For a pair of studies, S1 and S2, we calculate the correlation of correlations for gene g. When there are more than 2 studies under consideration, all pairwise correlation of correlations are calculated and averaged.

### Value

The output is an object of class mergeCor.

**See Also**

[mergeCor-class](#), [intcorDens](#)

**Examples**

```

if(require(Biobase) & require(MASS)){
  data(mergeData)
  merged <-mergeExprs(sample1, sample2, sample3)
  corcor <-intCor(merged,method="spearman")

  plot(merged)
  hist(corcor)

  corcor <-intCor(merged,method="pearson",exact=FALSE)
  corcor <-intCor(merged[1:2])
  corcor <-intCor(merged,exact=TRUE)

  vv<-c(1,3)
  corcor1 <-intCor(merged[vv])
  plot(merged,xlab="study A",ylab="study B",main="CORRELATION OF CORRELATION",col=3,pch=4)
  hist(corcor1,xlab="CORRELATION OF CORRELATION")
}

```

---

intcorDens

*plot of density functions of integrative correlations*

---

**Description**

Given a `mergeExpressionSet`, this function calculates and plots the density function of the approximate integrative correlations, as well as densities for the "null distributions" obtained by randomly permuting sample IDs.

**Usage**

```
intcorDens(x, method, ...)
```

**Arguments**

<code>x</code>	Object of class <code>mergeExpressionSet</code> .
<code>method</code>	The available method to use is "pearson".
<code>...</code>	Graphical parameters to be passed to plot.

**Details**

Here we use the approximate method to calculate the integrative correlation.

**Value**

The value is null. Returns a plot.

**See Also**

[mergeExpressionSet-class,intCor,modelOutcome](#)

**Examples**

```
if(require(Biobase) & require(MASS)){
  data(mergeData)
  merged <-mergeExprs(sample1, sample2, sample3)

  intcorDens(merged)

  intcorDens(merged, cex.legend=1.5)

  intcorDens(merged, lty=2)
}
```

---

intersection

*ExpressionSet with all common genes in a mergeExpressionSet*

---

**Description**

Given a mergeExpressionSet, this function returns a single ExpressionSet. Only genes common to all studies are included. Expression data for all studies sits side by side in the 'exprs' slot. The 'notes' slot is used for information about the study identity of each sample.

**Usage**

```
intersection(x)
```

**Arguments**

x                    Object of class mergeExpressionSet.

**Value**

Returns an object of class ExpressionSet

**See Also**

[mergeExpressionSet-class](#)

**Examples**

```
data(mergeData)
merged <-mergeExprs(sample1, sample2, sample3)

inter <- intersection(merged)
```

---

`mergeData`*MergeMaid instance data for merging*

---

**Description**

These are 3 artificial data sets, generated using `rnorm()`, for the purpose of illustrating the package. All are list objects. For some of the "genes", the expression values in the 3 datasets are not independent.

**Usage**

```
data(mergeData)
```

---

`mergeExprs`*Merge gene expression data sets*

---

**Description**

Merges gene expression data from different studies.

**Usage**

```
mergeExprs(...)
```

**Arguments**

... Input objects can be any combination of `mergeExpressionSet`, `ExpressionSet`, matrix or a list. A list should have the following slots: expression matrix, pheno data matrix, gene names vector, notes. The order of the four slots is fixed. A matrix should have genes ids as its row names, as should the `exprs` slot of an `ExpressionSet`. Since merging depends on geneids, these conventions are essential.

**Details**

The `mergeExpressionSet` object is the standard input for all functions in the `MergeMaid` package. Use the `mergeExprs` function when creating `mergeExpressionSet` objects to ensure that all necessary information is available for further analysis.

**Value**

The output is a `mergeExpressionSet`.

**See Also**

[mergeExpressionSet-class](#)



**Examples**

```

if(require(Biobase) & require(MASS)){
  data(mergedData)
  merged <-mergeExprs(sample1, sample2, sample3)

  rr<-rnorm(200*22,0,1)
  mm<-matrix(rr,200,22)
  rownames(mm)<-sample2[[3]]
  merge.m<-mergeExprs(sample1,mm,sample2)
  intcor.m<-intCor(merge.m)
  plot(merge.m)

  rr<-rnorm(200*50,0,1)
  mm2<-matrix(rr,200,50)
  ph.ll<-as.data.frame(rbinom(50,1,.5))
  ll<-list(mm2,ph.ll,sample2[[3]],"list 2")
  merge.t<-mergeExprs(sample1,mm,sample2,ll)
  intcor.t<-intCor(merge.t)
  plot(merge.t)

  merge.a<-mergeExprs(sample3,merge.m,ll)
  inter<-intersection(merge.a)
  summary(merge.a)
}

```

---

modelOutcome

*Compare regression coefficients across studies*


---

**Description**

Given a set of merged studies, this function calculates study specific regression coefficients for each gene.

**Usage**

```
modelOutcome(x, outcome, outcome2=NULL, method=c("linear", "logistic", "cox"), ...)
```

**Arguments**

<code>x</code>	Object of class <code>mergeExpressionSet</code> .
<code>method</code>	Method specifies the model used to generate coefficients. At this time only linear regression, logistic regression, and Cox hazard rates are implemented.
<code>outcome, outcome2</code>	The format for the outcome variable depends on the model used. For linear regression, outcome should be a continuous response variable, for logistic regression, it should be a binary response variable, and for Cox hazard rates it should be time of event. Outcome 2 is currently used only in the calculation of hazard rates, and should be a binary variable indicating censoring status for each subject. If outcome is a vector of length equal to number of studies, then each element represents the column in the <code>ExpressionSet</code> <code>phenoData</code> slot for that study. If outcome is a list, then each list element should have actual outcome data for the corresponding study.
<code>...</code>	Not implemented at this time

**Value**

The output is a mergeCoeff object.

**See Also**

[modelOutcome](#), [mergeCoeff-class](#)

**Examples**

```
if(require(Biobase) & require(MASS) & require(survival)){
  data(mergedData)
  merged <- mergeExprs(sample1, sample2, sample3)

  log.coeff <- modelOutcome(merged, outcome=c(1,1,1), method="logistic")
  plot(coeff(log.coeff))

  linear.coeff <- modelOutcome(merged[1:2], outcome=c(3,3), method="linear")
  plot(zscore(linear.coeff), xlab="study 1", ylab="study 2")

  event1<-rbinom(100,1,.5)
  event2<-rbinom(50,1,.5)
  event3<-rbinom(70,1,.5)

  out1<-rnorm(100,5,1)
  out2<-rnorm(50,5,1)
  out3<-rnorm(70,5,1)

  out<-list(out1,out2,out3)
  even<-list(event1,event2,event3)

  cox.coeff<-modelOutcome(merged, outcome2=even, outcome=out, method="cox")
  plot(coeff(cox.coeff))
}
```

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