

# Package ‘cqn’

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**Version** 1.2.0

**Title** Conditional quantile normalization

**Description** A normalization tool for RNA-Seq data, implementing the conditional quantile normalization method.

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**Depends** R (>= 2.10.0), mclust, nor1mix, stats, preprocessCore, splines, quantreg

**Imports** splines

**Suggests** scales, edgeR

**License** Artistic-2.0

**LazyLoad** yes

**biocViews** RNAseq, Preprocessing, DifferentialExpression

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cqn	<i>CQN (conditional quantile normalization) for RNA-Seq data</i>
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## Description

This function implements CQN (conditional quantile normalization) for RNA-Seq data.

## Usage

```
cqn(counts, x, lengths, sizeFactors = NULL, subindex = NULL, tau = 0.5, sqn = TRUE, verbose = FALSE)
cqn.fixedlength(counts, x, lengths, sizeFactors = NULL, subindex = NULL, tau = 0.5, sqn = TRUE, verbose = FALSE)
```

**Arguments**

counts	An object that can be coerced to a matrix of region by sample counts. Ought to have integer values.
x	This is a covariate whose systematic influence on the counts will be removed. Typically the GC content. Has to have the same length as the number of rows of counts.
lengths	The lengths (in bp) of the regions in counts. Has to have the same length as the number of rows of counts.
sizeFactors	An optional vector of sizeFactors, ie. the sequencing effort of the various samples. If NULL this is calculated as the column sums of counts.
subindex	An optional vector of indices into the rows of counts. If not given, this becomes the indices of genes with row means of counts greater than 50.
tau	This argument is passed to rq, it indicates what quantile is being fit. The default should only be changed by expert users..
sqn	This argument indicates whether the residuals from the systematic fit are (subset) quantile normalized. The default should only be changed by expert users.
verbose	Is the function verbose?

**Details**

These functions implement the CQN (conditional quantile normalization) for RNA-Seq data. The functions remove a single systematic effect, contained in the argument `x`, which will typically be GC content. The effect of `lengths` will either be modelled as a smooth function (which we recommend), if you are using `cqn` or as an offset (equivalent to modelling using RPKMs), if you are using `cqn.fixedlength`.

Final corrected values are equal to `value$y + value$offset`.

**Value**

A list with the following components

counts	The value of argument counts.
x	The value of argument x.
lengths	The value of argument lengths.
sizeFactors	The value of argument sizeFactors. In case the argument was NULL, this is the value used internally.
subindex	The value of argument subindex. In case the argument was NULL, this is the value used internally.
y	The dependent value used in the systematic effect fit. Equal to log <sub>2</sub> transformed reads per millions.
offset	The estimated offset.
offset0	A single number used internally for identifiability.
func1	The estimated effect of function 1 (argument x). This is a matrix of function values on a grid. Columns are samples and rows are grid points.
grid1	The grid points on which function 1 (argument x) was evaluated.
knots1	The knots used for function 1 (argument x).

func2	The estimated effect of function 2 (lengths). This is a matrix of function values on a grid. Columns are samples and rows are grid points.
grid2	The grid points on which function 2 (lengths) was evaluated.
knots2	The knots used for function 2 (lengths).

**Note**

Internally, the function uses a custom implementation of subset quantile normalization, contained in the (not exported) SQN2 function.

**Author(s)**

Kasper Daniel Hansen, Zhijin Wu

**References**

Hansen, K.D., Irizarry, R.A. and Wu Z., Removing technical variability in RNA-seq data using conditional quantile normalization, Johns Hopkins, Dept of Biostatistics Working Papers. Working Paper 227, <http://www.bepress.com/jhubiostat/paper227>

**See Also**

The package vignette.

**Examples**

```
data(montgomery.subset)
data(sizeFactors.subset)
data(uCovar)
cqn.subset <- cqn(montgomery.subset, lengths = uCovar$length,
                 x = uCovar$gccontent, sizeFactors = sizeFactors.subset,
                 verbose = TRUE)
```

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cqnplot

*Plot the systematic effect estimated as part of a CQN normalization.~*

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**Description**

This function plots the estimated systematic effect which are removed during CQN normalization.

**Usage**

```
cqnplot(x, n = 1, col = "grey60", ylab = "QR fit", xlab = "", type = "l", lty = 1, ...)
```

**Arguments**

x	The result of a call to cqn; an object of class cqn.
n	Which systematic effect is plotted.
col	A vector of colors, as in plot.
ylab	y-label as in plot.
xlab	x-label as in plot.

type            type, as in plot.  
 lty            line type, as in plot.  
 ...            These arguments are passed to `matplot`

### Value

This function is invoked for its side effect.

### Author(s)

Kasper Daniel Hansen

### Examples

```

data(montgomery.subset)
data(sizeFactors.subset)
data(uCovar)
cqn.subset <- cqn(montgomery.subset, lengths = uCovar$length,
                 x = uCovar$gccontent, sizeFactors = sizeFactors.subset,
                 verbose = TRUE)
cqnplot(cqn.subset, n = 1)

```

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montgomery.subset      *Montgomery RNA-seq data.*

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### Description

A gene by sample count matrix for 10 samples from from Montgomery et al. Also included is information about these genes (length and gc content) as well as sequencing depth for each of the samples.

### Usage

```

data(montgomery.subset)
data(sizeFactors.subset)
data(uCovar)

```

### Format

`montgomery.subset` is a data frame with 23552 observations on 10 different samples, the column names are the sample ids. `sizeFactors.subset` a a named vector of length 10 containing the number of mapped reads for each of the 10 samples. `uCovar` is a data frame with 23552 observations on 2 different covariates: gc content and genic length in bp.

### Details

Gene models are union models based on Ensembl 61. These gene models were constructed using Genominator. Genes that have zero counts in all 10 samples were excluded.

### References

SB Montgomery, M Sammeth, M Gutierrez-Arcelus, RP Lach, C Ingle, J Nisbett, R Guigo, ET Dermitzakis, (2010) "Transcriptome genetics using second generation sequencing in a Caucasian population". *Nature* 464(7289), 773-777.

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