

# Package ‘Category’

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**Title** Category Analysis

**Version** 2.28.0

**Author** R. Gentleman with contributions from S. Falcon and D.Sarkar

**Description** A collection of tools for performing category analysis.

**Maintainer** Bioconductor Package Maintainer <maintainer@bioconductor.org>

**License** Artistic-2.0

**Depends** BiocGenerics, AnnotationDbi, Biobase, Matrix, GO.db, methods

**Suggests** EBarrays, ALL, Rgraphviz, RColorBrewer, xtable (>= 1.4-6), hgu95av2.db, KEGG.db, SNPchip, geneplotter, limma, lattice, graph, Biobase, gene-filter, methods, RUnit, org.Sc.sgd.db, GOstats

**Imports** BiocGenerics, graph, methods, Biobase, AnnotationDbi, RBGL, GSEABase (>= 1.19.3), genefilter, annotate (>= 1.15.6), stats, utils

**LazyLoad** Yes

**Collate** AllClasses.R AllGenerics.R categoryToEntrezBuilder-methods.R  
categoryName-methods.R hyperg-methods.R hyperGTest-methods.R  
linearMTest-methods.R DatPkg-accessors.R ChrBandTree.R  
tree\_visitor.R cb\_test.R initialize-methods.R  
HyperGParams-accessors.R LinearMParams-accessors.R  
HyperGResult-accessors.R LinearMResult-accessors.R  
ChrMapHyperGResult-accessors.R ChrMapLinearMResult-accessors.R  
show-methods.R universeBuilder-methods.R utils.R MAPcode.R  
catcode.R cateGOryMatrix.R gseaperm.R summary-methods.R mouse.R

**biocViews** Annotation, GO, Pathways, GeneSetEnrichment

**R topics documented:**

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applyByCategory	<i>Apply a function to a vector of statistics, by category</i>
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## Description

For each category, apply the function FUN to the set of values of stats belonging to that category.

## Usage

```
applyByCategory(stats, Amat, FUN = mean, ...)
```

## Arguments

stats	Numeric vector with test statistics of interest.
Amat	A logical or numeric matrix: the adjacency matrix of the bipartite genes - category graph. Its rows correspond to the categories, columns to the genes, and TRUE or a numeric value different from 0 indicates membership. The columns are assumed to be aligned with the elements of stats.
FUN	A function to apply to the subsets stats by categories.
...	Extra parameters passed to FUN.

## Details

For GO categories, the function [cateGOry](#) might be useful for the construction of Amat.

## Value

The return value is a list or vector of length equal to the number of categories. Each element corresponds to the values obtained by applying FUN to the subset of values in stats according to the category defined for that row.

## Author(s)

R. Gentleman, contributions from W. Huber

## See Also

[apply](#)

## Examples

```
set.seed(0xabcd)
st = rnorm(20)
names(st) = paste("gene", 1:20)

a = matrix(sample(c(FALSE, TRUE), 60, replace=TRUE), nrow=3,
            dimnames = list(paste("category", LETTERS[1:3]), names(st)))

applyByCategory(st, a, median)
```

---

cateGOry	<i>Construct a category membership matrix from a list of gene identifiers and their annotated GO categories.</i>
----------	--

---

## Description

The function constructs a category membership matrix, such as used by [applyByCategory](#), from a list of gene identifiers and their annotated GO categories. For each of the GO categories stated in `categ`, all less specific terms (ancestors) are also included, thus one need only obtain the most specific set of GO term mappings, which can be obtained from Bioconductor annotation packages or via **biomaRt**. The ancestor relationships are obtained from the **GO.db** package.

## Usage

```
cateGOry(x, categ, sparse=FALSE)
```

## Arguments

<code>x</code>	Character vector with (arbitrary) gene identifiers. They will be used for the column names of the resulting matrix.
<code>categ</code>	A character vector of the same length as <code>x</code> with GO annotations for the genes in <code>x</code> . If a gene has multiple GO annotations, it is expected to occur multiple times in <code>x</code> , once for each different annotation.
<code>sparse</code>	Logical. If TRUE, the resulting matrix is constructed using <a href="#">Matrix</a> , otherwise, R's base <code>matrix</code> is used.

## Details

The function requires the [GO](#) package.

For subsequent analyses, it is often useful to remove categories that have only a small number of members. Use the normal matrix subsetting syntax for this, see example.

If a GO category in `categ` is not found in the GO annotation package, a warning will be generated, and no ancestors for that GO category are added (but that category itself will be part of the returned adjacency matrix).

## Value

The adjacency matrix of the bipartite category membership graph, rows are categories and columns genes.

## Author(s)

Wolfgang Huber

## See Also

[applyByCategory](#)

**Examples**

```

g = cateGOry(c("CG2671", "CG2671", "CG2950"),
             c("GO:0090079", "GO:0001738", "GO:0003676"), sparse=TRUE)

g

rowSums(g)  ## number of genes in each category

## Filter out categories with less than minMem and more than maxMem members.
## This is toy data, in real applications, a choice of minMem higher
## than 2 will be more appropriate.
filter = function(x, minMem = 2, maxMem = 35) ((x>=minMem) & (x<=maxMem))
g[filter(rowSums(g)),,drop=FALSE ]

```

Category-defunct

*Defunct Functions in Package* **Category****Description**

The functions or variables listed here are no longer part of the Category package.

**Usage**

```

condGeneIdUniverse()
isConditional()
geneGoHyperGeoTest()
geneKeggHyperGeoTest()
cb_parse_band_hsa()
chrBandInciMat()

```

**See Also**

[Defunct](#)

categoryToEntrezBuilder

*Return a list mapping category ids to Entrez Gene ids***Description**

Return a list mapping category ids to the Entrez Gene ids annotated at the category id. Only those category ids that have at least one annotation in the set of Entrez Gene ids specified by the geneIds slot of p are included.

**Usage**

```
categoryToEntrezBuilder(p)
```

**Arguments**

p                      A subclass of HyperGParams-class

**Details**

End users **should not** call this directly. This method gets called from hyperGTest. To add support for a new category, a new method for this generic must be defined. Its signature should match a subclass of HyperGParams-class appropriate for the new category.

**Value**

A list mapping category ids to Entrez Gene identifiers.

**Author(s)**

S. Falcon

**See Also**

[hyperGTest HyperGParams-class](#)

---

cb_contingency	<i>Create and Test Contingency Tables of Chromosome Band Annotations</i>
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---

**Description**

For each chromosome band identifier in chrVect, cb\_contingency builds and performs a test on a 2 x k contingency table for the genes from selids found in the child bands of the given chrVect element.

cb\_sigBands extracts the chromosome band identifiers that were in a contingency table that tested significant given the specified p-value cutoff.

cb\_children returns the child bands of a given band in the chromosome band graph. The argument must have length equal to one.

**Usage**

```
cb_contingency(selids, chrVect, chrGraph, testFun = chisq.test,
               min.expected = 5L, min.k = 1L)

cb_sigBands(b, p.value = 0.01)

cb_children(n, chrGraph)
```

**Arguments**

<code>selids</code>	A vector of the selected gene identifiers (usual Entrez IDs).
<code>chrVect</code>	A character vector of chromosome band identifiers
<code>chrGraph</code>	A graph object as returned by <code>makeChrBandGraph</code> . The nodes should be chromosome band IDs and the edges should represent the tree structure of the bands. Furthermore, the graph is expected to have a "geneIds" node attribute providing a vector of gene IDs annotated at each band.
<code>testFun</code>	The function to use for testing the 2 x k contingency tables. The default is <code>chisq.test</code> . It will be called with a single argument, a 2 x k matrix representing the contingency table.
<code>min.expected</code>	A numeric value specifying the minimum expected count for columns to be included in the contingency table. The expected count is $(\text{rowSum} * \text{colSum}) / n$ . Chromosome bands with a select cell count less than <code>min.expected</code> are dropped from the table before testing occurs. If <code>NULL</code> , then no bands will be dropped.
<code>min.k</code>	An integer giving the minimum number of chromosome bands that must be present in a contingency table in order to proceed with testing.
<code>b</code>	A list as returned by <code>cb_contingency</code>
<code>p.value</code>	A p-value cutoff to use in selecting significant contingency tables.
<code>n</code>	A length one character vector specifying a chromosome band annotation. Bands not found in <code>chrGraph</code> will return <code>character(0)</code> when passed to <code>cb_children</code> .

**Details**

`cb_sigBands` assumes that the p-value associated with a result of `testFun` can be accessed as `testFun(t)$p.value`. We should improve this to be a method call which can then be specialized based on the class of the object returned by `testFun`.

**Value**

`cb_contingency` returns a list with an element for each test performed. This will most often be shorter than `length(chrVect)` due to skipped tests based on `min.found` and `min.k`. Each element of the returned list is itself a list with components:

<code>table</code>	A 2 x k contingency table
<code>result</code>	The output of <code>testFun</code> applied to the table.

`cb_sigBands` returns a character vector of chromosome band identifiers that are in one of the contingency tables that had a p-value less than the cutoff specified by `p.value`.

**Author(s)**

Seth Falcon

---

`cb_parse_band_Hs`*Parse Homo Sapiens Chromosome Band Annotations*

---

**Description**

This function parses chromosome band annotations as found in the <chip>MAP map of Bioconductor annotation data packages. The return value is a vector of parent bands up to the relevant chromosome.

**Usage**

```
cb_parse_band_Hs(x)
```

**Arguments**

`x` A chromosome band annotation given as a string.

**Details**

The former function `cb\_parse\_band\_hsa` is now deprecated.

**Value**

A character vector giving the path to the relevant chromosome.

**Author(s)**

Seth Falcon

**Examples**

```
cb_parse_band_Hs("12q32.12")
```

---

`cb_parse_band_Mm`*Parse Mus Musculus Chromosome Band Annotations*

---

**Description**

This function parses chromosome band annotations as found in the <chip>MAP map of Bioconductor annotation data packages. The return value is a vector of parent bands up to the relevant chromosome.

**Usage**

```
cb_parse_band_Mm(x)
```

**Arguments**

x                      A chromosome band annotation given as a string.

**Value**

A character vector giving the path to the relevant chromosome.

**Author(s)**

Seth Falcon & Nolwenn Le Meur

**Examples**

```
cb_parse_band_Mm("10 B3")
```

---

cb_test	<i>Chromosome Band Tree-Based Hypothesis Testing</i>
---------	--

---

**Description**

cb\_test is a flexible tool for discovering interesting chromosome bands relative to a selected gene list. The function supports local and global tests which can be carried out in a top down or bottom up fashion on the tree of chromosome bands.

**Usage**

```
cb_test(selids, chrtree, level, dir = c("up", "down"),
        type = c("local", "global"), next.pval = 0.05,
        cond.pval = 0.05, conditional = FALSE)
```

**Arguments**

selids	A vector of gene IDs. The IDs should match those used to annotate the ChrBandTree given by chrtree. In most cases, these will be Entrez Gene IDs.
chrtree	A ChrBandTree object representing the chromosome bands and the mapping to gene identifiers. The genes in the ChrBandTree are limited to the universe of gene IDs specified at object creation time.
level	An integer giving the level of the chromosome band tree at which testing should begin. The level is conceptualized as the set of nodes with a given path length to the root (organism) node of the chromosome band tree. So level 1 is the chromosome and level 2 is the chromosome arms. You can get a better sense by calling <code>exampleLevels(chrtree)</code>
dir	A string giving the direction in which the chromosome band tree will be traversed when carrying out the tests. A bottom up traversal, from leaves to root, is specified by "up". A top down, from root to leaves, traversal is specified by "down".

type	A string giving the type of test to perform. The current choices are "local" and "global". A local test carries out a <code>chisq.test</code> on each 2 x K contingency table induced by each set of siblings at a given level in the tree. A global test uses the Hypergeometric distribution to compute a p-value for the 2 x 2 tables induced by each band treated independently.
next.pval	The p-value cutoff used to determine whether the parents or children of a node should be tested. After testing a given level of the tree, the decision of whether or not to continue testing the children (or parents) of the already tested nodes is made by comparing the p-value result for a given node with this cutoff; relatives of nodes with values strictly greater than the cutoff are skipped.
cond.pval	The p-value cutoff used to determine whether a node is significant during a conditional test. See <code>conditional</code> .
conditional	A logical value. Can only be used when <code>dir="up"</code> and <code>type="global"</code> . In this case, a TRUE value causes a conditional Hypergeometric calculation to be performed. The genes annotated at significant children of a given band are removed before testing.

### Value

A list with an element for each level of the tree that was tested. Note that the first element will correspond to the level given by `level` and that subsequent elements will be the next or previous depending on `dir`.

Each level element is itself a list consisting of a result list for each node or set of nodes tested. These inner-most lists will have, at least, the following components:

nodes	A character vector of the nodes involved in the test.
p.value	The p-value for the test
observed	The contingency table
method	A brief description of the test method

### Author(s)

Seth Falcon

---

ChrBandTree-class	<i>Class "ChrBandTree"</i>
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---

### Description

This class represents chromosome band annotation data for a given experiment. The class is responsible for storing the mapping of band to set of gene IDs located within that band as well as for representing the tree structured relationship among the bands.

### Objects from the Class

Objects should be created using `NewChrBandTree` or `ChrBandTreeFromGraph`.

**Slots**

**toParentGraph:** Object of class "graph" representing the tree of chromosome bands. Edges in this directed graph go from child to parent.

**toChildGraph:** Object of class "graph". This is the same as toParentGraph, but with the edge directions reversed. This is not an ideal implementation due to the duplication of data, but it provides quick access to parents or children of a given node.

**root:** Object of class "character" giving the name of the root node. The convention is to use "ORGANISM:<organism>".

**level2nodes:** Object of class "list" providing a mapping of levels in the tree to the set of nodes at that level. Levels X is defined as the set of nodes with a path length of X from the root node.

**Methods**

**allGeneIds** Return a vector of gene IDs representing the gene universe for this ChrBandTree

**childrenOf** Return a list with an element for each the character vector n. Each element is a character vector of node names of the children of the named element.

**geneIds** Return a vector of gene IDs for a single band.

**lgeneIds** Return a list of vectors of gene IDs when given more than one band. The "l" prefix is for list.

**parentOf** Return the parents of the specified bands. See childrenOf for a description of the structure of the return value.

**treeLevels** Return an integer vector identifying the levels of the tree.

**level2nodes(g, level)** Return the nodes in the tree that are at the level specified by level. The level argument can be either numeric or character, but should match a level returned by treeLevels.

**Note**

Not all known chromosome bands will be represented in a given instance. The set of bands that will be present is determined by the available annotation data and the specified gene universe. The annotation source maps genes to their most specific band. Such bands and all bands on the path to the root will be represented in the resulting tree.

Currently there is only support for human and mouse data.

**Author(s)**

S. Falcon

**Examples**

```
library("hgu95av2.db")
set.seed(0xfeee)
univ = NULL ## use all Entrez Gene IDs on the chip (not recommended)
ct = NewChrBandTree("hgu95av2.db", univ)

length(allGeneIds(ct))
```

```

exampleLevels(ct)

geneIds(ct, "10p11")
lgeneIds(ct, "10p11")
lgeneIds(ct, c("10p11", "Yq11.22"))

pp = parentOf(ct, c("10p11", "Yq11.22"))
childrenOf(ct, unlist(pp))

treeLevels(ct)

level2nodes(ct, 0)
level2nodes(ct, 0L)
level2nodes(ct, "0")

level2nodes(ct, 1)

```

---

ChrMapHyperGParams-class

*Class "ChrMapHyperGParams"*


---

## Description

This class encapsulates parameters needed for Hypergeometric testing of over or under representation of chromosome bands among a selected gene list using [hyperGTest](#).

## Objects from the Class

Objects can be created by calls of the form `new("ChrMapHyperGParams", ...)`.

## Slots

**chrGraph:** Object of class "graph". The nodes are the chromosome bands and the edges describe the tree structure of the bands. Each node has a "geneIds" node attributes (see `nodeData`) which contains a vector of gene IDs annotated at the given band.

**conditional:** Object of class "logical", indicating whether the test performed should be a conditional test.

**geneIds:** Object of class "ANY": A vector of gene identifiers. Numeric and character vectors are probably the only things that make sense. These are the gene ids for the selected gene set.

**universeGeneIds:** Object of class "ANY": A vector of gene ids in the same format as `geneIds` defining a subset of the gene ids on the chip that will be used as the universe for the hypergeometric calculation. If this is NULL or has length zero, then all gene ids on the chip will be used.

**annotation:** A string giving the name of the annotation data package for the chip used to generate the data.

**categorySubsetIds:** Object of class "ANY": If the test method supports it, can be used to specify a subset of category ids to include in the test instead of all possible category ids.

**categoryName:** A string describing the category. Usually set automatically by subclasses. For example "GO".

**pvalueCutoff:** The p-value to use as a cutoff for significance for testing methods that require it. This value will also be passed on to the result instance and used for display and counting of significant results. The default is 0.01.

**testDirection:** A string indicating whether the test should be for overrepresentation ("over") or underrepresentation ("under").

**datPkg:** Object of class "DatPkg" used to assist with dispatch based on type of annotation data available.

## Extends

Class "[HyperGParams](#)", directly.

## Methods

No methods defined with class "ChrMapHyperGParams" in the signature.

## Author(s)

Seth Falcon

## Examples

```
showClass("ChrMapHyperGParams")
```

---

ChrMapHyperGResult-class

*Class "ChrMapHyperGResult"*

---

## Description

This class represents the results of a Hypergeometric test for over-representation of genes in a selected gene list in the chromosome band annotation. The `hyperGTest` function returns an instance of `ChrMapHyperGResult` when given a parameter object of class `ChrMapHyperGParams`. For details on accessing the results, see [HyperGResult-accessors](#).

## Objects from the Class

Objects can be created by calls of the form `new("ChrMapHyperGResult", ...)`.

**Slots**

**pvalue.order:** Object of class "integer" that gives the order of the p-values.

**conditional:** Object of class "logical" is a flag indicating whether or not this result is from a conditional analysis.

**chrGraph:** Object of class "graph". The nodes are the chromosome bands with edges representing the tree structure of the bands. Each node has a "geneIds" attribute that gives the gene IDs annotated at that band.

**annotation:** A string giving the name of the chip annotation data package used.

**geneIds:** Object of class "ANY": the input vector of gene identifiers intersected with the universe of gene identifiers used in the computation. The class of this slot is specified as "ANY" because gene IDs may be integer or character vectors depending on the annotation package.

**testName:** A string identifying the testing method used.

**pvalueCutoff:** Numeric value used as a p-value cutoff. Used by the show method to count number of significant terms.

**testDirection:** Object of class "character" indicating whether the test was for over-representation ("over") or under-representation ("under").

**Extends**

Class "[HyperGResultBase](#)", directly.

**Methods**

See [HyperGResult-accessors](#).

**Author(s)**

Seth Falcon

**Examples**

```
showClass("ChrMapHyperGResult")
## For details on accessing the results:
##   help("HyperGResult-accessors")
```

---

ChrMapLinearMParams-class

*Class "ChrMapLinearMParams"*

---

**Description**

This class encapsulates parameters needed for testing systematic variations in some gene-level statistic by chromosome bands using [linearMTest](#).

## Objects from the Class

Objects can be created by calls of the form `new("ChrMapLinearMParams", ...)`.

## Slots

**graph:** Object of class "graph". The nodes are the chromosome bands and the edges describe the tree structure of the bands. Each node has a "geneIds" node attributes (see `nodeData`) which contains a vector of gene IDs annotated at the given band.

**conditional:** Object of class "logical", indicating whether the test performed should be a conditional test.

**gsc:** The [GeneSetCollection](#) object grouping the gene ids into sets.

**geneStats:** Named vector of class "numeric", giving the gene-level statistics to be used in the tests.

**universeGeneIds:** Object of class "ANY": A vector of gene ids defining a subset of the gene ids on the chip that will be used as the universe for the hypergeometric calculation. If this is NULL or has length zero, then all gene ids on the chip will be used.

**annotation:** A string giving the name of the annotation data package for the chip used to generate the data.

**datPkg:** Object of class "DatPkg" used to assist with dispatch based on type of annotation data available.

**categorySubsetIds:** Object of class "ANY": If the test method supports it, can be used to specify a subset of category ids to include in the test instead of all possible category ids.

**categoryName:** A string describing the category. Usually set automatically by subclasses. For example "GO".

**pvalueCutoff:** The p-value to use as a cutoff for significance for testing methods that require it. This value will also be passed on to the result instance and used for display and counting of significant results. The default is 0.01.

**minSize:** An integer giving a minimum size for a gene set for it to be tested. The default is 5.

**testDirection:** A string indicating whether the test should test for systematic increase ("up") or decrease ("down") in the `geneStats` values within a gene set relative to the remaining genes.

## Extends

Class "[LinearMParams](#)", directly.

## Author(s)

Deepayan Sarkar

## See Also

[linearMTest](#)

## Examples

```
showClass("ChrMapLinearMParams")
```

---

ChrMapLinearMResult-class

*Class "ChrMapLinearMResult"*


---

## Description

This class represents the results of a linear model-based test for systematic changes in a per-gene statistic by chromosome band annotation. The [linearMTest](#) function returns an instance of ChrMapLinearMResult when given a parameter object of class ChrMapLinearMParams. Most slots can be queried using accessors.

## Objects from the Class

Objects can be created by calls of the form `new("ChrMapLinearMResult", ...)`, but is more commonly created by calling [linearMTest](#)

## Slots

**pvalues:** Object of class "numeric", with the p-values for each term.  
**pvalue.order:** Object of class "integer", the order vector (increasing) for the p-values.  
**effectSize:** Object of class "numeric", with the effect size for each term.  
**annotation:** Object of class "character" ~~  
**geneIds:** Object of class "ANY" ~~  
**testName:** Object of class "character" ~~  
**pvalueCutoff:** Object of class "numeric" ~~  
**minSize:** Object of class "integer" ~~  
**testDirection:** Object of class "character" ~~  
**conditional:** Object of class "logical" ~~  
**graph:** Object of class "graph" ~~  
**gsc:** Object of class "GeneSetCollection" ~~

## Extends

Class "[LinearMResult](#)", directly.  
 Class "[LinearMResultBase](#)", by class "LinearMResult", distance 2.

## Methods

None

## Author(s)

Deepayan Sarkar, Michael Lawrence

**See Also**

[linearMTest](#), [ChrMapLinearMParams](#), [LinearMResult](#), [LinearMResultBase](#),

**Examples**

```
showClass("ChrMapLinearMResult")
```

---

DatPkg-class	<i>Class "DatPkg"</i>
--------------	-----------------------

---

**Description**

DatPkg is a VIRTUAL class for representing annotation data packages.

AffyDatPkg is a subclass of DatPkg used to represent standard annotation data packages that follow the format of Affymetrix expression array annotation.

YeastDatPkg is a subclass of DatPkg used to represent the annotation data packages for yeast. The yeast chip packages are based on sgd and are internally different from the AffyDataPkg conforming packages.

Org.XX.egDatPkg is a subclass of DatPkg used to represent the org.\*.eg.db organism-level Entrez Gene based annotation data packages.

GeneSetCollectionDatPkg is a subclass of DatPkg used to represent annotations in the form of GeneSetCollection objects which are not based on any annotation packages but are instead derived from custom (user supplied) annotations.

**Objects from the Class**

A virtual Class: No objects may be created from it.

Given the name of an annotation data package, DatPkgFactory can be used to create an appropriate DatPkg subclass.

**Slots**

**name** A string giving the name of the annotation data package.

**Methods**

See `showMethods(classes="DatPkg")`.

The set of methods, ID2EntrezID map between the standard IDs for an organism, or Chip and EntrezIDs, typically to give a way to get the GO terms. Different organisms, such as *S. cerevisiae* and *A. thaliana* have their own internal IDs, so we need specialized methods for them.

**Author(s)**

Seth Falcon

**Examples**

```
DatPkgFactory("hgu95av2")
## Not run:
DatPkgFactory("org.Sc.sgd")
DatPkgFactory("org.Hs.eg.db")
DatPkgFactory("ag")

## End(Not run)
```

---

effectSize*Extract estimated effect sizes*

---

**Description**

This function extracts estimated effect sizes from the results of a linear model-based gene-set / category enrichment test.

**Usage**

```
effectSize(r)
```

**Arguments**

**r**                      The results of the test

**Value**

A numeric vector.

**Author(s)**

Deepayan Sarkar

**See Also**

```
linkS4class{LinearMResult}
```

exampleLevels

*Display a sample node from each level of a ChrBandTree object***Description**

The "levels" of a chromosome band tree represented by a ChrBandTree object are the sets of nodes with a given path length to the root node. This function displays the available levels along with an example node from each level.

**Usage**

```
exampleLevels(g)
```

**Arguments**

`g`                      A ChrBandTree object

**Value**

A list with an element for each level. The names of the list are the levels. Each element is an example of a node from the given level.

**Author(s)**

S. Falcon

findAMstats

*Compute per category summary statistics***Description**

For a given incidence matrix, Amat, compute some per category statistics.

**Usage**

```
findAMstats(Amat, tstats)
```

**Arguments**

`Amat`                      An incidence matrix, with categories as the rows and probes as the columns.  
`tstats`                    A vector of per probe test statistics (should be the same length as `ncol(Amat)`).

**Details**

Simple summary statistics are computed, such as the row sums and the vector of per category sums of the test statistics, `tstats`.

**Value**

A list with components,

eDE	per category sums of the test statistics
lens	row sums of Amat

**Author(s)**

R. Gentleman

**See Also**

[applyByCategory](#)

**Examples**

```
ts = rnorm(100)
Am = matrix(sample(c(0,1), 1000, replace=TRUE), ncol=100)
findAMstats(Am, ts)
```

---

getPathNames

*A function to print pathway names given their numeric ID.*

---

**Description**

Given a KEGG pathway ID this function returns the character name of the pathway.

**Usage**

```
getPathNames(iPW)
```

**Arguments**

iPW	A vector of KEGG pathway IDs.
-----	-------------------------------

**Details**

This function simply does a look up in KEGGPATHID2NAME and returns a list of the pathway names. Possible extensions would be to extend it to work with the cMAP library as well.

**Value**

A list of pathway names.

**Author(s)**

R. Gentleman

**See Also**[KEGGPATHID2NAME](#)**Examples**

```
nms = "00031"
getPathNames(nms)
```

---

GOHyperGParams-class    *Class "GOHyperGParams"*


---

**Description**

A parameter class for representing all parameters needed for running the hyperGTest method with one of the GO ontologies (BP, CC, MF) as the category.

**Objects from the Class**

Objects can be created by calls of the form `new("GOHyperGParams", ...)`.

**Slots**

**ontology:** A string specifying the GO ontology to use. Must be one of "BP", "CC", or "MF".

**conditional:** A logical indicating whether the calculation should condition on the GO structure.

**geneIds:** Object of class "ANY": A vector of gene identifiers. Numeric and character vectors are probably the only things that make sense. These are the gene ids for the selected gene set.

**universeGeneIds:** Object of class "ANY": A vector of gene ids in the same format as geneIds defining a subset of the gene ids on the chip that will be used as the universe for the hypergeometric calculation. If this is NULL or has length zero, then all gene ids on the chip will be used.

**annotation:** A string giving the name of the annotation data package for the chip used to generate the data.

**categorySubsetIds:** Object of class "ANY": If the test method supports it, can be used to specify a subset of category ids to include in the test instead of all possible category ids.

**categoryName:** A string describing the category. Usually set automatically by subclasses. For example "GO".

**datPkg:** Holds a DatPkg object which is of a particular type that in turn varies with the kind of annotation package this is.

**pvalueCutoff:** A numeric values between zero and one used as a p-value cutoff for p-values generated by the Hypergeometric test. When the test being performed is non-conditional, this is only used as a default value for printing and summarizing the results. For a conditional analysis, the cutoff is used during the computation to determine perform the conditioning: child terms with a p-value less than pvalueCutoff are conditioned out of the test for their parent term.

**testDirection:** A string which can be either "over" or "under". This determines whether the test performed detects over or under represented GO terms.

**Extends**

Class "HyperGParams", directly.

**Methods**

`hyperGTest(p)` Perform hypergeometric tests to assess overrepresentation of category ids in the gene set. See the documentation for the generic function for details. This method must be called with a proper subclass of HyperGParams.

`ontology(p), ontology(p) <- value` Accessors for the GO ontology. When setting, value should be one of "BP", "CC", or "MF".

`conditional(p), conditional(p) <- value` Accessors for the conditional flag. When setting, value must be TRUE or FALSE.

**Author(s)**

S. Falcon

**See Also**

[HyperGResult-class](#) [GOHyperGParams-class](#) [hyperGTest](#)

---

GSEAGOHyperGParams	<i>Helper function for constructing a GOHyperGParams objects or KEGGHyperGParams objects from a GeneSetCollection</i>
--------------------	---

---

**Description**

Helps to create A parameter class for representing all parameters needed for running the hyperGTest method. If it is a GOHyperGParams object, being made, then with one of the GO ontologies (BP, CC, MF) as the category. This function will construct the parameter object from a GeneSetCollection object and if necessary will also try to check to make sure that the object is based on a GO2ALL mapping.

**Usage**

```
GSEAGOHyperGParams(name, geneSetCollection, geneIds, universeGeneIds,
ontology, pvalueCutoff, conditional, testDirection, ...)
```

```
GSEAKEGGHyperGParams(name, geneSetCollection, geneIds, universeGeneIds,
pvalueCutoff, testDirection, ...)
```

**Arguments**

name	String specifying name of the GeneSetCollection.
geneSetCollection	A GeneSetCollection Object. If a GOHyperGParams object is sought, then this GeneSetCollection should be based on a GO2ALLFrame object and so the idType of that GeneSetCollection should be GOAllFrameIdentifier. If a KEGGHyperGParams object is sought then a GeneSetCollection based on a KEGGFrame object should be used and the idType will be a KEGGFrameIdentifier.
geneIds	Object of class "ANY": A vector of gene identifiers. Numeric and character vectors are probably the only things that make sense. These are the gene ids for the selected gene set.
universeGeneIds	Object of class "ANY": A vector of gene ids in the same format as geneIds defining a subset of the gene ids on the chip that will be used as the universe for the hypergeometric calculation. If this is NULL or has length zero, then all gene ids on the chip will be used.
ontology	A string specifying the GO ontology to use. Must be one of "BP", "CC", or "MF". (used with GO only)
pvalueCutoff	A numeric values between zero and one used as a p-value cutoff for p-values generated by the Hypergeometric test. When the test being performed is non-conditional, this is only used as a default value for printing and summarizing the results. For a conditional analysis, the cutoff is used during the computation to determine perform the conditioning: child terms with a p-value less than pvalueCutoff are conditioned out of the test for their parent term.
conditional	A logical indicating whether the calculation should condition on the GO structure. (GO only)
testDirection	A string which can be either "over" or "under". This determines whether the test performed detects over or under represented GO terms.
...	optional arguments to configure the GOHyperGParams object.

**Author(s)**

M. Carlson

**See Also**[HyperGResult-class GOHyperGParams-class hyperGTest](#)

gseattperm

*Permutation p-values for GSEA***Description**

This function performs GSEA computations and returns p-values for each gene set based on repeated permutation of the phenotype labels.

**Usage**

```
gseattperm(eset, fac, mat, nperm)
```

**Arguments**

eset	An ExpressionSet object
fac	A factor identifying the phenotypes in eset. Usually, this will be one of the columns in the phenotype data associated with eset.
mat	A 0/1 incidence matrix with each row representing a gene set and each column representing a gene. A 1 indicates membership of a gene in a gene set.
nperm	Number of permutations to test to build the reference distribution.

**Details**

The t-statistic is used (via rowttests) to test for a difference in means between the phenotypes determined by fac within each gene set (given as a row of mat).

A reference distribution for these statistics is established by permuting fac and repeating the test B times.

**Value**

A matrix with the same number of rows as mat and two columns, "Lower" and "Upper". The "Lower" ("Upper") column gives the probability of seeing a t-statistic smaller (larger) than the observed.

**Author(s)**

Seth Falcon

**Examples**

```
## This example uses a random sample of probesets and a randomly
## generated category matrix. The results, therefore, are not
## meaningful, but the code demonstrates how to use gseattperm without
## requiring any expensive computations.

## Obtain an ExpressionSet with two types of samples (mol.biol)
haveALL <- require("ALL")
if (haveALL) {
  data(ALL)
  set.seed(0xabcd)
  rndIdx <- sample(1:nrow(ALL), 500)
  Bcell <- grep("^B", as.character(ALL$BT))
  typeNames <- c("NEG", "BCR/ABL")
  bcrAblOrNegIdx <- which(as.character(ALL$mol.biol) %in% typeNames)
  s <- ALL[rndIdx, intersect(Bcell, bcrAblOrNegIdx)]
  s$mol.biol <- factor(s$mol.biol)

  ## Generate a random category matrix
```

```

nCats <- 100
set.seed(0xdcba)
rndCatMat <- matrix(sample(c(0L, 1L), replace=TRUE),
                     nrow=nCats, ncol=nrow(s),
                     dimnames=list(
                       paste("c", 1:nCats, sep=""),
                       featureNames(s)))

## Demonstrate use of gseattperm
N <- 10
pvals <- gseattperm(s, s$mol.biol, rndCatMat, N)
pvals[1:5, ]
}

```

hyperg

*Hypergeometric (gene set enrichment) tests on character vectors.***Description**

This function performs a hypergeometric test for over- or under-representation of significant ‘genes’ amongst those assayed in a universe of genes. It provides an interface based on character vectors of identifying member of gene sets and the gene universe.

**Usage**

```
hyperg(assayed, significant, universe,
       representation = c("over", "under"), ...)
```

**Arguments**

assayed	A vector of assayed genes (or other identifiers). assayed may be a character vector (defining a single gene set) or list of character vectors (defining a collection of gene sets).
significant	A vector of assayed genes that were differentially expressed. If assayed is a character vector, then significant must also be a character vector; likewise when assayed is a list.
universe	A character vector defining the universe of genes.
representation	Either “over” or “under”, to indicate testing for over- or under-representation, respectively, of differentially expressed genes.
...	Additional arguments, unused.

**Value**

When invoked with a character vector of assayed genes, a named numeric vector providing the input values, P-value, odds ratio, and expected number of significantly expressed genes.

When invoked with a list of character vectors of assayed genes, a data frame with columns of input values, P-value, odds ratio, and expected number of significantly expressed genes.

**Author(s)**

Martin Morgan [mtmorgan@fhcrc.org](mailto:mtmorgan@fhcrc.org) with contributions from Paul Shannon.

**See Also**

[hyperGTest](#) for convenience functions using Bioconductor annotation resources such as GO.db.

**Examples**

```
set.seed(123)

## artificial sets -- affy probes grouped by protein family
library(hgu95av2.db)
map <- select(hgu95av2.db, keys(hgu95av2.db), "PFAM")
sets <- Filter(function(x) length(x) >= 10, split(map$PROBEID, map$PFAM))

universe <- unlist(sets, use.names=FALSE)
siggenes <- sample(universe, length(universe) / 20) ## simulate
sigsets <- Map(function(x, y) x[x %in% y], sets, MoreArgs=list(y=siggenes))

result <- hyperg(sets, sigsets, universe)
head(result)
```

---

HyperGParams-class	<i>Class "HyperGParams"</i>
--------------------	-----------------------------

---

**Description**

An abstract (VIRTUAL) parameter class for representing all parameters needed by a method specializing the hyperGTest generic. You should only use subclasses of this class directly.

**Objects from the Class**

Objects of this class cannot be instantiated directly.

**Slots**

**geneIds:** Object of class "ANY": A vector of gene identifiers. Numeric and character vectors are probably the only things that make sense. These are the gene ids for the selected gene set.

**universeGeneIds:** Object of class "ANY": A vector of gene ids in the same format as geneIds defining a subset of the gene ids on the chip that will be used as the universe for the hypergeometric calculation. If this is NULL or has length zero, then all gene ids on the chip will be used.

**annotation:** A string giving the name of the annotation data package for the chip used to generate the data.

**categorySubsetIds:** Object of class "ANY": If the test method supports it, can be used to specify a subset of category ids to include in the test instead of all possible category ids.

**categoryName:** A string describing the category. Usually set automatically by subclasses. For example "GO".

**pvalueCutoff:** The p-value to use as a cutoff for significance for testing methods that require it. This value will also be passed on to the result instance and used for display and counting of significant results. The default is 0.01.

**testDirection:** A string indicating whether the test should be for overrepresentation ("over") or underrepresentation ("under").

**datPkg:** Holds a DatPkg object which is of a particular type that in turn varies with the kind of annotation package this is.

## Methods

**hyperGTest** signature(p = "HyperGParams"): Perform hypergeometric tests to assess overrepresentation of category ids in the gene set. See the documentation for the generic function for details. This method must be called with a proper subclass of HyperGParams.

**geneIds(object), geneIds(object) <- value** Accessors for the gene identifiers that will be used as the selected gene list.

**codeannotation(object)** Accessor for annotation. If you want to change the annotation for an existing instance, use the replacement form.

**ontology(object)** Accessor for GO ontology.

**pvalueCutoff(r), pvalueCutoff(r) <- value** Accessor for the p-value cutoff. When setting, value should be a numeric value between zero and one.

**testDirection** Accessor for the test direction. When setting, value must be either "over" or "under".

**universeGeneIds(r)** accessor for vector of gene identifiers.

## Author(s)

S. Falcon

## See Also

[HyperGResult-class](#) [GOHyperGParams-class](#) [KEGGHyperGParams-class](#) [hyperGTest](#)

---

HyperGResult-accessors

*Accessors for HyperGResult Objects*

---

## Description

This manual page documents generic functions for extracting data from the result object returned from a call to `hyperGTest`. The result object will be a subclass of `HyperGResultBase`. Methods apply to all result object classes unless otherwise noted.

**Usage**

```

pvalues(r)
oddsRatios(r)
expectedCounts(r)

geneCounts(r)
universeCounts(r)
universeMappedCount(r)
geneMappedCount(r)

geneIds(object, ...)
geneIdUniverse(r, cond = TRUE)
geneIdsByCategory(r, catids = NULL)
sigCategories(r, p)

## R CMD check doesnt like these
## annotation(r)
## description(r)

testName(r)
pvalueCutoff(r)
testDirection(r)

chrGraph(r)

```

**Arguments**

<code>r, object</code>	An instance of a subclass of <code>HyperGResultBase</code> .
<code>catids</code>	A character vector of category identifiers.
<code>p</code>	Numeric p-value used as a cutoff for selecting a subset of the result.
<code>cond</code>	A logical value indicating whether to return conditional results for a conditional test. The default is <code>TRUE</code> . For non-conditional results, this argument is ignored.
<code>...</code>	Additional arguments that may be used by specializing methods.

**Accessor Methods (Generic Functions)**

**geneCounts** returns an "integer" vector: for each category term tested, the number of genes from the gene set that are annotated at the term.

**pvalues** returns a "numeric" vector: the ordered p-values for each category term tested.

**universeCounts** returns an "integer" vector: for each category term tested, the number of genes from the gene universe that are annotated at the term.

**universeMappedCount** returns an "integer" vector of length one giving the size of the gene universe set.

**expectedCounts** returns a "numeric" vector giving the expected number of genes in the selected gene list to be found at each tested category term. These values may surprise you if you forget that your gene list and gene universe might have had to undergo further filtering to ensure that each gene has been labeled by at least one GO term.

**oddsRatios** returns a "numeric" vector giving the odds ratio for each category term tested.

**annotation** returns the name of the annotation data package used.

**geneIds** returns the input vector of gene identifiers intersected with the universe of gene identifiers used in the computation.

**geneIdUniverse** returns a list named by the tested categories. Each element of the list is a vector of gene identifiers (from the gene universe) annotated at the corresponding category term.

**geneIdsByCategory** returns a list similar to `geneIdUniverse`, but each vector of gene IDs is intersected with the list of selected gene IDs from `geneIds`. The result is the selected gene IDs annotated at each category.

**sigCategories** returns a character vector of category identifiers with a significant p-value. If argument `p` is missing, then the cutoff obtained from `pvalueCutoff(r)` will be used.

**geneMappedCount** returns the size of the selected gene set used in the computation. This is simply `length(geneIds(obj))`.

**pvalueCutoff** accessor for the `pvalueCutoff` slot.

**testDirection** accessor for the `testDirection` slot. Contains a string indicating whether the test was for "over" or "under" representation of the categories.

**description** returns a character string description of the test result.

**testName** returns a string describing the testing method used.

**summary** returns a `data.frame` summarizing the test result. Optional arguments `pvalue` and `categorySize` allow specification of maximum p-value and minimum categorySize, respectively. The data frame contains the `GOID`, `Pvalue`, `OddsRatio`, `ExpCount`, `Count`, and `Size`. `ExpCount` is the expected count and the `Count` is how many instances of that term were actually observed in your gene list while the `Size` is the number that could have been found in your gene list if every instance had turned up. Values like the `ExpCount` and the `Size` are going to be affected by what is included in the gene universe as well as by whether or not it was a conditional test.

**htmlReport** writes an HTML version of the table produced by the `summary` method. The first argument should be a `HyperGResult` instance (or subclass). The path of a file to write the report to can be specified using the `file` argument. The default is `file=""` which will cause the report to be printed to the screen. If you wish to create a single report comprising multiple results you can set `append=TRUE`. The default is `FALSE` (overwrite pre-existing report file). You can specify a string to use as an identifier for each table by providing a value for the `label` argument. The number of digits displayed in numerical columns can be controlled using `digits` (defaults to 3). The `summary` method is called on the `HyperGResult` instance to generate a data frame that is transformed to HTML. You can pass additional arguments to the `summary` method which is used to generate the data frame that is transformed to HTML by specifying a named list using `summary.args`.

## Author(s)

Seth Falcon

**See Also**

[hyperGTest](#) [HyperGResult-class](#) [HyperGParams-class](#) [GOHyperGParams-class](#) [KEGGHyperGParams-class](#)

**Examples**

```
## Note that more in-depth examples can be found in the G0stats
## vignette (Hypergeometric tests using G0stats).
library("hgu95av2.db")
library("annotate")

probinds <- ls(hgu95av2GENENAME)[1:300]
## Select for probeids that have PFAM ids
hasPFAM <- sapply(mget(probinds, hgu95av2PFAM), function(ids)
  if(!is.na(ids) && length(ids) > 1) TRUE else FALSE)
probinds <- probinds[hasPFAM]
## get unique Entrez Gene IDs
probinds <- unique(getEG(probinds, "hgu95av2"))
## Now do the same for the universe
univ <- ls(hgu95av2GENENAME)
univHasPFAM <- sapply(mget(univ, hgu95av2PFAM), function(ids)
  if(!is.na(ids) && length(ids) > 1) TRUE else
  FALSE)
univ <- univ[univHasPFAM]
univ <- unique(getEG(univ, "hgu95av2"))

p <- new("PFAMHyperGParams", geneIds=probinds, universeGeneIds=univ,
  annotation="hgu95av2")
## this takes a while...
if(interactive()){
  hypt <- hyperGTest(p)
  summary(hypt)
  htmlReport(hypt, file="temp.html", summary.args=list("htmlLinks"=TRUE))
}
```

---

HyperGResult-class      *Class "HyperGResult"*

---

**Description**

This class represents the results of a test for over-representation of categories among genes in a selected gene set based upon the Hypergeometric distribution. The `hyperGTest` generic function returns an instance of the `HyperGResult` class. For details on accessing the results, see [HyperGResult-accessors](#).

**Objects from the Class**

Objects can be created by calls of the form `new("HyperGResult", ...)`.

**Slots**

**pvalues:** "numeric" vector: the ordered p-values for each category term tested.

**catToGeneId:** Object of class "list". The names of the list are category IDs. Each element is a vector of gene IDs annotated at the given category ID and in the specified gene universe.

**annotation:** A string giving the name of the chip annotation data package used.

**geneIds:** Object of class "ANY": the input vector of gene identifiers intersected with the universe of gene identifiers used in the computation. The class of this slot is specified as "ANY" because gene IDs may be integer or character vectors depending on the annotation package.

**testName:** A string identifying the testing method used.

**pvalueCutoff:** Numeric value used as a p-value cutoff. Used by the show method to count number of significant terms.

**testDirection:** A string indicating whether the test should be for overrepresentation ("over") or underrepresentation ("under").

**oddsRatios** a "numeric" vector giving the odds ratio for each category term tested.

**expectedCounts** a "numeric" vector giving the expected number of genes in the selected gene list to be found at each tested category term.

**Extends**

Class "HyperGResultBase", directly.

**Methods**

See [HyperGResult-accessors](#).

**Author(s)**

Seth Falcon

**See Also**

[HyperGResultBase-class](#) [GOHyperGResult-class](#) [HyperGResult-accessors](#)

---

HyperGResultBase-class

*Class "HyperGResultBase"*

---

**Description**

This VIRTUAL class represents common elements of the return values of generic functions like `hyperGTest`. All subclasses are intended to implement the accessor functions documented at [HyperGResult-accessors](#).

**Objects from the Class**

A virtual Class: No objects may be created from it.

**Slots**

**annotation:** Object of class "character" giving the name of the annotation data package used.

**geneIds:** Object of class "ANY" (usually a character vector, but sometimes an integer vector). The input vector of gene identifiers intersected with the universe of gene identifiers used in the computation.

**testName:** Object of class "character" identifying the testing method used.

**pvalueCutoff:** Numeric value used by the testing method as a p-value cutoff. Not all testing methods use this. Also used by the show method to count number of significant terms.

**testDirection:** A string indicating whether the test performed was for overrepresentation ("over") or underrepresentation("under").

**Methods**

See [HyperGResult-accessors](#).

**Author(s)**

Seth Falcon

**See Also**

[HyperGResult-class](#) [GOHyperGResult-class](#) [HyperGResult-accessors](#)

---

hyperGTest

---

*Hypergeometric Test for association of categories and genes*


---

**Description**

Given a subclass of HyperGParams, compute Hypergeometric p-values for over or under-representation of each term in the specified category among the specified gene set.

**Usage**

```
hyperGTest(p)
```

**Arguments**

**p** An instance of a subclass of HyperGParams. This parameter object determines the category of interest (e.g., GO or KEGG) as well as the gene set.

**Details**

The gene identifiers in the geneIds slot of p define the selected set of genes. The universe of gene ids is determined by the chip annotation found in the annotation slot of p. Both the selected genes and the universe are reduced by removing identifiers that do not have any annotations in the specified category.

For each term in the specified category that has at least one annotation in the selected gene set, we determine how many of its annotations are in the universe set and how many are in the selected set. With these counts we perform a Hypergeometric test using phyper. This is equivalent to using Fisher's exact test.

It is important that the correct chip annotation data package be identified as it determines the universe of gene identifiers and is often used to determine the mapping between the category term and the gene identifiers.

For *S. cerevisiae* if the annotation slot of p is set to "org.Sc.sgd" then comparisons and statistics are computed using common names and are with respect to all genes annotated in the *S. cerevisiae* genome not with respect to any microarray chip. This will *not* be the right thing to do if you are working with a yeast microarray.

**Value**

A HyperGResult instance.

**Implementation Notes**

In most cases, the provided method with signature matching any subclass of HyperGParams is all that will be needed. This method follows a template pattern. To add support for a new FOO category type, a developer would need to create a FooHyperGParams subclass and then define two methods specialized to the new subclass that get called from inside hyperGTest: universeBuilder and categoryToEntrezBuilder.

**Author(s)**

S. Falcon

**See Also**

[HyperGResult-class](#) [HyperGParams-class](#) [GOHyperGParams-class](#) [KEGGHyperGParams-class](#)

---

KEGGHyperGParams-class

*Class "KEGGHyperGParams" and "PFAMHyperGParams"*

---

**Description**

Parameter classes for representing all parameters needed for running the hyperGTest method with KEGG or PFAM as the category.

## Objects from the Class

Objects can be created by calls of the form `new("KEGGHyperGParams", ...)` or `new("PFAMHyperGParams", ...)`.

## Slots

**geneIds:** Object of class "ANY": A vector of gene identifiers. Numeric and character vectors are probably the only things that make sense. These are the gene ids for the selected gene set.

**universeGeneIds:** Object of class "ANY": A vector of gene ids in the same format as **geneIds** defining a subset of the gene ids on the chip that will be used as the universe for the hypergeometric calculation. If this is `NULL` or has length zero, then all gene ids on the chip will be used.

**annotation:** A string giving the name of the annotation data package for the chip used to generate the data.

**categorySubsetIds:** Object of class "ANY": If the test method supports it, can be used to specify a subset of category ids to include in the test instead of all possible category ids.

**categoryName:** A string describing the category. This will be automatically set to "KEGG" or "PFAM" via the class's prototype.

**pvalueCutoff:** The p-value to use as a cutoff for significance for testing methods that require it. This value will also be passed on to the result instance and used for display and counting of significant results. The default is 0.01.

**testDirection:** A string indicating whether the test should be for overrepresentation ("over") or underrepresentation ("under").

## Extends

Class "HyperGParams", directly.

## Methods

**hyperGTest** signature(`p = "HyperGParams"`): Perform hypergeometric tests to assess overrepresentation of category ids in the gene set. See the documentation for the generic function for details. This method must be called with a proper subclass of `HyperGParams`.

## Author(s)

S. Falcon

## See Also

[HyperGResult-class](#) [GOHyperGParams-class](#) [hyperGTest](#)

---

LinearMParams-class      *Class "LinearMParams"*


---

## Description

A parameter class for representing all parameters needed by a method specializing the [linearMTest](#) generic.

## Objects from the Class

Objects can be created by calls of the form `new("LinearMParams", ...)`.

## Slots

**geneStats:** Named vector of class "numeric", giving the gene-level statistics to be used in the tests. The names should correspond to the gene identifiers in gsc.

**universeGeneIds:** Object of class "ANY": A vector of gene ids defining a subset of the gene ids on the chip that will be used as the universe for the hypergeometric calculation. If this is NULL or has length zero, then all gene ids on the chip will be used. Currently this parameter is ignored by the base `linearMTest` method.

**annotation:** A string giving the name of the annotation data package for the chip used to generate the data.

**datPkg:** Object of class "DatPkg" used to assist with dispatch based on type of annotation data available. Currently this parameter is ignored by the base `linearMTest` method.

**categorySubsetIds:** Object of class "ANY": If the test method supports it, can be used to specify a subset of category ids to include in the test instead of all possible category ids. Currently this parameter is ignored by the base `linearMTest` method.

**categoryName:** A string describing the category. Usually set automatically by subclasses. For example "ChrMap".

**pvalueCutoff:** The p-value to use as a cutoff for significance for testing methods that require it. This value will also be passed on to the result instance and used for display and counting of significant results. The default is 0.01.

**minSize:** An integer giving a minimum size for a gene set for it to be tested. The default is 5.

**testDirection:** A string indicating whether the test should test for systematic increase ("up") or decrease ("down") in the `geneStats` values within a gene set relative to the remaining genes.

**graph:** The [graph](#) object indicating the hierarchical relationship among terms of the ontology or other grouping.

**conditional:** A logical indicating whether conditional tests should be performed. This tests whether a term is still significant even when including its sub-terms in the model.

**gsc:** The [GeneSetCollection](#) object grouping the gene ids into sets.

## Methods

These are accessor methods for the various parameter slots:

```
annotation<- signature(object = "LinearMParams", value = "character"): ...
annotation signature(object = "LinearMParams"): ...
categoryName signature(r = "LinearMParams"): ...
conditional signature(r = "LinearMParams"): ...
geneIds<- signature(object = "LinearMParams"): ...
geneIds signature(object = "LinearMParams"): ...
pvalueCutoff<- signature(r = "LinearMParams"): ...
pvalueCutoff signature(r = "LinearMParams"): ...
show signature(object = "LinearMParams"): ...
testDirection<- signature(r = "LinearMParams"): ...
testDirection signature(r = "LinearMParams"): ...
conditional<- signature(r = "LinearMParams"): ...
conditional signature(r = "LinearMParams"): ...
universeGeneIds signature(r = "LinearMParams"): ...
```

## Author(s)

Deepayan Sarkar, Michael Lawrence

## See Also

See [linearMTest](#) for examples. [ChrMapLinearMParams](#) is a specialization of this class for chromosome maps.

---

LinearMResult-class	<i>Class "LinearMResult"</i>
---------------------	------------------------------

---

## Description

This class represents the results of a test for systematic change in some gene-level statistic by gene sets. The `linearMTest` generic function returns an instance of the `LinearMResult` class.

## Objects from the Class

Objects can be created by calls of the form `new("LinearMResult", ...)`, but is more commonly created using a call to [linearMTest](#).

**Slots**

**pvalues:** Object of class "numeric", with the p-values for each term.  
**pvalue.order:** Object of class "integer", the order vector (increasing) for the p-values.  
**effectSize:** Object of class "numeric", with the effect size for each term.  
**annotation:** Object of class "character" ~~  
**geneIds:** Object of class "ANY" ~~  
**testName:** Object of class "character" ~~  
**pvalueCutoff:** Object of class "numeric" ~~  
**minSize:** Object of class "integer" ~~  
**testDirection:** Object of class "character" ~~  
**conditional:** Object of class "logical" ~~  
**graph:** Object of class "graph" ~~  
**gsc:** Object of class "GeneSetCollection" ~~

**Extends**

Class "[LinearMResultBase](#)", directly.

**Methods**

**effectSize** signature(r = "LinearMResult"): ...  
**pvalues** signature(r = "LinearMResult"): ...  
**summary** signature(r = "LinearMResult"): returns a data.frame with a row for each gene set tested the following columns: ID, Pvalue, Effect size, Size (number of members), Conditional (whether the test used the conditional test), and TestDirection (for up or down).

**Author(s)**

Deepayan Sarkar, Michael Lawrence

**See Also**

[linearMTest](#)

**Examples**

```
showClass("LinearMResult")
```

---

LinearMResultBase-class

Class "LinearMResultBase"

---

## Description

This VIRTUAL class represents common elements of the return values of generic functions like `linearMTest`. These elements are essentially those that are passed through from the input parameters. See [LinearMResult](#) for a concrete result class with the basic outputs.

## Objects from the Class

A virtual Class: No objects may be created from it.

## Slots

All of these slots correspond to slots in the [LinearMParams](#) class.

**annotation:** Object of class "character" ~~

**geneIds:** Object of class "ANY" ~~

**testName:** Object of class "character" ~~

**pvalueCutoff:** Object of class "numeric" ~~

**minSize:** Object of class "integer" ~~

**testDirection:** Object of class "character" ~~

**conditional:** Object of class "logical" ~~

**graph:** Object of class "graph" ~~

**gsc:** Object of class "GeneSetCollection" ~~

## Methods

**annotation** signature(object = "LinearMResultBase"): ...

**conditional** signature(r = "LinearMResultBase"): ...

**description** signature(object = "LinearMResultBase"): ...

**geneIdsByCategory** signature(object = "LinearMResultBase"): ...

**geneIds** signature(object = "LinearMResultBase"): ...

**geneIdUniverse** signature(r = "LinearMResultBase"): ...

**geneMappedCount** signature(r = "LinearMResultBase"): ...

**pvalueCutoff** signature(r = "LinearMResultBase"): ...

**show** signature(object = "LinearMResultBase"): ...

**sigCategories** signature(r = "LinearMResultBase"): ...

**summary** signature(object = "LinearMResultBase"): ...

**testDirection** signature(r = "LinearMResultBase"): ...

**conditional** signature(object = "LinearMResultBase"): ...  
**testName** signature(r = "LinearMResultBase"): ...  
**universeCounts** signature(r = "LinearMResultBase"): ...  
**universeMappedCount** signature(r = "LinearMResultBase"): ...

**Author(s)**

Deepayan Sarkar, Michael Lawrence

**See Also**

[LinearMResult](#), [LinearMParams](#), [linearMTest](#)

---

linearMTest	<i>A linear model-based test to detect enrichment of unusual genes in categories</i>
-------------	--

---

**Description**

Given a subclass of LinearMParams, compute p-values for detecting systematic up or downregulation of the specified gene set in the specified category.

**Usage**

```
linearMTest(p)
```

**Arguments**

p	An instance of a subclass of LinearMParams. This parameter object determines the category of interest (currently, only chromosome bands) as well as the gene set.
---	---

**Details**

The per-gene statistics in the geneStats slot of p give a measure of up or down regulation of the individual genes in the universe.

**Value**

A LinearMResult instance.

**Author(s)**

D. Sarkar

**See Also**

[LinearMResult-class](#) [LinearMParams-class](#)

---

`makeChrBandGraph`*Create a graph representing chromosome band annotation data*

---

### Description

This function returns a graph object representing the nested structure of chromosome bands (also known as cytogenetic bands). The nodes of the graph are band identifiers. Each node has a `geneIds` node attribute that lists the gene IDs that are annotated at the band (the gene IDs will be Entrez IDs in most cases).

### Usage

```
makeChrBandGraph(chip, univ = NULL)
```

### Arguments

<code>chip</code>	A string giving the annotation source. For example, "hgu133plus2"
<code>univ</code>	A vector of gene IDs (these should be Entrez IDs for most annotation sources). The annotations attached to the graph will be limited to those specified by <code>univ</code> . If <code>univ</code> is <code>NULL</code> (default), then the gene IDs are those found in the annotation data source.

### Details

This function parses the data stored in the `<chip>MAP` map from the appropriate annotation data package. Although cytogenetic bands are observed in all organisms, currently, only human and mouse band nomenclatures are supported.

### Value

A [graph-class](#) instance. The graph will be a tree and the root node is labeled for the organism.

### Author(s)

Seth Falcon

### Examples

```
chrGraph <- makeChrBandGraph("hgu95av2.db")
chrGraph
```

---

makeEBcontr*A function to make the contrast vectors needed for EBarrays*

---

## Description

Using EBarrays to detect differential expression requires the construction of a set of contrasts. This little helper function computes these contrasts for a two level factor.

## Usage

```
makeEBcontr(f1, hival)
```

## Arguments

f1	The factor that will define the contrasts.
hival	The level of the factor to treat as the high level.

## Details

Not much more to add, see EBarrays for more details. This is used in the Category package to let users compute the posterior probability of differential expression, and hence to compute expected numbers of differentially expressed genes, per category.

## Value

An object of class “ebarraysPatterns”.

## Author(s)

R. Gentleman

## See Also

[ebPatterns](#)

## Examples

```
if( require("EBarrays") ) {  
  myfac = factor(rep(c("A", "B"), c(12, 24)))  
  makeEBcontr(myfac, "B")  
}
```

---

makeValidParams

*Non-standard Generic for Checking Validity of Parameter Objects*


---

### Description

This function is not intended for end-users, but may be useful for developers extending the Hypergeometric testing capabilities provided by the Category package.

makeValidParams is intended to validate a parameter object instance (e.g. HyperGParams or subclass). The idea is that unlike validObject, methods for this generic attempt to fix invalid instances when possible, and in this case issuing a warning, and only give an error if the object cannot be fixed.

### Usage

```
makeValidParams(object)
```

### Arguments

object                      A parameter object. Consult showMethods to see signatures currently supported.

### Value

The value must have the same class as the object argument.

### Author(s)

Seth Falcon

---

MAPAmat

*Mapping chromosome bands to genes*


---

### Description

These functions return a mapping of chromosome bands to genes. makeChrBandGSC returns a [GeneSetCollection](#) object, with a GeneSet for each band. The other functions return a 0/1 incidence matrix with a row for each chromosome band and a column for each gene. Only those chromosome bands with at least one gene annotation will be included.

### Usage

```
MAPAmat(chip, univ = NULL, minCount = 0)
makeChrBandInciMat(chrGraph)
makeChrBandGSC(chrGraph)
```

**Arguments**

chip	A string giving the annotation source. For example, "hgu133plus2"
univ	A vector of gene IDs (these should be Entrez IDs for most annotation sources). The the annotations will be limited to those in the set specified by univ. If univ is NULL (default), then the gene IDs are those found in the annotation data source.
chrGraph	A graph object as returned by makeChrBandGraph
minCount	Bands with less than minCount genes will be excluded from the returned matrix. If minCount is 0, no bands will be removed, this is the default.

**Value**

For makeChrBandGSC, a GeneSetCollection object with a GeneSet for each band.

For the other functions, (0/1) incidence matrix with chromosome bands as rows and gene IDs as columns. A 1 in  $m[i, j]$  indicates that the chromosome band `rownames(m)[i]` contains the geneID `colnames(m)[j]`.

**Author(s)**

Seth Falcon, Michael Lawrence

**See Also**

[makeChrBandGraph](#), [cateG0ry](#), [probes2MAP](#)

**Examples**

```
have_hgu95av2.db <- suppressWarnings(require("hgu95av2.db"))
if (have_hgu95av2.db)
  mam <- MAPAmat("hgu95av2.db")
```

---

NewChrBandTree	<i>Create a new ChrBandTree object</i>
----------------	--

---

**Description**

NewChrBandTree and ChrBandTreeFromGraph provide constructors for the ChrBandTree class.

**Usage**

```
NewChrBandTree(chip, univ)
ChrBandTreeFromGraph(g)
```

**Arguments**

chip	The name of an annotation data package
univ	A vector of gene identifiers that defines the universe of genes. Usually, this will be a vector of Entez Gene IDs. If univ is NULL, then all genes probed on the specified chip will be in the universe. We strongly recommend using the set of genes that remains after applying a non-specific filter as the universe.
g	A graph instance as returned by makeChrBandGraph

**Value**

A new ChrBandTree instance.

**Author(s)**

S. Falcon

**See Also**

[ChrBandTree-class](#)

---

probes2MAP

*Map probe IDs to MAP regions.*

---

**Description**

This function maps probe identifiers to MAP positions using the appropriate Bioconductor meta-data package.

**Usage**

```
probes2MAP(pids, data = "hgu133plus2")
```

**Arguments**

pids	A vector of probe IDs for the chip in use.
data	The name of the chip, as a character string.

**Details**

Probes are mapped to regions, no checking for duplicate Entrez gene IDs is done.

**Value**

A vector, the same length as pids, with the MAP locations.

**Author(s)**

R. Gentleman

**See Also**[probes2Path](#)**Examples**

```
set.seed(123)
library("hgu95av2.db")
v1 = sample(names(as.list(hgu95av2MAP)), 100)
pp = probes2MAP(v1, "hgu95av2.db")
```

---

probes2Path*A function to map probe identifiers to pathways.*

---

**Description**

Given a set of probe identifiers from a microarray this function looks up all KEGG pathways that the probe is documented to be involved in.

**Usage**

```
probes2Path(pids, data = "hgu133plus2")
```

**Arguments**

pids	A vector of probe identifiers.
data	The character name of the chip.

**Details**

This is a simple look up in the appropriate chip PATH data environment.

**Value**

A list of pathway vectors. One element for each value of pid that is mapped to at least one pathway.

**Author(s)**

R. Gentleman

**See Also**[findAMstats](#)**Examples**

```
library("hgu95av2.db")
x = c("1001_at", "1000_at")
probes2Path(x, "hgu95av2.db")
```

---

`tperm`*A simple function to compute a permutation t-test.*

---

**Description**

The data matrix, `x`, with two-level factor, `fac`, is used to compute t-tests. The values of `fac` are permuted `B` times and the complete set of t-tests is performed for each permutation.

**Usage**

```
tperm(x, fac, B = 100, ts0 = TRUE)
```

**Arguments**

<code>x</code>	A data matrix. The number of columns should be the same as the length of <code>fac</code> .
<code>fac</code>	A factor with two levels.
<code>B</code>	An integer specifying the number of permutations.
<code>ts0</code>	A logical indicating whether to compute only the t-test statistic for each permutation. If <code>FALSE</code> then p-values are also computed - but this can be very slow.

**Details**

Not much more to say. Probably there is a generic function somewhere, but I could not find it.

**Value**

A list, the first element is named `obs` and contains the true, observed, values of the t-statistic. The second element is named `ans` and contains a list of length `B` containing the different permutations.

**Author(s)**

R. Gentleman

**See Also**

[rowttests](#)

**Examples**

```
x=matrix(rnorm(100), nc=10)
y = factor(rep(c("A","B"), c(5,5)))
tperm(x, y, 10)
```

---

universeBuilder	<i>Return a vector of gene identifiers with category annotations</i>
-----------------	--

---

### Description

Return all gene ids that are annotated at one or more terms in the category. If the universeGeneIds slot of p has length greater than zero, then the intersection of the gene ids specified in that slot and the normal return value is given.

### Usage

```
universeBuilder(p)
```

### Arguments

p	A subclass of HyperGParams-class
---	----------------------------------

### Details

End users **should not** call this directly. This method gets called from hyperGTest. To add support for a new category, a new method for this generic must be defined. Its signature should match a subclass of HyperGParams-class appropriate for the new category.

### Value

A vector of gene identifiers.

### Author(s)

S. Falcon

### See Also

[hyperGTest](#) [HyperGParams-class](#)

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